

Usher syndrome type 1C: Mechanisms, Animal Models and the hunt for a Cure

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Usher Coalition
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1. Lentz Lab Mission
2. Usher syndrome type 1C
3. Acadian Usher syndrome
4. *USH1C* and Harmonin
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7. Future directions

Lentz Lab
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Mission

**To develop a therapeutic approach to
prevent or cure the deafness and
blindness associated with Usher syndrome**

Focus: Usher syndrome type 1C

Congenital Deafness – born with severe to profound hearing impairment

Vestibular Areflexia – difficulty with balance

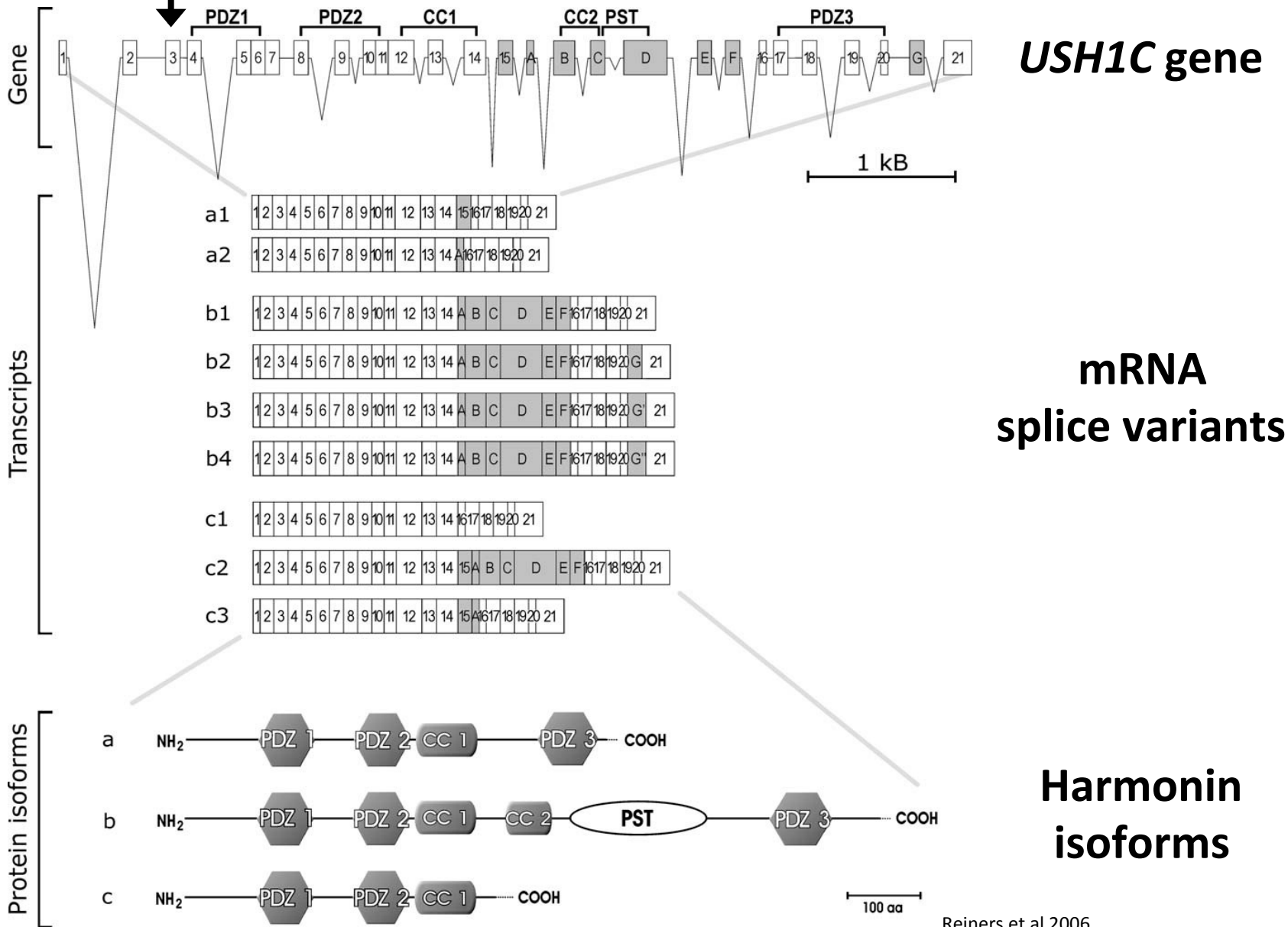
Retinitis Pigmentosa – begins in early adolescence with night-blindness

6-8% of Usher 1 cases are caused by mutations in the *USH1C* gene, which encodes the protein harmonin

All cases of Usher 1 in Acadian populations (South Louisiana and Canada) are caused by the *USH1C.216G>A* mutation (216A)

