



Using Zebrafish to Develop the First Pharmacotherapy for the Treatment of Hearing Loss Caused by Usher Syndrome Type I due to Variants in MYO7A

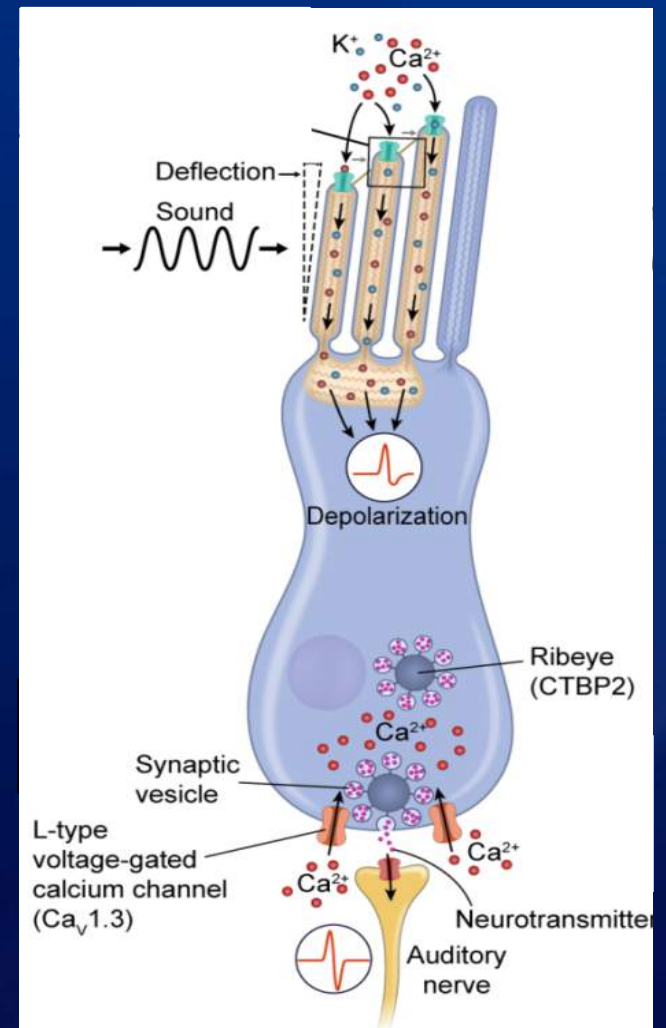
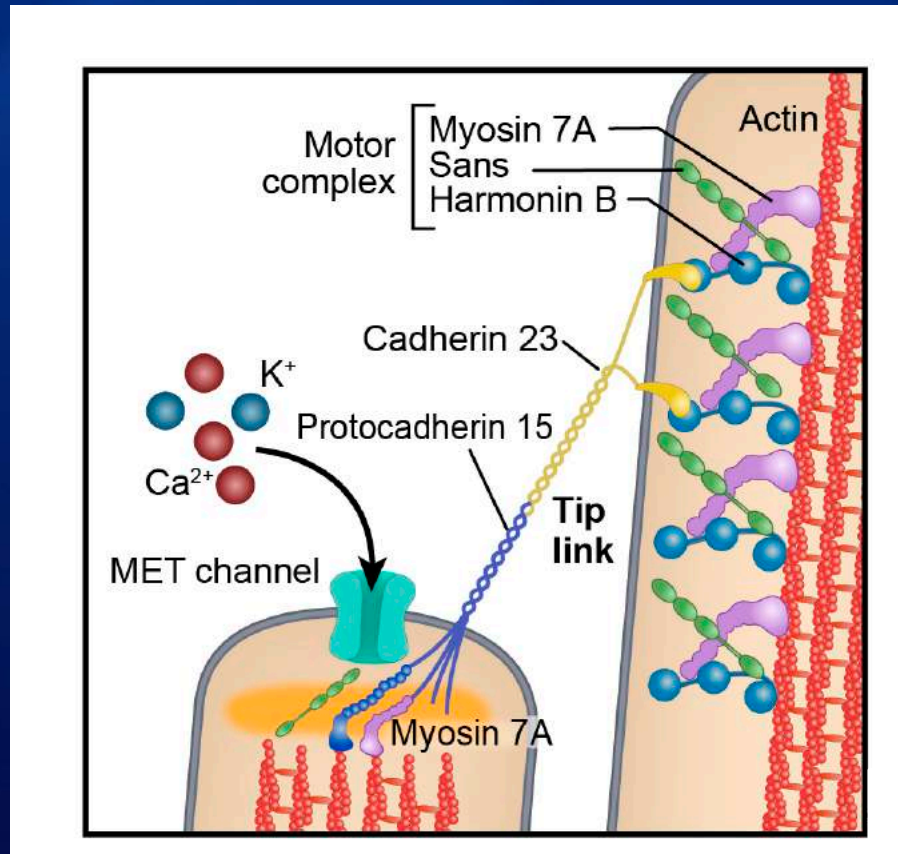
Alaa Koleilat, Ph.D.

USH Connections Week 2020

Usher Syndrome

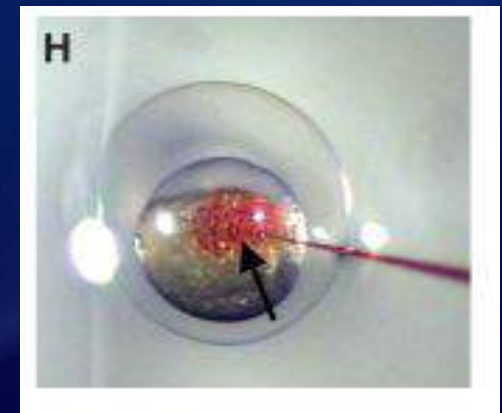
Type	Hearing Impairment	Onset of Hearing Loss	Vestibular Impairment	Vision Loss	Genes
I	Severe to profound hearing loss	At birth	Severe (E.g. walk at a later age, etc.)	Onset in the first decade of life	<i>MYO7A</i> <i>USH1C</i> <i>CDH23</i> <i>PCDH15</i> <i>SANS/USH1G</i>
II	Moderate to severe hearing loss	At birth	none	Onset in first to second decade of life	<i>USH2A</i> <i>VLGR1</i> <i>WHRN</i>
III	Variable, progressive hearing loss	Adolescence	Variable	Variable	<i>CLRN1</i> <i>HARS</i>

MYO7A and the inner ear hair cell



Zebrafish as Model Organism to Study Human Disease

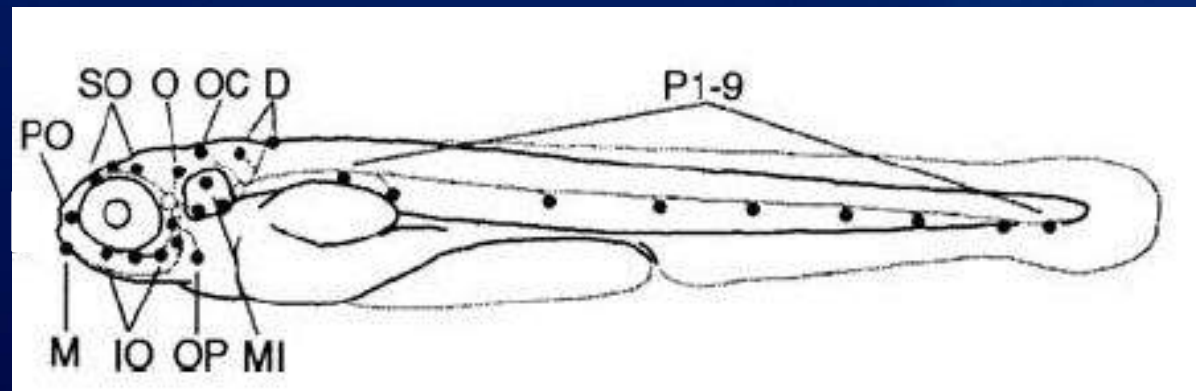
- Availability of a zebrafish genome
- Ease of gene editing to model human diseases
- Produce hundreds of embryos in one day
- Ease of administering drugs



Bill et al. 2009 *Zebrafish*

Zebrafish as Model Organism to Study Genetic Hearing Loss

- Acoustic startle response present at 5 days post fertilization
- External sensory organ → lateral line



Harris et al. 2003 *JARO*

Genetic Analysis of Vertebrate Sensory Hair Cell Mechanosensation: the Zebrafish Circler Mutants