USH1C and Harmonin Protein Isoforms

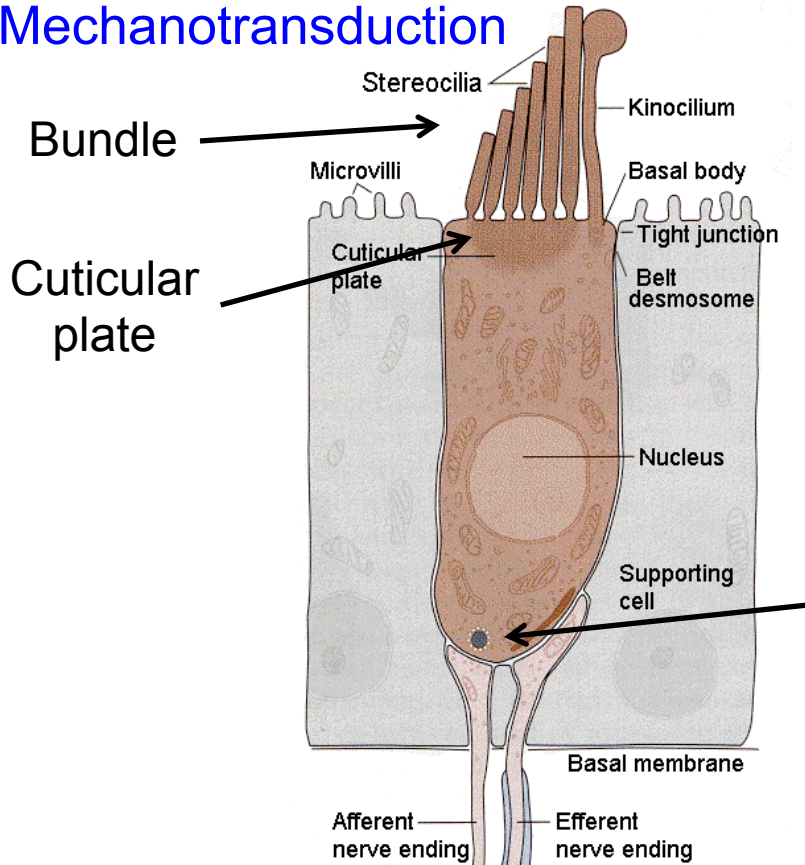
216G>A mutation



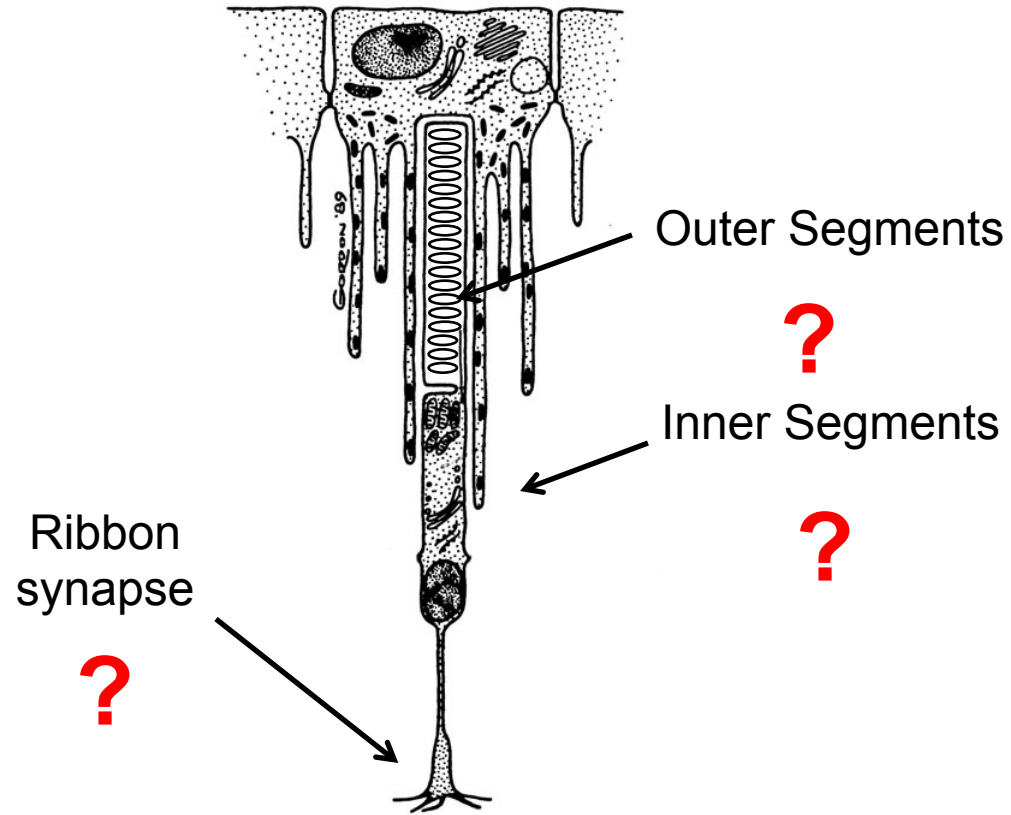
Harmonin

Cochlear Hair Cell

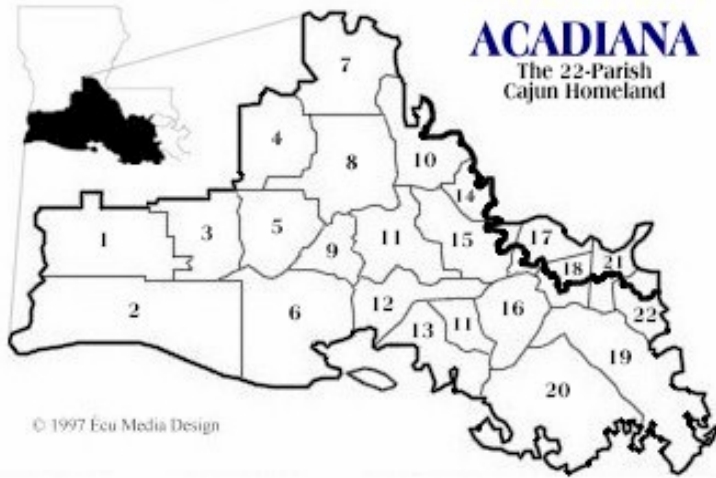
Bundle morphogenesis
Mechanotransduction