Teresa Nicolson,*# Alfons Rüschi,‡
Rainer W. Friedrich,† Michael Granato,*¶
Johann Peter Ruppertsberg,‡§
and Christiane Nüsslein-Volhard*

Table 1. Mutations Affecting Larval Vestibular Function

Gene	Abbreviation
<i>sputnik</i>	<i>spu</i>
<i>mariner</i>	<i>mar</i>
<i>orbiter</i>	<i>orb</i>
<i>mercury</i>	<i>mrc</i>
<i>gemini</i>	<i>gem</i>
<i>skylab</i>	<i>skb</i>
<i>astronaut</i>	<i>asn</i>
<i>cosmonaut</i>	<i>csm</i>

Genetic Analysis of Vertebrate Sensory Hair Cell Mechanosensation: the Zebrafish Circler Mutants

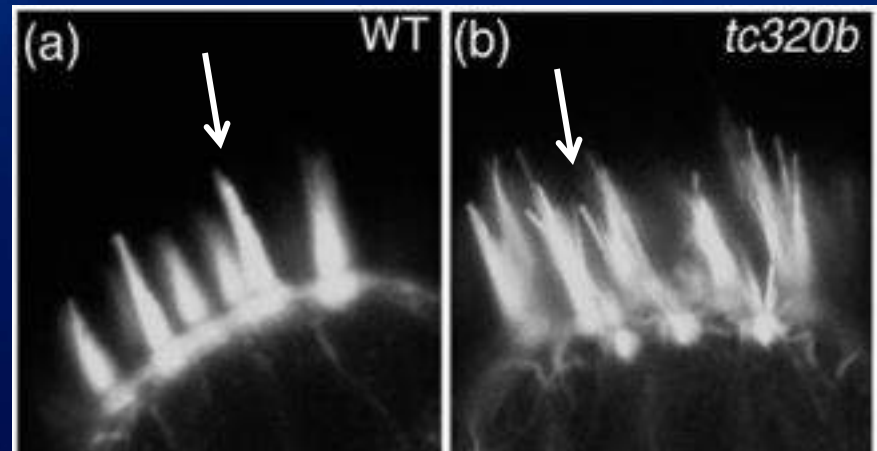
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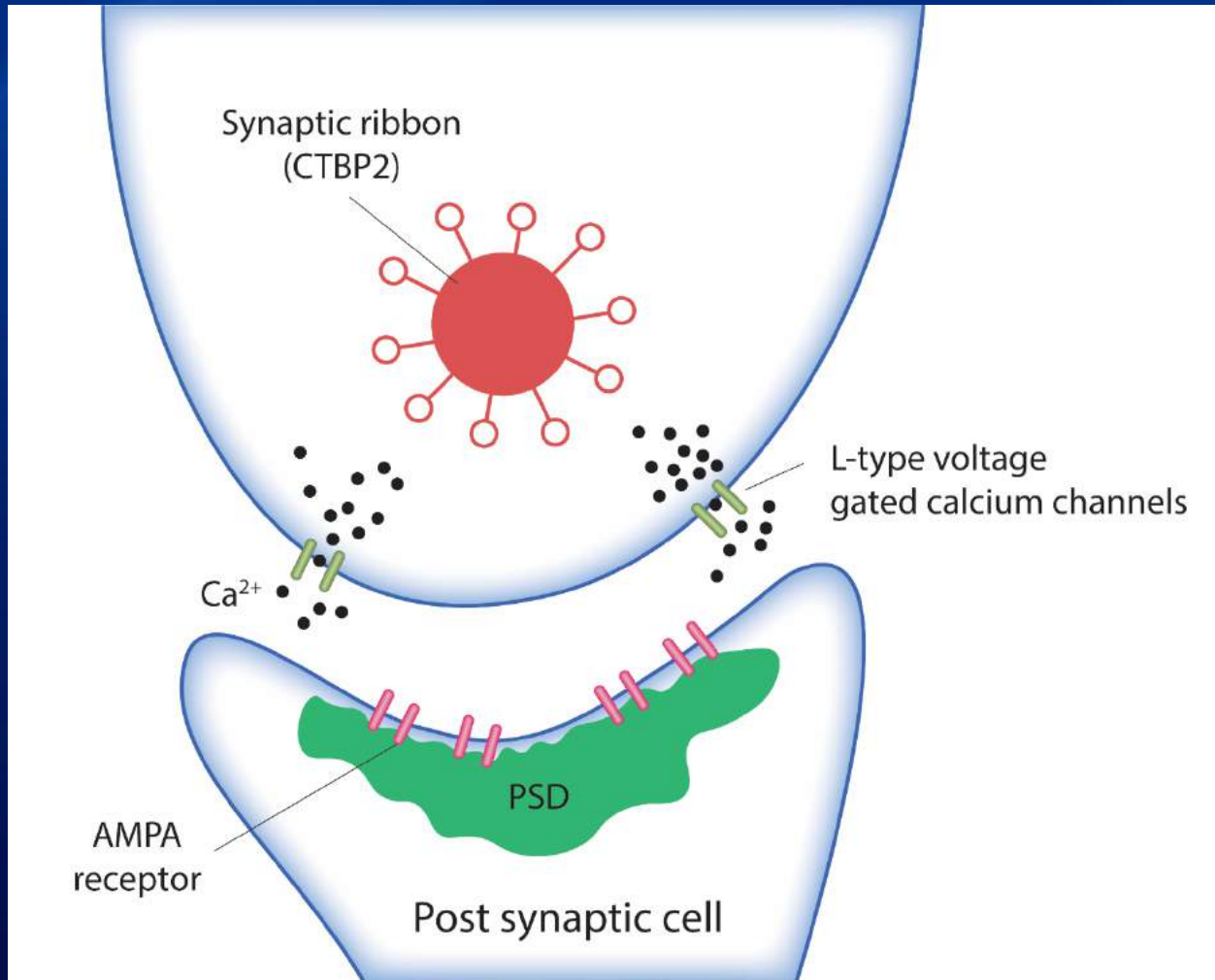
Table 2. Summary of the phenotypes of the vestibular mutants

Strain	Allele	Hair cell morphology	Acoustic vibrational sensitivity	Startle reflex Ca ²⁺ signal (%)
wildtype		Normal	Present	100 ± 56
<i>mariner</i> (<i>mar</i>)	<i>tc320b</i>	Bundle defect	Absent	9 ± 8

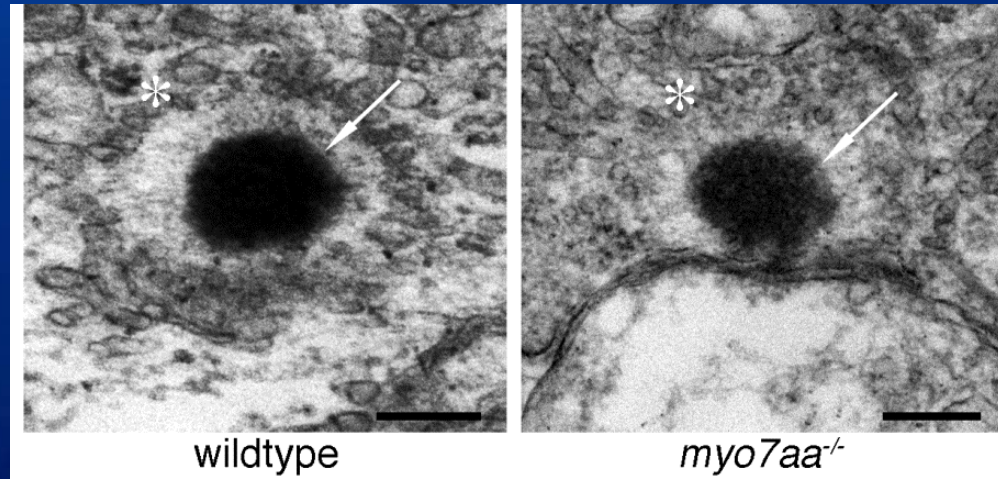
***Mariner* is defective in *myosin VIIA*: a zebrafish model for human hereditary deafness**

Sylvain Ernest, Gerd-Jörg Rauch¹, Pascal Haffter¹, Robert Geisler¹, Christine Petit and Teresa Nicolson²⁺

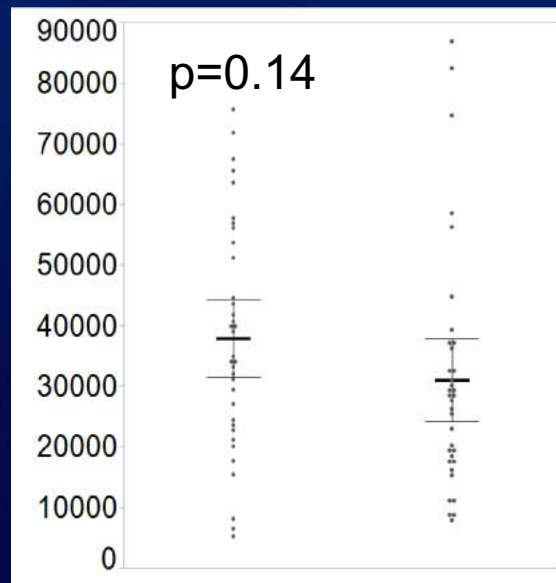




myo7aa^{-/-} mutant zebrafish have comparable ribbon area, but fewer glutamatergic vesicles

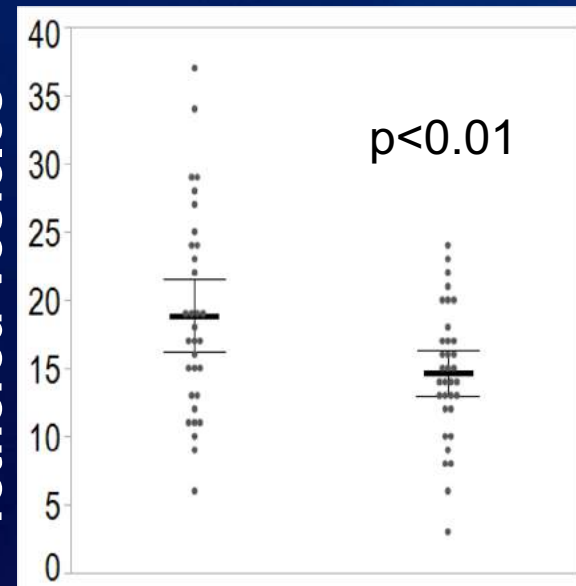


Ribbon Area (nm²)



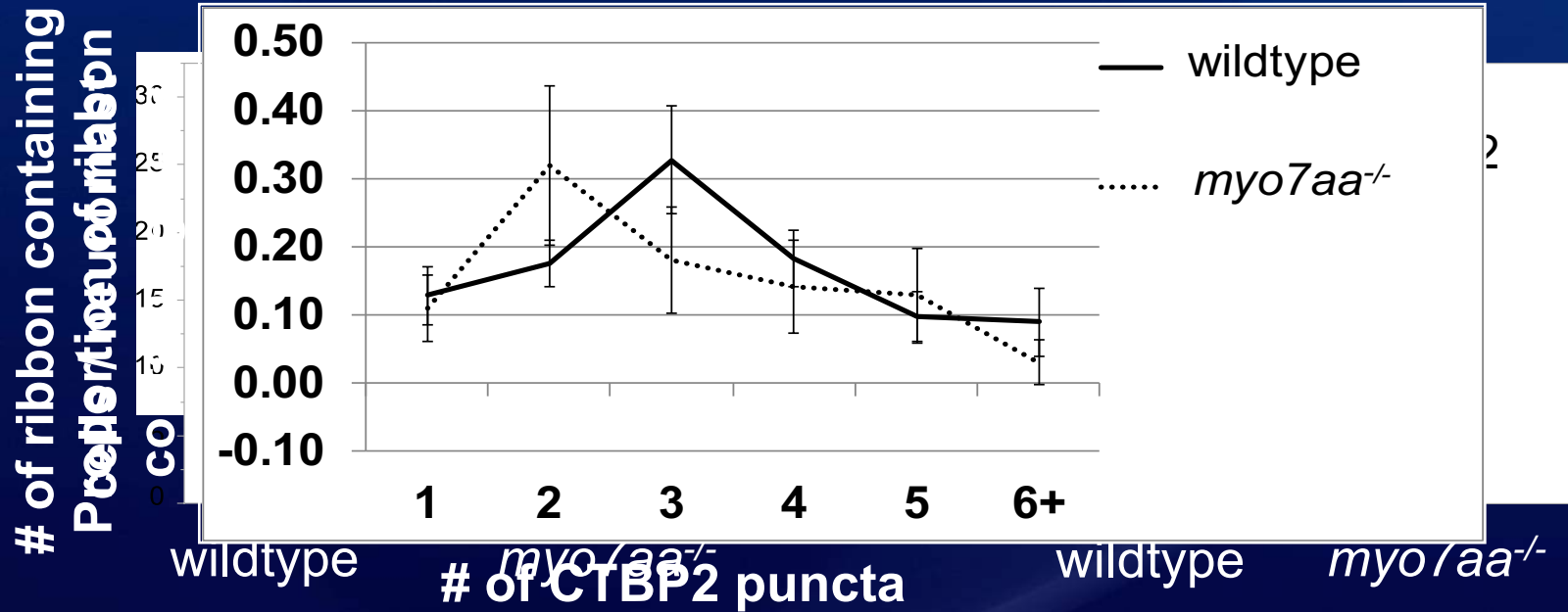
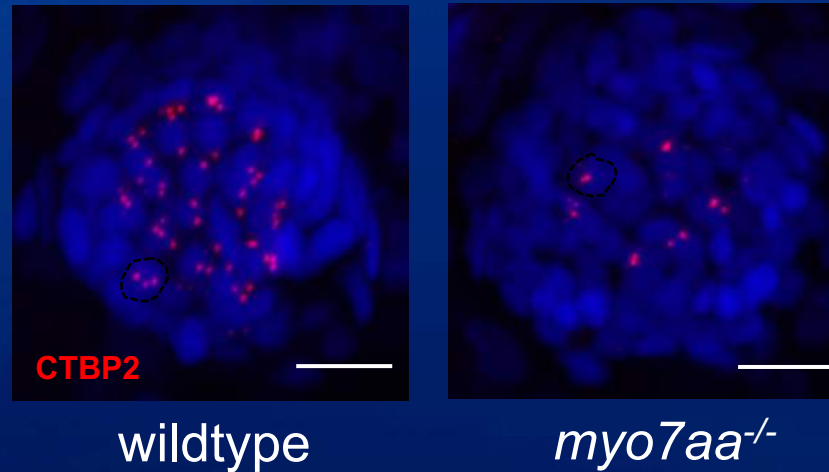
wildtype *myo7aa*^{-/-}