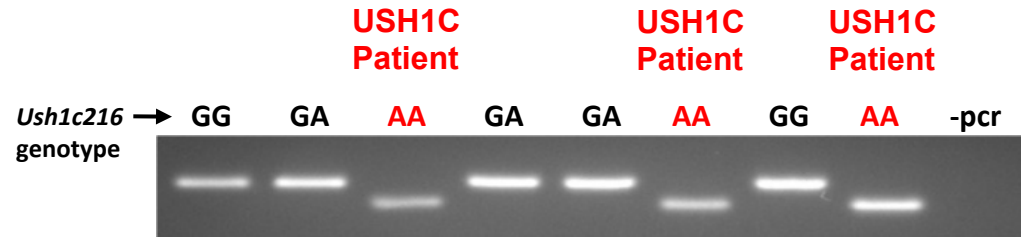
ROD photoreceptor



Acadian Usher Syndrome type 1C



Gene Expression in USH1C Patients



**Truncated mRNA
with 35 bp deletion
at the end of exon 3**

Predicted to encode a truncated harmonin

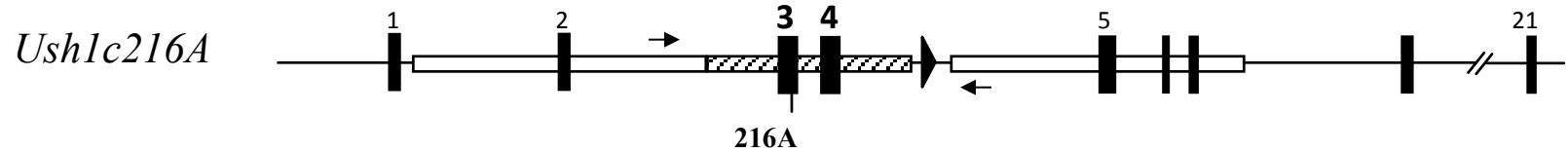
Wild-type Harmonin

MDRKVAREFRHKVDFLIENDAEDKDYLDVLRMYHQTMDDAVLVGDLKLVINEPSRPLPFDIAIRPLIPL
KHQVEYDQLTPRRSRKLKEVRLDRLHPEGLGLSVRGGLFEGCGLETSHLTKGGQADSVGLQVGDPIV
INGYSISSCTHEEVINLIRTKKTVSIVKVRHIGLIPVKS SPDEPLTWQYVDQFVSES GGVRGSLGSPGN
RENKEKKVFISLVGSRGLGCSISSGPIOKPGIFTSHVKPGSLSAEVGLEFIGDOIVEVNGVDFESNLDHK
EAVNVLKNSRSLTISIVAAAGRELFMTDRERLAEARQRELQRQELLMQKRLAMESNKILQEQQEMERQ
RRKEIAQKAAEENERYRKEMEQIVVEEEKFKKQWEEDWGSKEQLLLPKTITAENVHPVPLRKPYPYDQGV
EPELEPADDLDGGTTEEQGEQDFRKYEEGFDPYSMFTPEQIMGKDVRLRLIKKEGSLDLALEGGVDSPI
GKVVVSAVYERGAERHGGIVKGDFTMATNGKIVTDYTLAEADAALQKAWNQQGDWIDLVVAVCPPKE
YDDELTFLLKSKRGNQIHALGNSELRPHLVNTKPRTSLEGRHMTHTRWHPWDLNLSPRNLKPLALNQ
GQIRNSSGHFFEGQCGGKAASRLGEDLKDPPDSHSFPLAQ

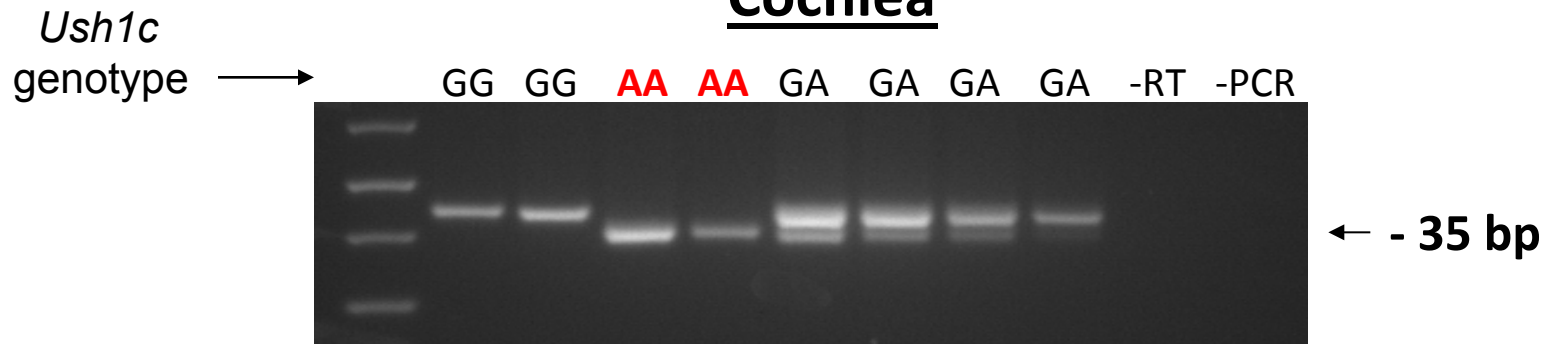
Truncated 216A mRNA Translation

MDRKVAREFRHKVDFLIENDAEDKDYLDVLRMYHQTMDDAVLVGDLKLVINEPSRPLPFDIAIRPLIPL
KHQEAEGGASGPSAPRRRPECAWNPVWLVWALHLPHPHQRSSGRQRRAPEGRRDRPDQWIFHLLLYP

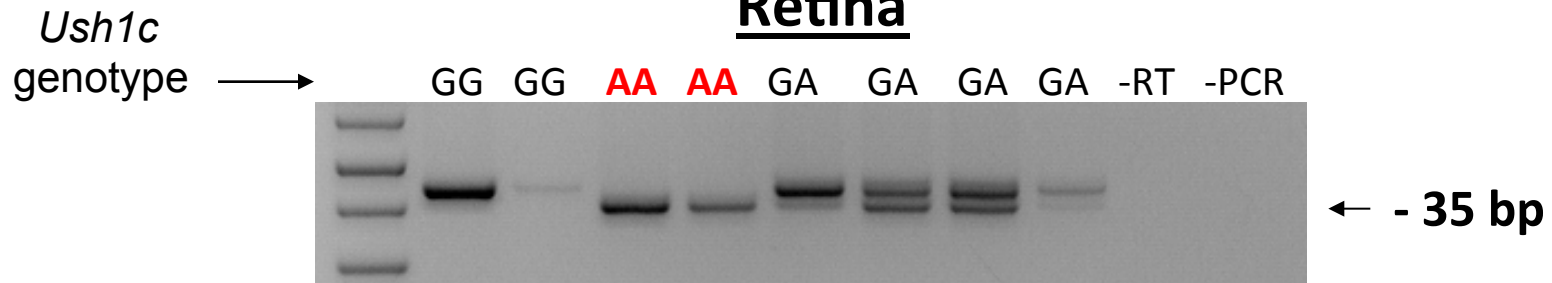
Put the Human 216A mutation into the Mouse *Ush1c* gene (knock-in)



Cochlea



Retina

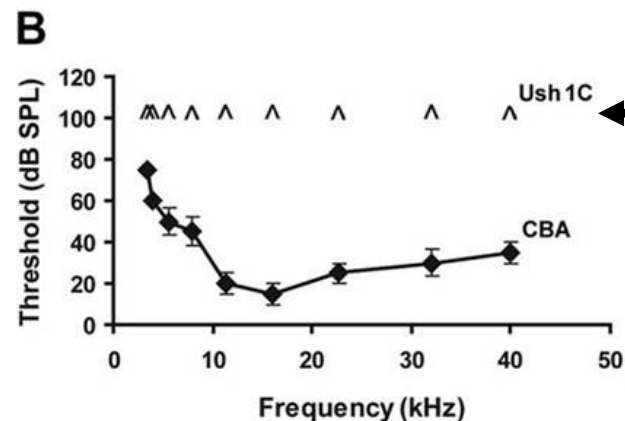
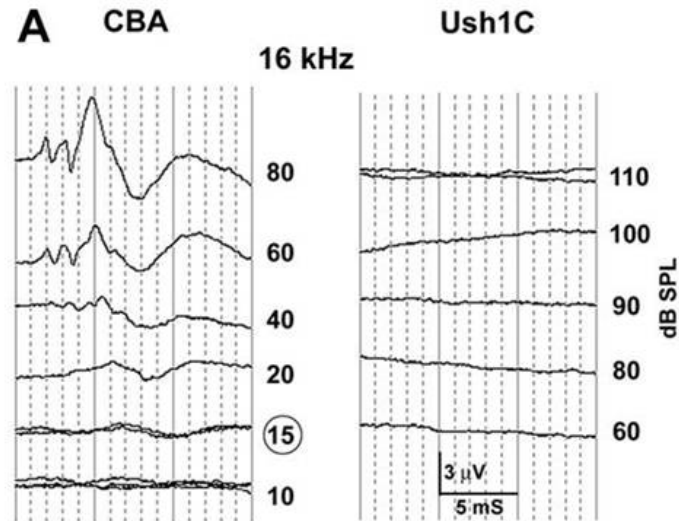