Tethered Vesicles

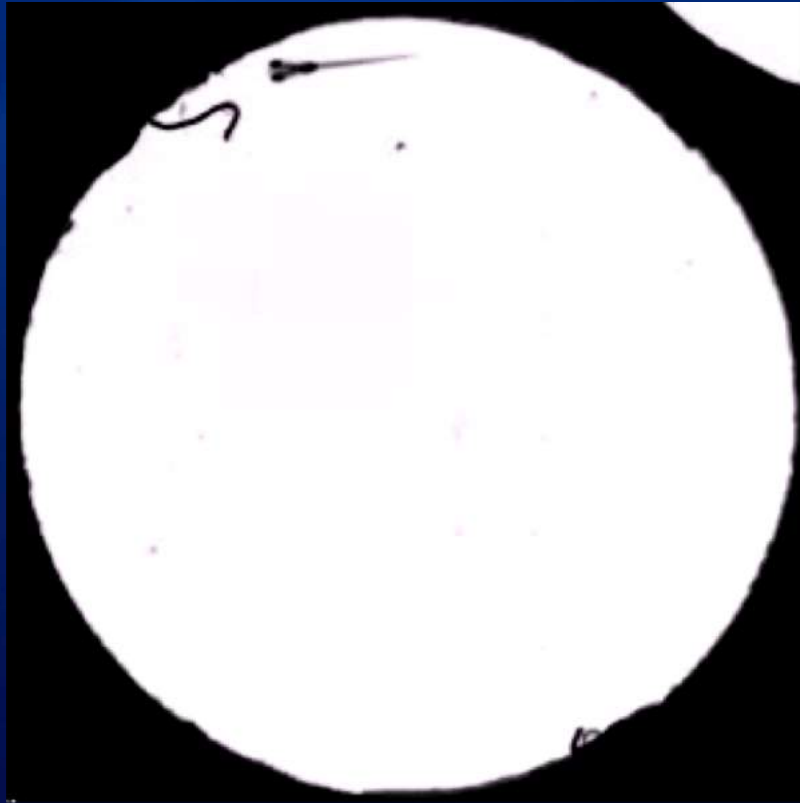


wildtype *myo7aa*^{-/-}

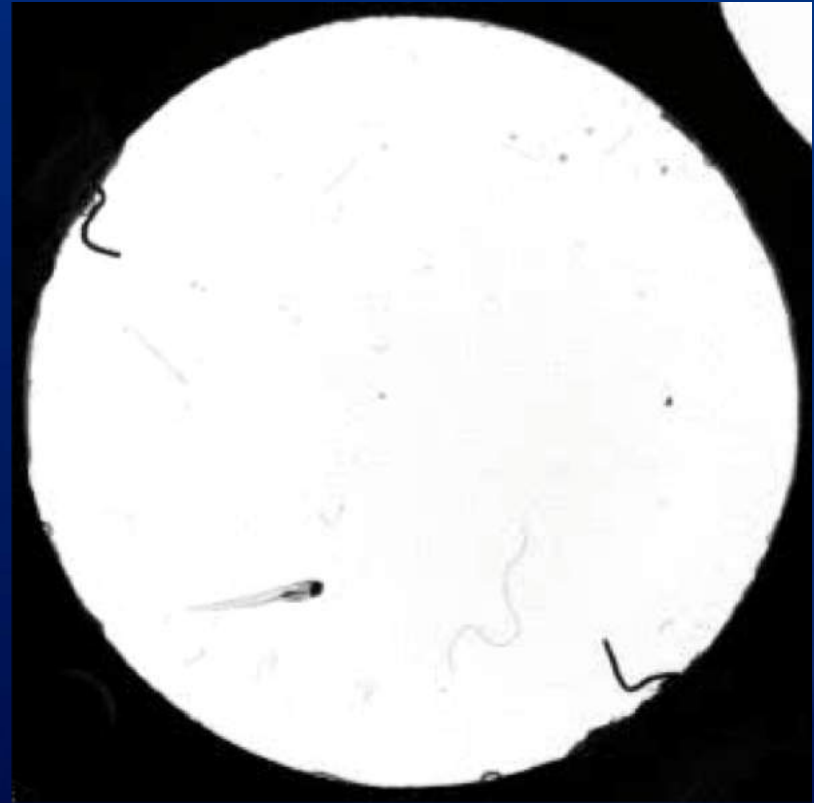
myo7aa^{-/-} mutant zebrafish have different distribution of CTBP2 puncta