216AA mice express truncated mRNA in cochlea and retina

216AA mice have vestibular dysfunction and are deaf

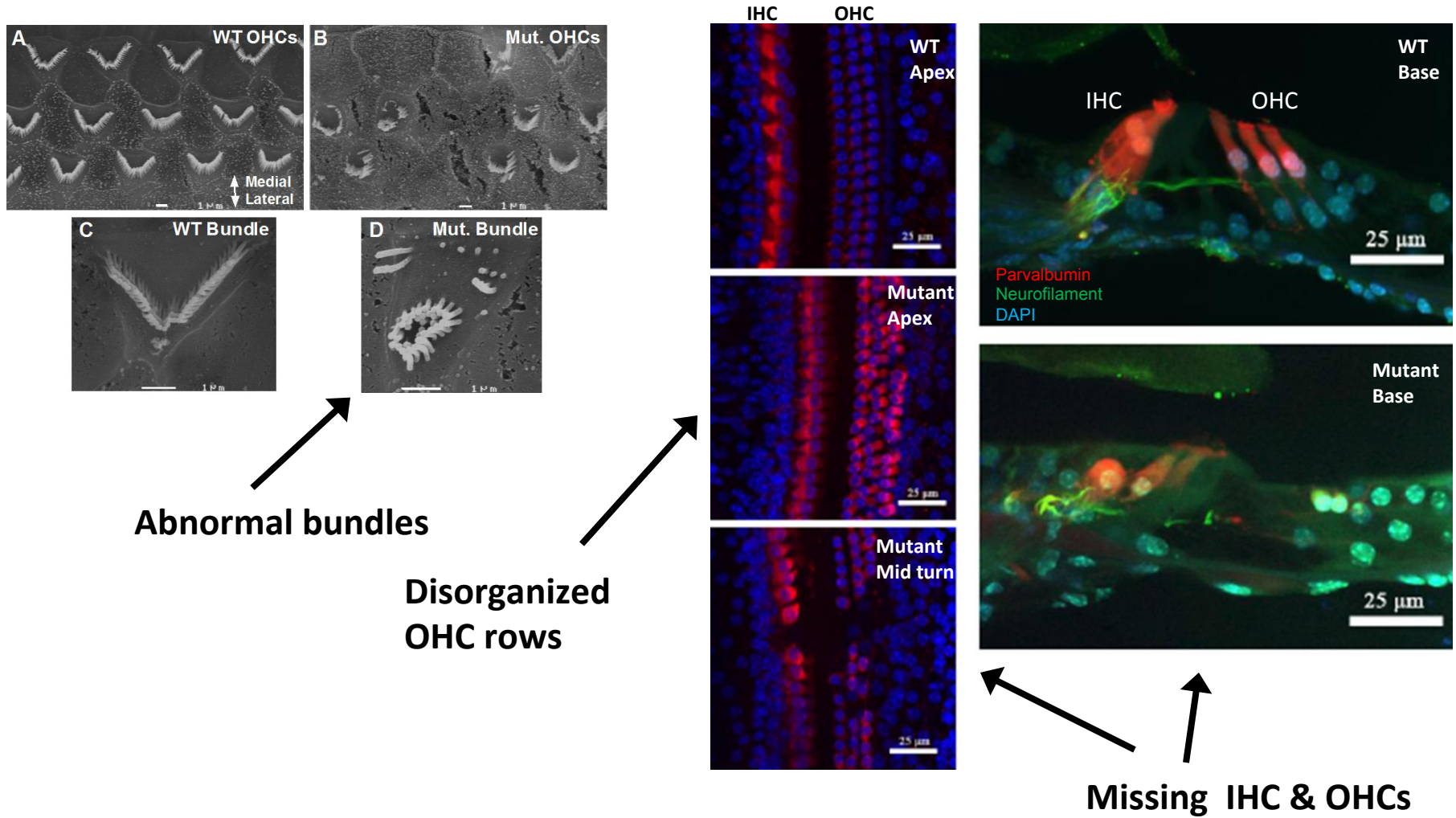
No Auditory-evoked Brainstem Response (ABR)