Quantification of behavior between the wildtype and *myo7aa*^{-/-} mutant

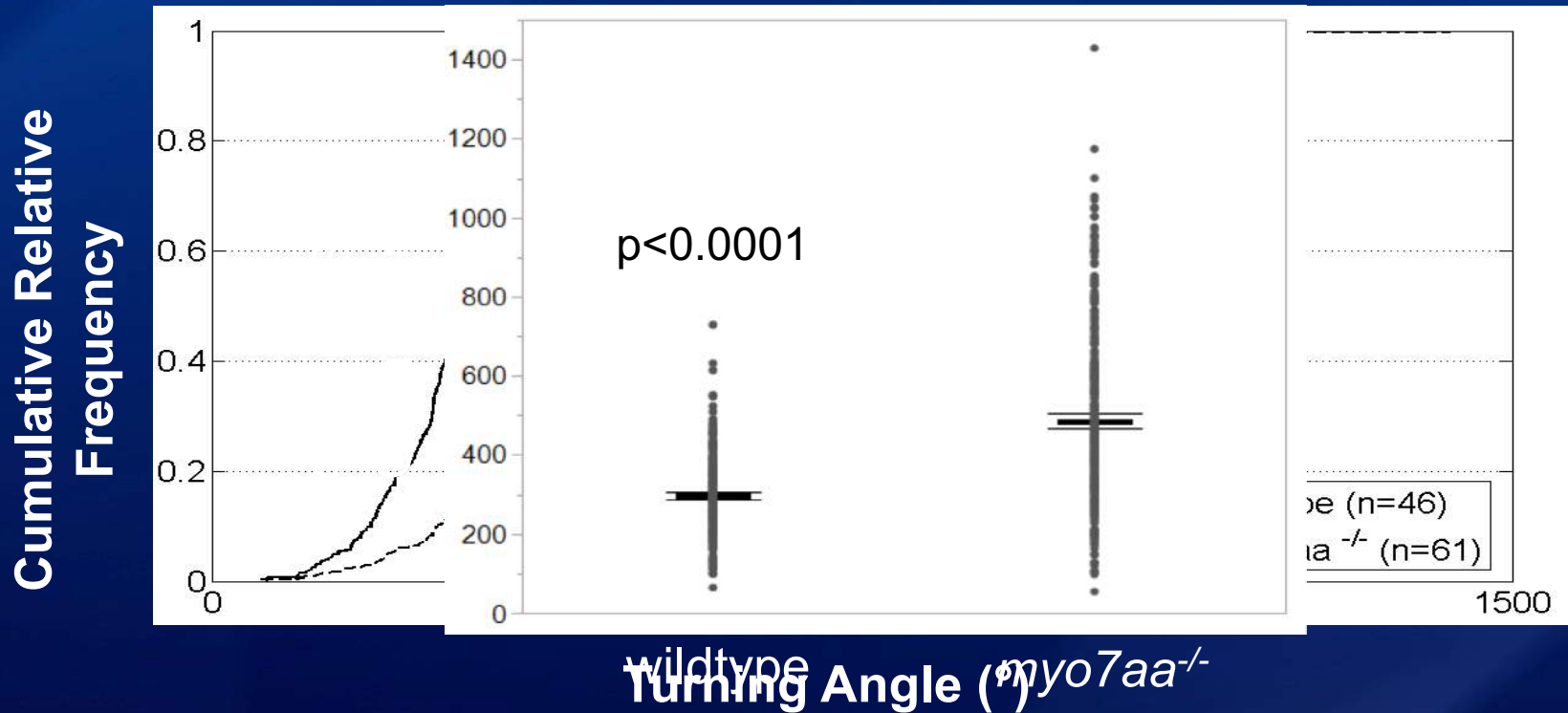


wildtype

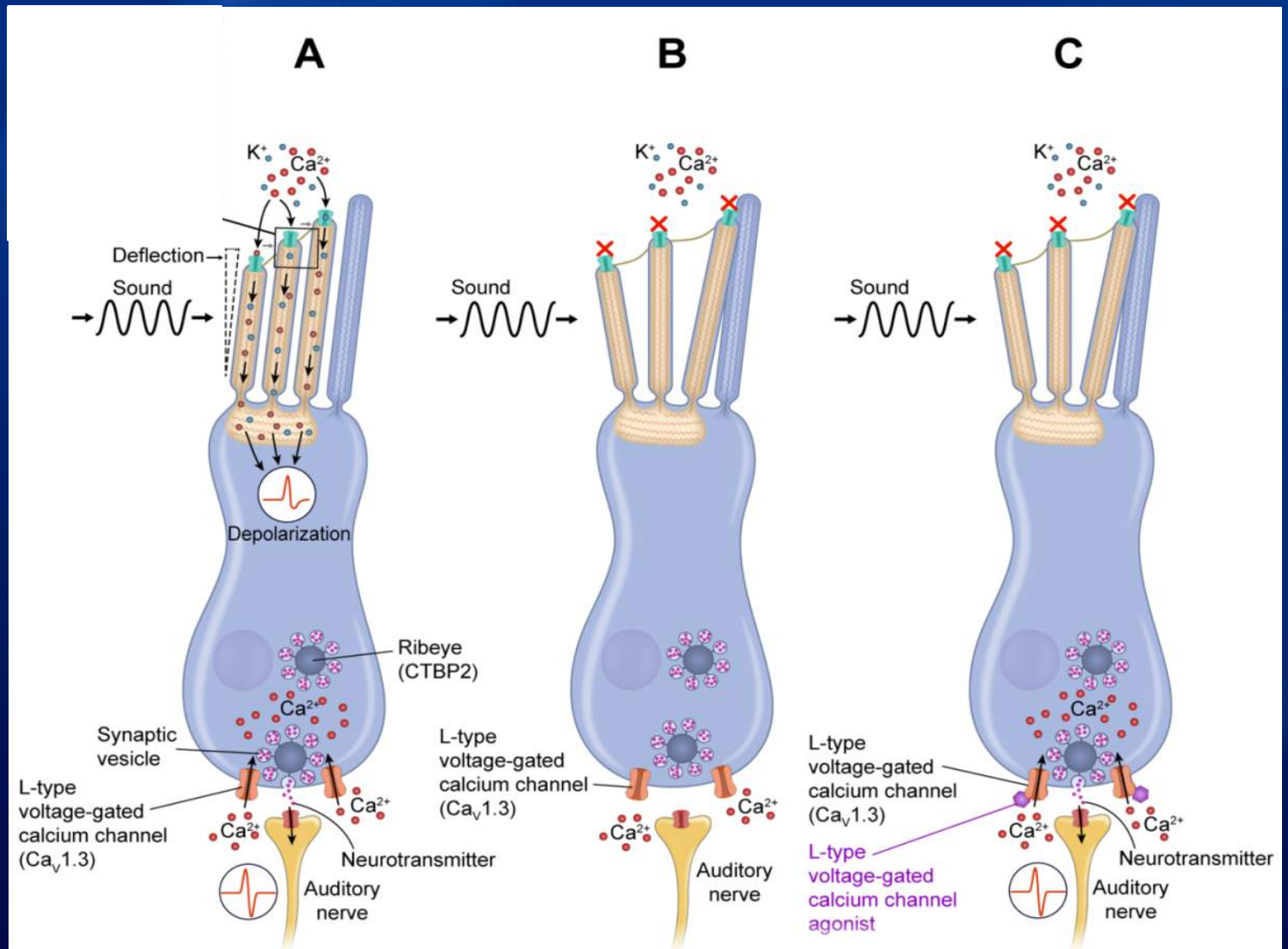


myo7aa^{-/-}

myo7aa^{-/-} mutant zebrafish have larger turning angles



Can modulating the $Ca_v1.3a$ channel using drugs provide a therapeutic effect in the *myo7aa*^{-/-} mutant zebrafish?



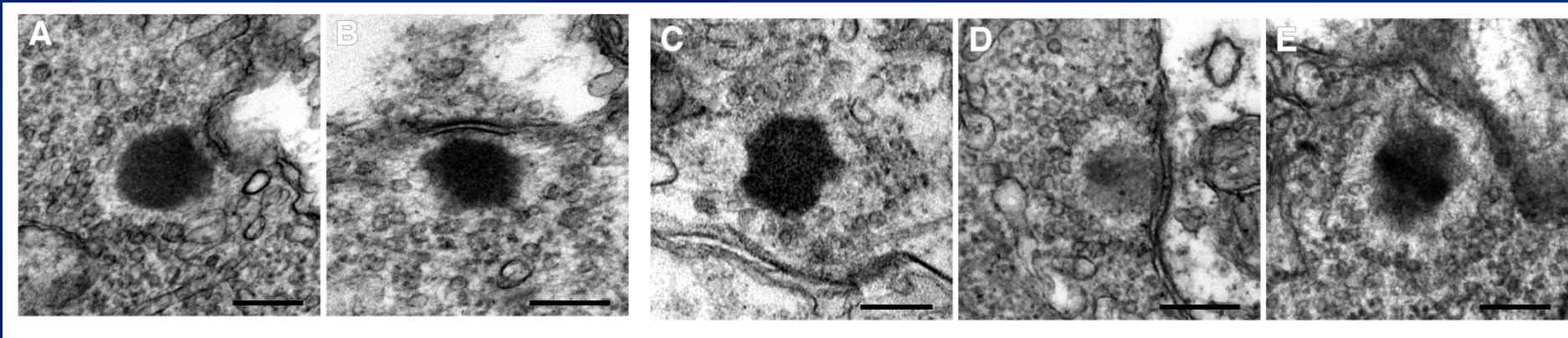
L-type voltage gated calcium channel agonists tested in this study:

(±)- Bay K
8644

Nefiracetam

(R)-
Baclofen

(R)-Baclofen increases ribbon area and number of tethered vesicles in *myo7aa*^{-/-} mutant zebrafish



wildtype
untreated

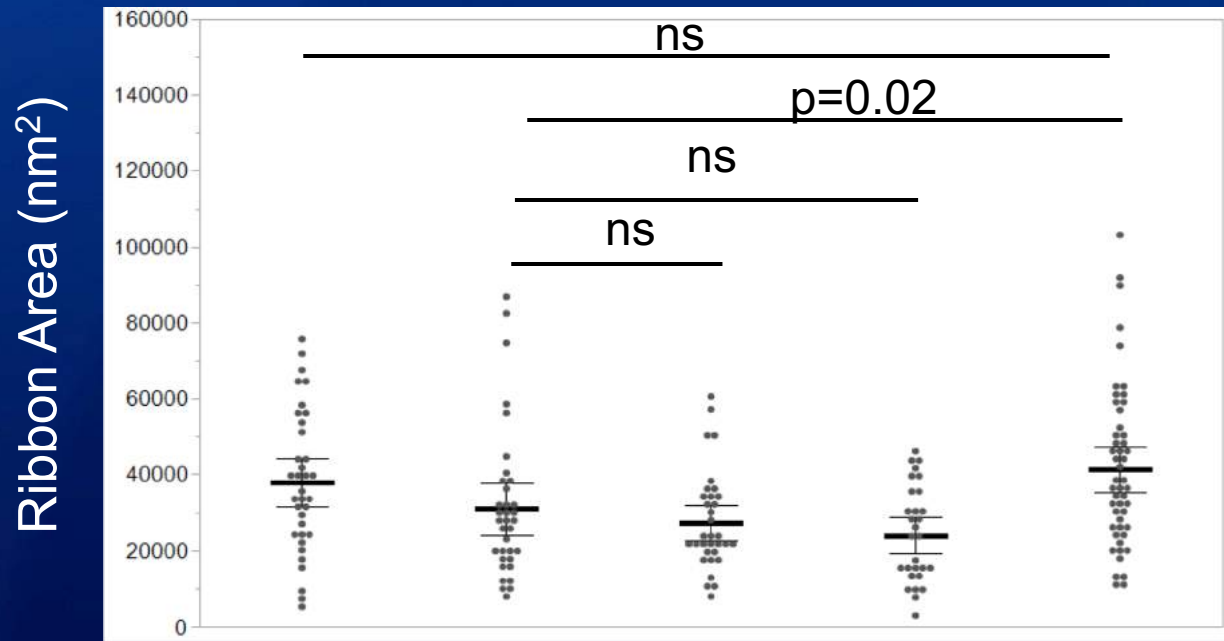
myo7aa^{-/-}
untreated

myo7aa^{-/-}
(±)-Bay K 8644

myo7aa^{-/-}
Nefiracetam

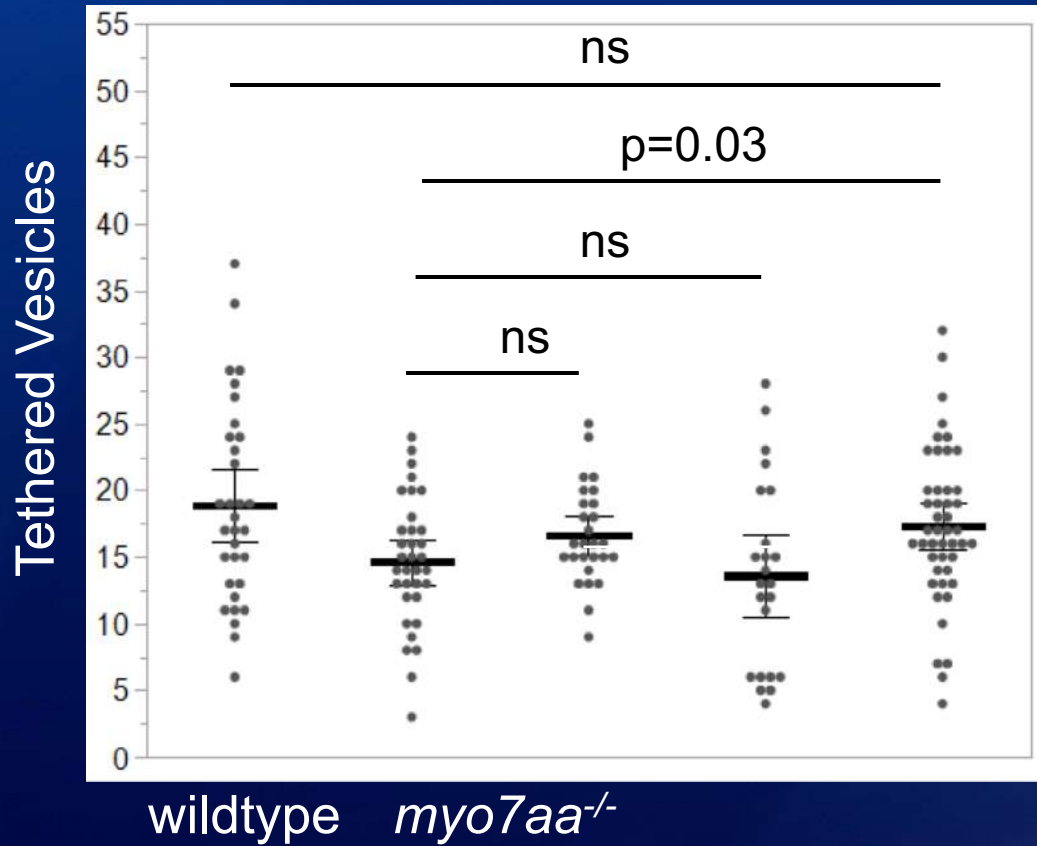
myo7aa^{-/-}
(R)-Baclofen

(R)-Baclofen increases ribbon area in *myo7aa*^{-/-} mutant zebrafish



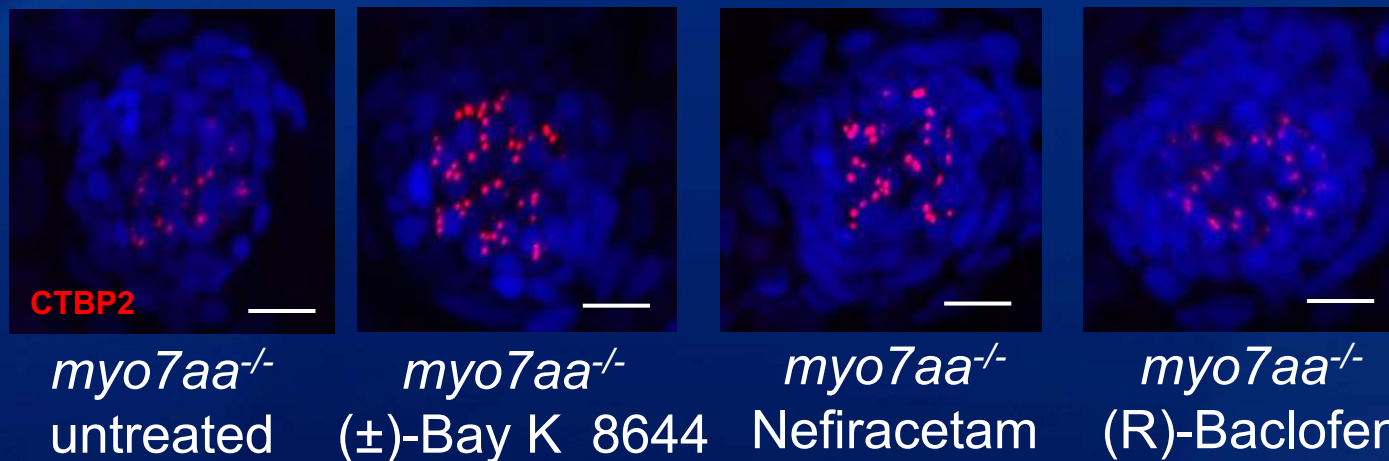
	wildtype	<i>myo7aa</i> ^{-/-}			
(±)-Bay K 8644	-	-	+	-	-
Nefiracetam	-	-	-	+	-
(R)-Baclofen	-	-	-	-	+

(R)-Baclofen increases number of tethered vesicles in *myo7aa*^{-/-} mutant zebrafish

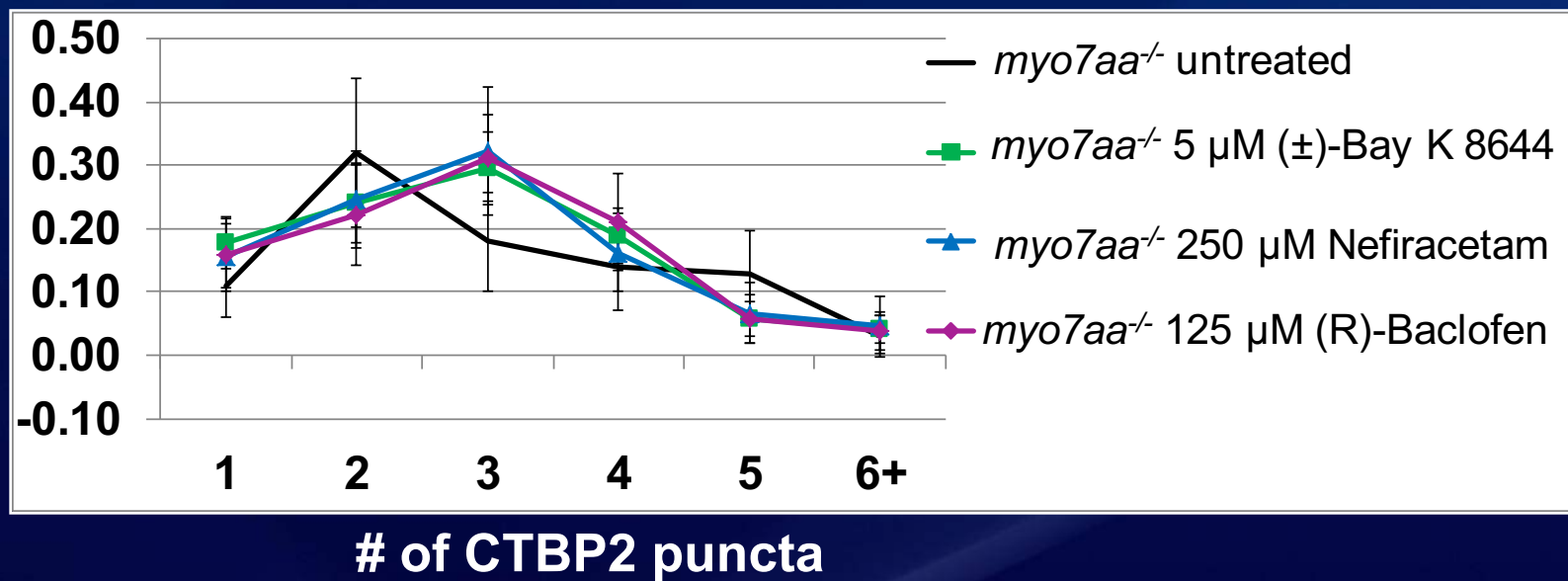


(±)-Bay K 8644	-	-	+	-	-
Nefiracetam	-	-	-	+	-
(R)-Baclofen	-	-	-	-	+

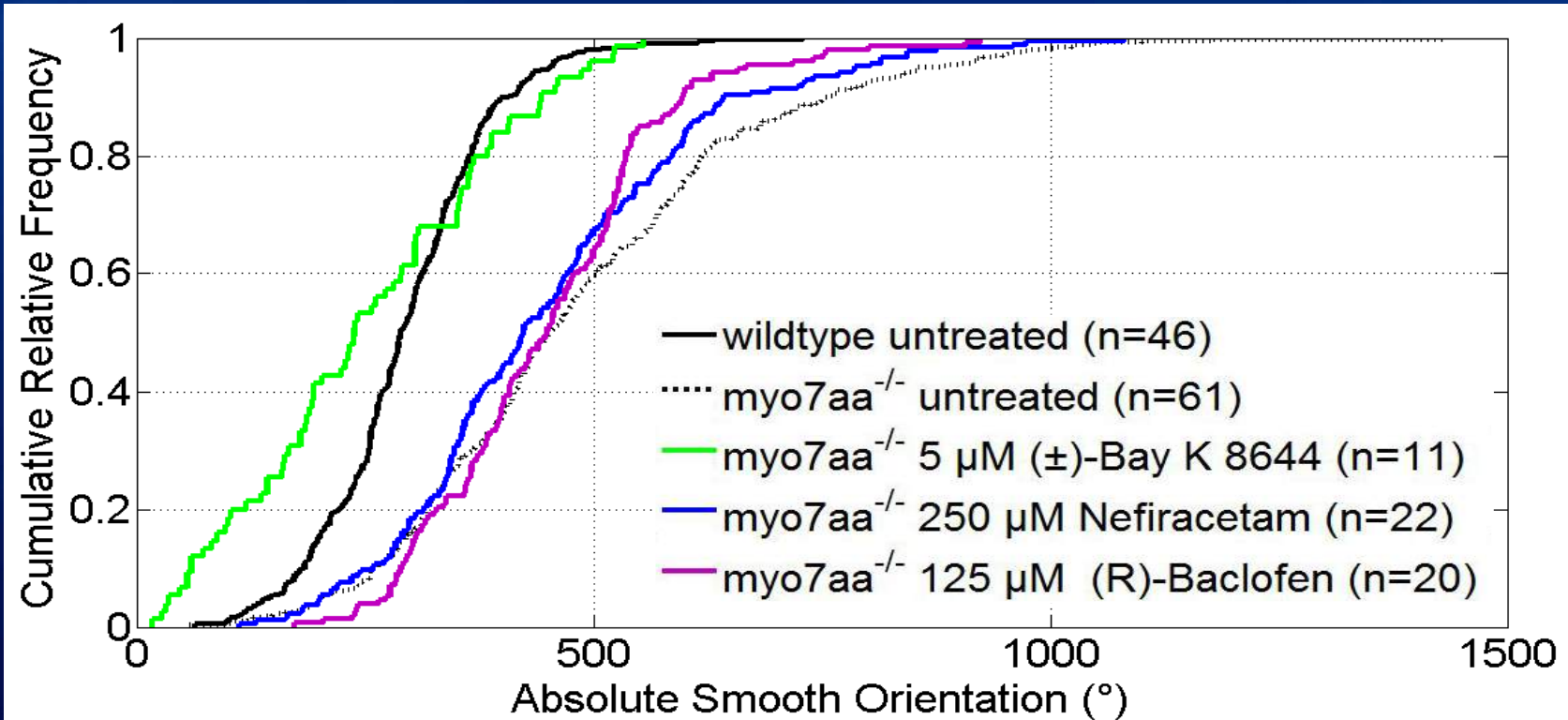
L-type voltage-gated calcium channel agonists alter CTBP2 distribution in *myo7aa*^{-/-} mutant zebrafish