Circling and head tossing behavior



← No ABR at high, mid or low frequencies

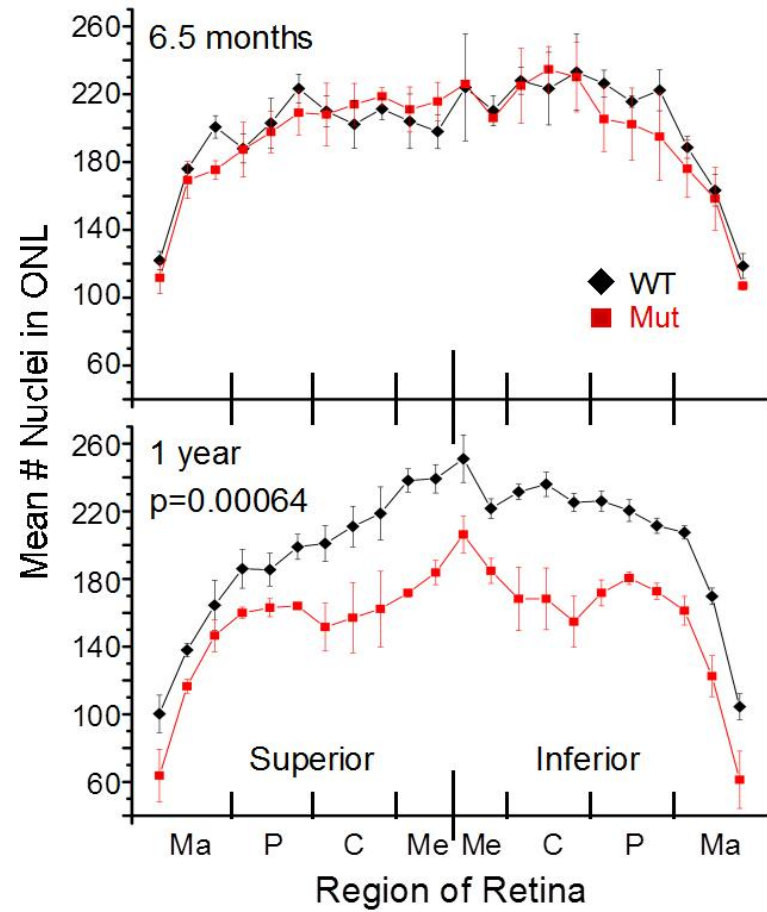
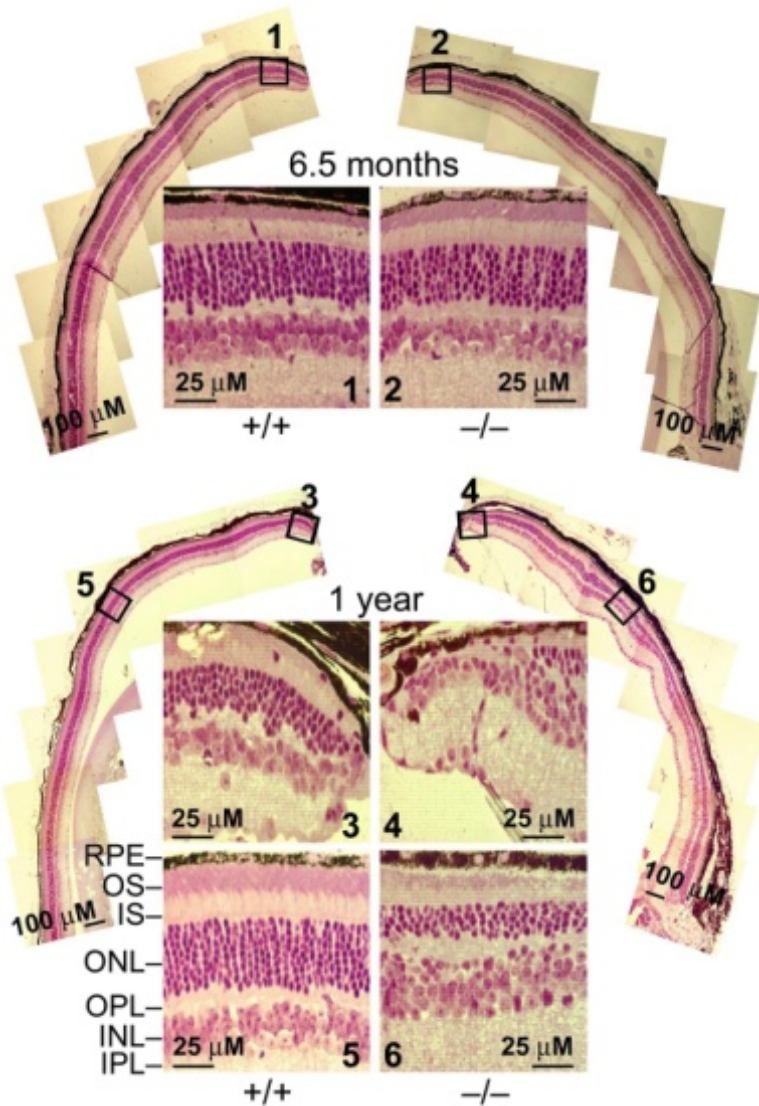
216AA mice have abnormal bundles and missing hair cells



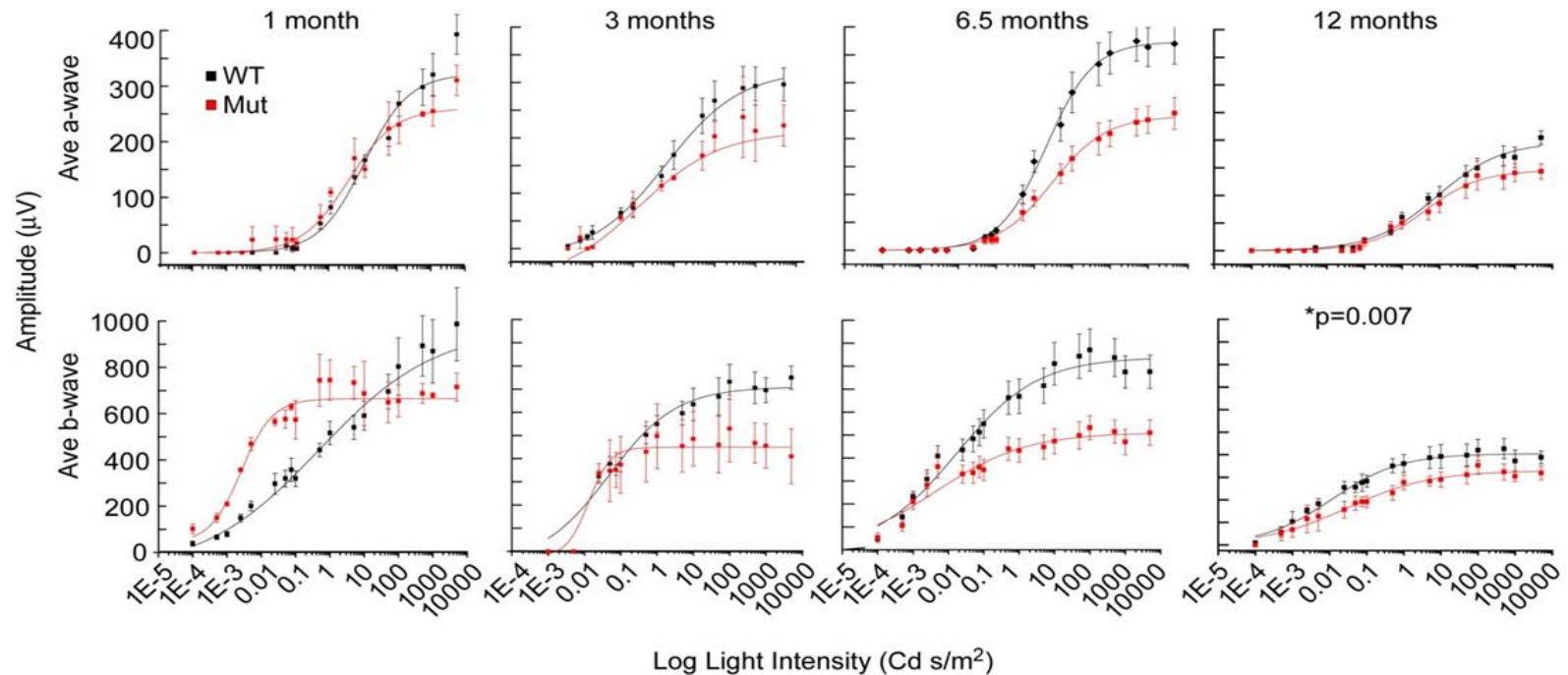
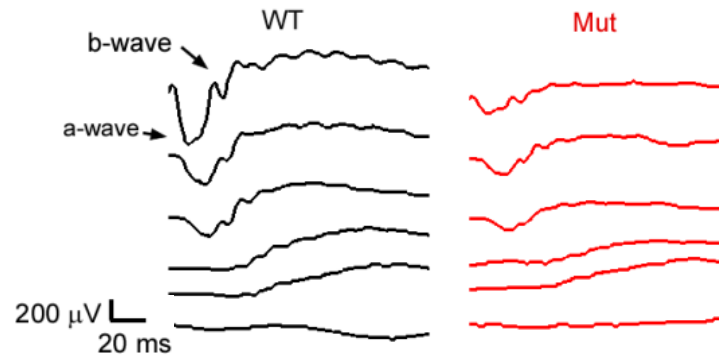
216AA mice have progressive photoreceptor degeneration

Wild Type

216AA



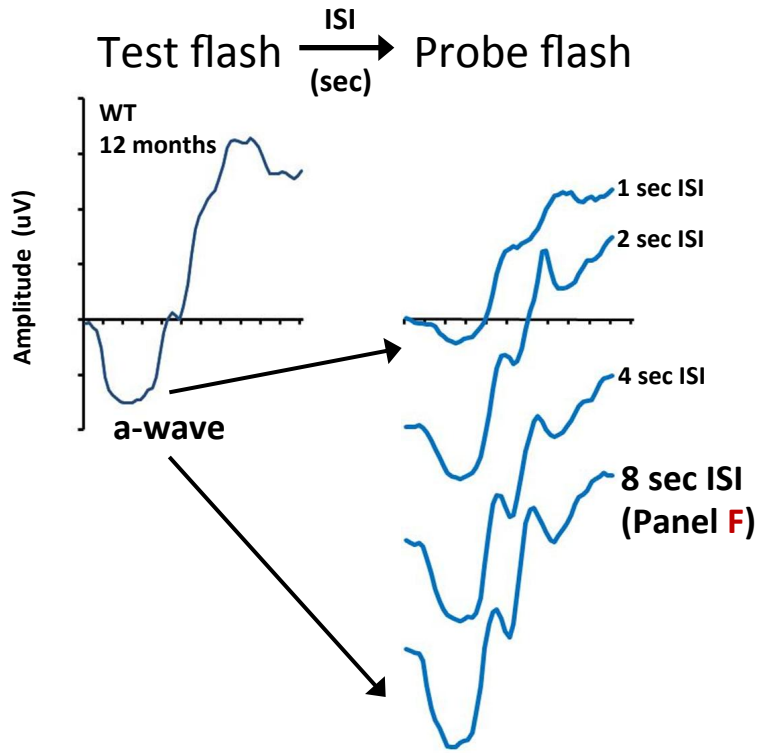
216AA mice have reduced visual function and retinal degeneration



216AA mice have slow rod adaptation

Twin flash protocol

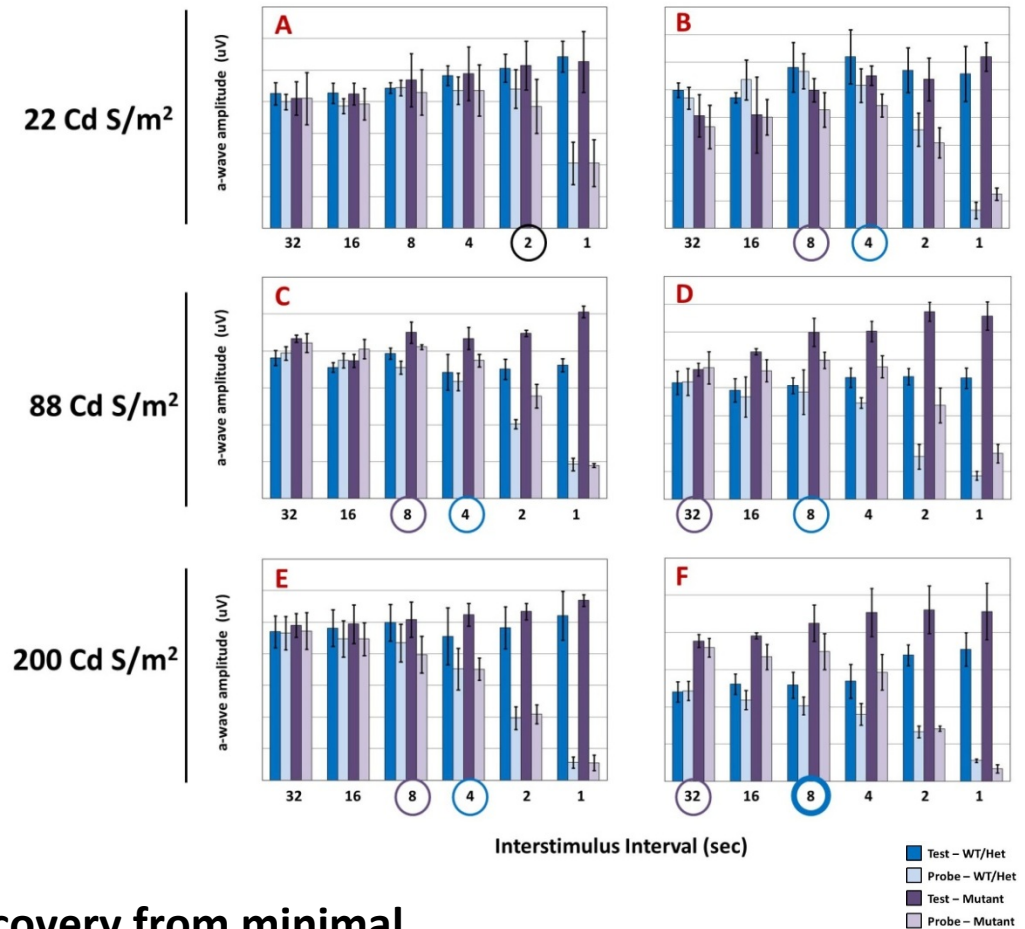
Minimal Bleach (> 1% rhodopsin)