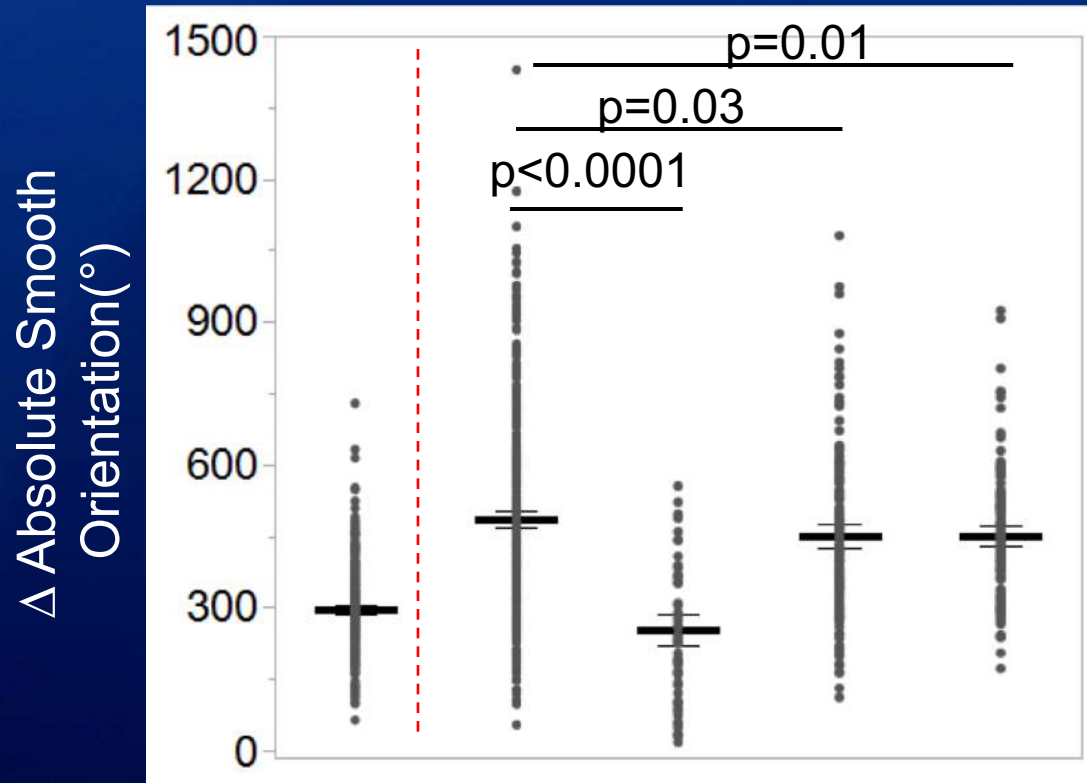
Proportion of ribbon containing cells



L-type voltage-gated calcium channel agonists decrease turning angle in *myo7aa*^{-/-} mutant zebrafish



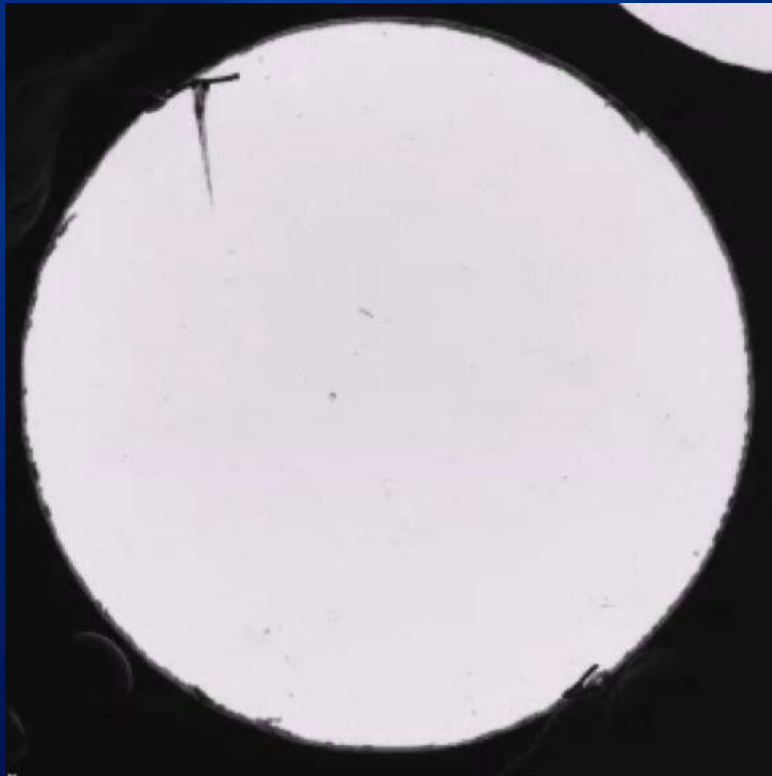
L-type voltage-gated calcium channel agonists decrease turning angle in *myo7aa*^{-/-} mutant zebrafish



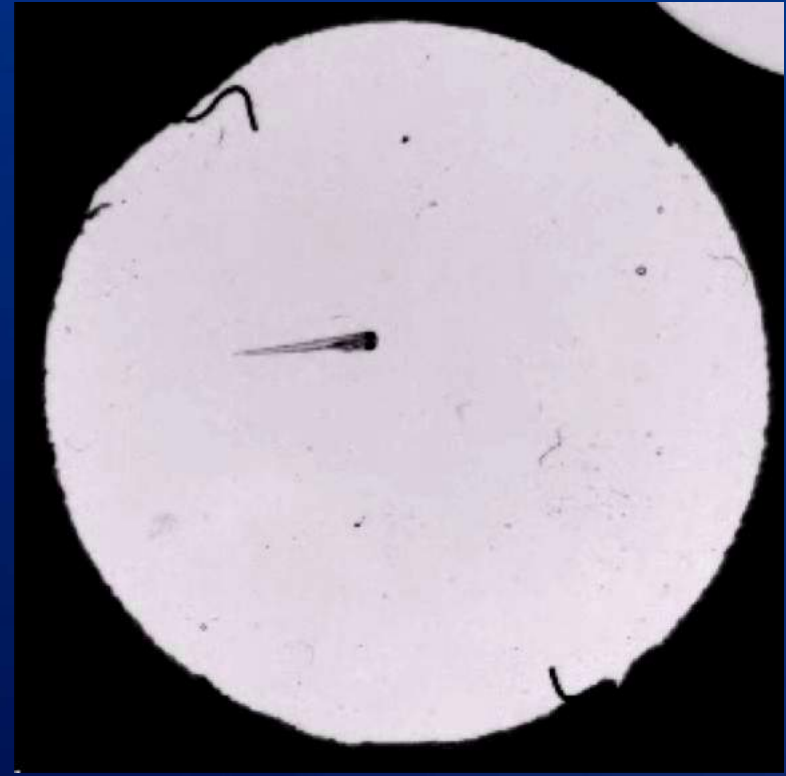
	wildtype		<i>myo7aa</i> ^{-/-}		
(±)-Bay K 8644	-	-	+	-	-
Nefiracetam	-	-	-	+	-
(R)-Baclofen	-	-	-	-	+