Deactivation of Rhodopsin Kinetics

6 months old

12 months old

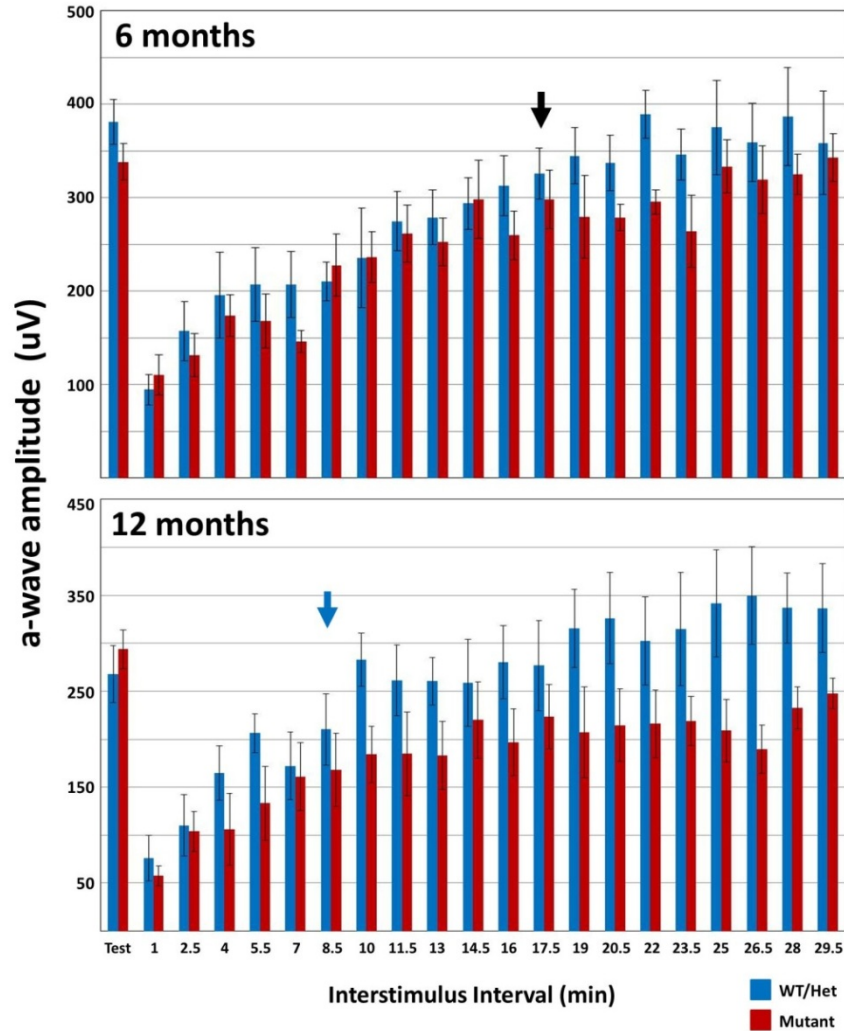
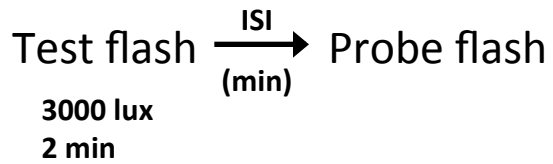


a-wave recovery from minimal bleach takes 2 – 4 times as long

Visual Cycle Kinetics

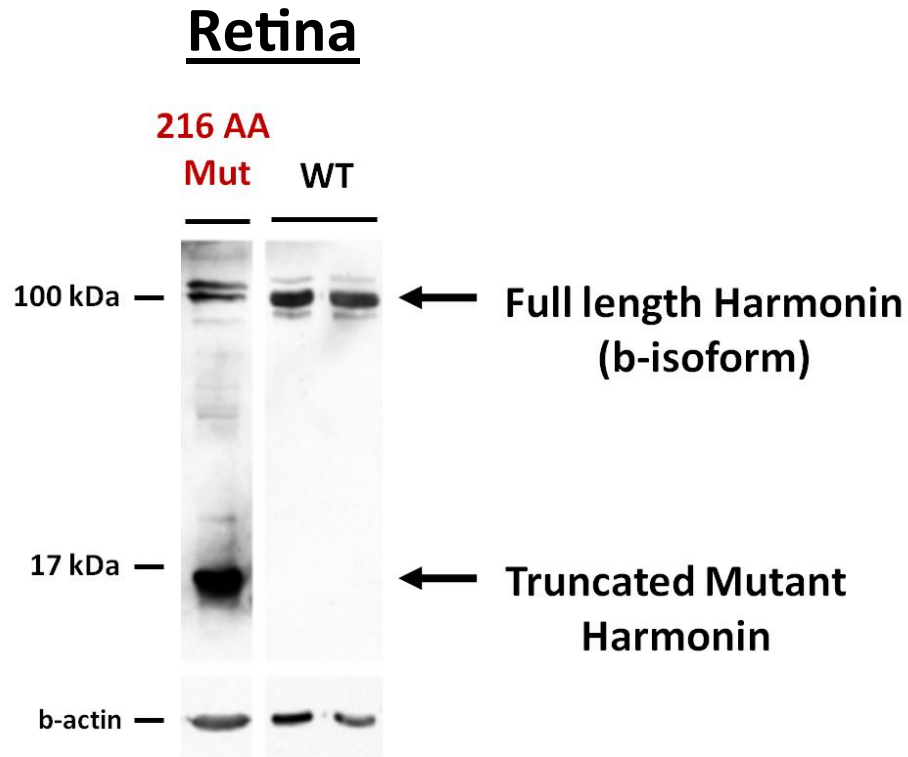
Paired flash protocol

Bleach 30-40 % Rhodopsin



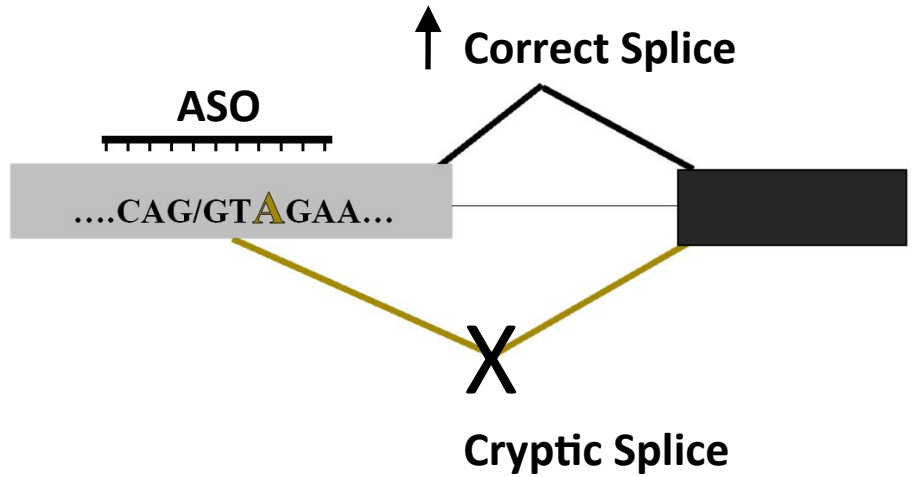
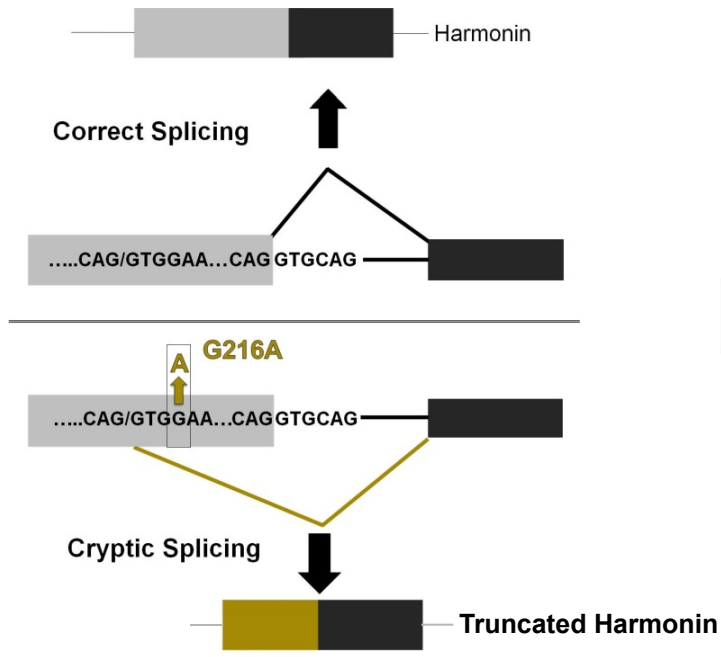
Mutant a-wave not recovered after 30 min

216AA mice express both wt and mutant harmonin



Can we detect both wt and mutant Ush1c mRNA?

Can we modulate the use of the mutant and wild type splice sites by treating with antisense oligonucleotides (ASOs)?



ASO targeted to 216 mutation prevents cryptic splicing and forces correct splicing

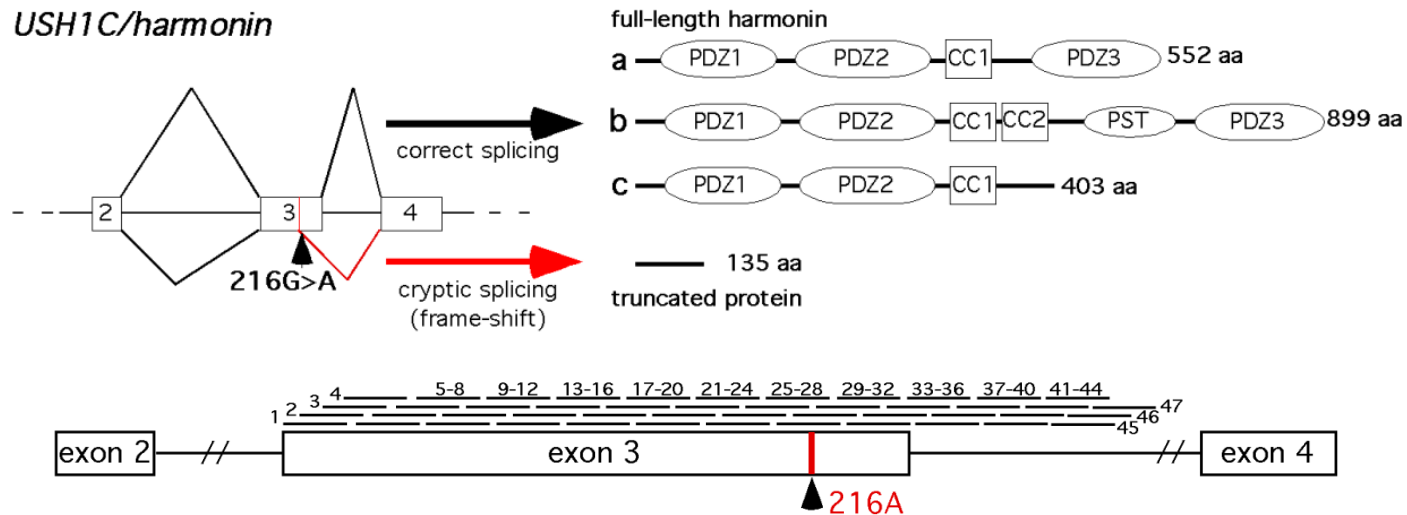
Antisense Oligonucleotides (ASOs)

ASOs are small molecules that are

- designed to have high affinity for their RNA target through unique sequence base pairing
- resistant to degradation, which allows for high potency and specificity, and low toxicity

ASO-based therapeutics have been FDA approved for other conditions and toxicity studies have proven the ASO chemistry is not toxicity for humans

ASO design & optimization



1. Tested 50 different ASOs in cell lines transfected with 216A minigenes
Selected an ASO that showed the largest increase in correct splicing and the largest decrease in cryptic splicing
2. Tested that ASO in patient cell lines and cells from 216AA mice
Ush-ASO corrected cryptic splicing and increased correct splicing
3. Tested the Ush-ASO in Adult mice
Ush-ASO corrected cryptic splicing and increased correct splicing in a dose dependent manor

Treatment Model with ASOs



Injection Time
(Postnatal Day)

P3-5, P10 or P16-18



Behavior

Vestibular function (Open-field chamber)

Physiology

Hearing function (ABRs, preyer reflex)

Histology

Hair cell morphology (Immunohistochemistry)

Molecular

Ush1c and Harmonin Expression

Treatment of 216AA mice with Ush-ASO

Vestibular Function-

Single treatment between postnatal day 5 (P5) – P13 rescues vestibular function

Hearing function-

Single treatment at P5 restores hearing at low – mid frequencies similar to wild type levels

Histology-

Single treatment at P5 rescues harmonin expression in the hair cell bundle and at the synapse; partially restores hair bundle structure; and rescues hair cells at the apex and middle turn

Molecular-

Single treatment at P5 decreases cryptic splicing and increases wt expression of *Ush1c* and harmonin in the ear

Conclusions

ASOs targeted to the 216A mutation rescue gene and protein expression in patient and mouse cell lines

A single systemic injection in 216AA mice-

Cures vestibular dysfunction

Rescues hearing at low and mid frequencies

Partially restores hair bundle morphology and decreases hair cell loss

Partially rescues *Ush1c* and harmonin expression in the ear

Future Directions

Further develop an ASO treatment regiment in our mouse model to prevent deafness and vestibular defects with the goal of providing pre-clinical data that would lay the framework for clinical trials in patients

Test the ASO for the treatment of blindness in our mouse model

Continue studies in the ear and eye to understand how the 216A mutation causes deafness and blindness

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