L-type voltage-gated calcium channel agonists decrease turning angle in *myo7aa*^{-/-} mutant zebrafish



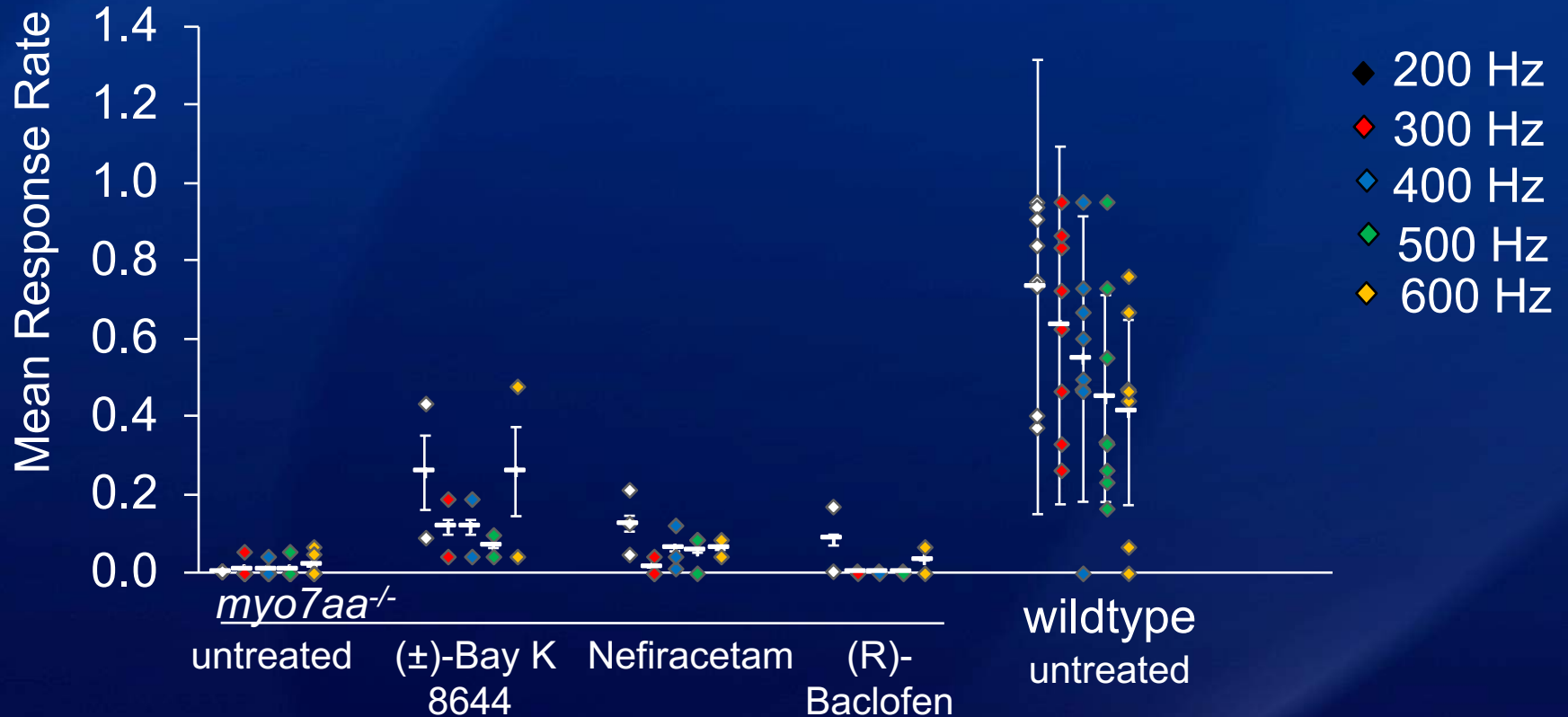


myo7aa^{-/-}
Nefiracetam



myo7aa^{-/-}
(R)-Baclofen

L-type voltage-gated calcium channel agonists increase acoustic startle response in *myo7aa*^{-/-} mutant zebrafish



Summary and Conclusions

- *myo7aa*^{-/-} mutants have a different synaptic morphology:
 - decreased number of vesicles tethered to ribeye
 - decreased number of ribbon containing cells
 - decreased total CTBP2 puncta per neuromast
 - different distribution of CTBP2 puncta
- *myo7aa*^{-/-} mutants have larger turning angles as part of their swimming behavior

Summary and Conclusions

- Behavioral and synaptic morphological differences can be modified by drugs with L-type voltage-gated calcium channel activity
 - (R)-Baclofen increases ribbon area and number of tethered vesicles
 - L-type voltage-gated calcium channel agonists shift distribution of CTBP2 puncta to more closely resemble wildtype
 - L-type voltage-gated calcium channel agonists decrease turning angles in swimming behavior and increase acoustic startle response

Future Directions

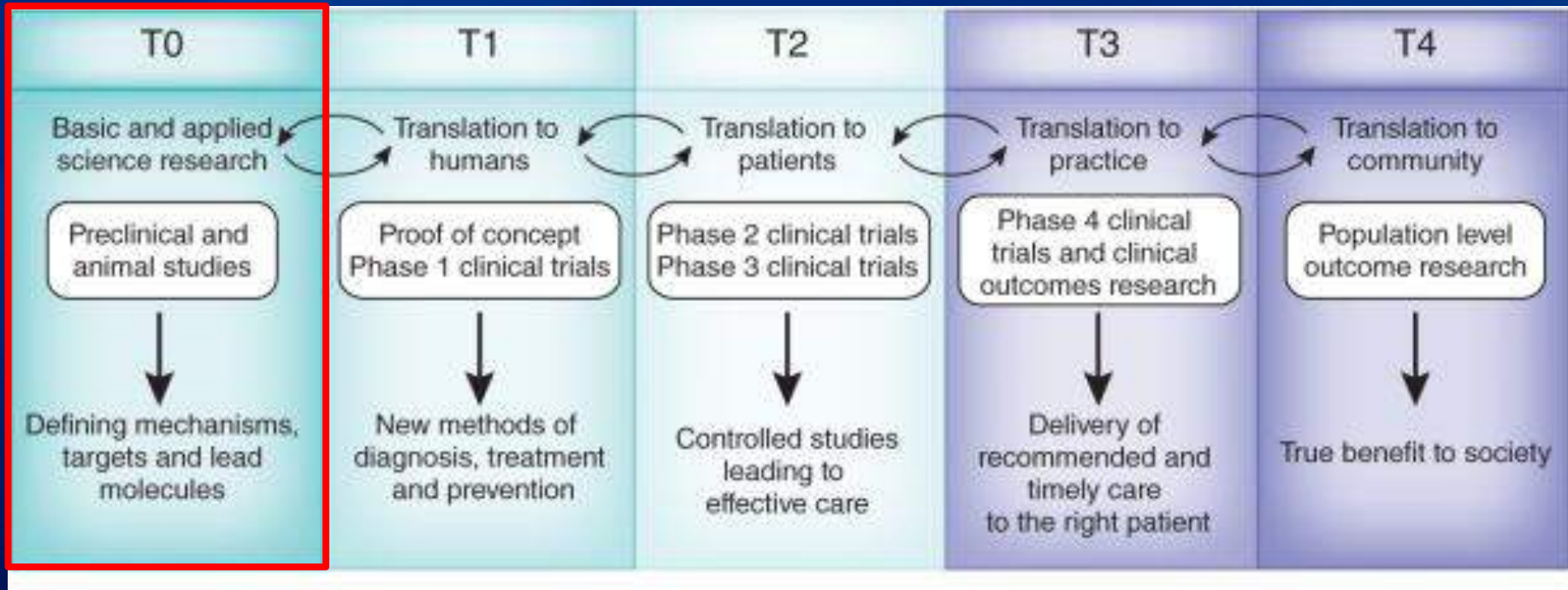
- Identify exact mechanism of action of Nefiracetam and (R)-Baclofen
- Assess calcium signal in hair cells with and without L-type voltage-gated calcium channel agonists

Future Directions

- Move to the mouse model of USH1 (*shaker-1*) and assess hearing thresholds through auditory brainstem responses upon injection with L-type voltage-gated calcium channel agonists.



Translational Research



Blumberg et al. 2012 Nat. Med

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