

Gene Therapy for Usher Syndrome (and other treatments)

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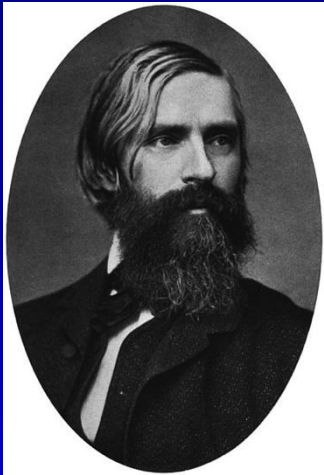
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Statistician

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Usher Syndrome - History



Albrecht von Graefe

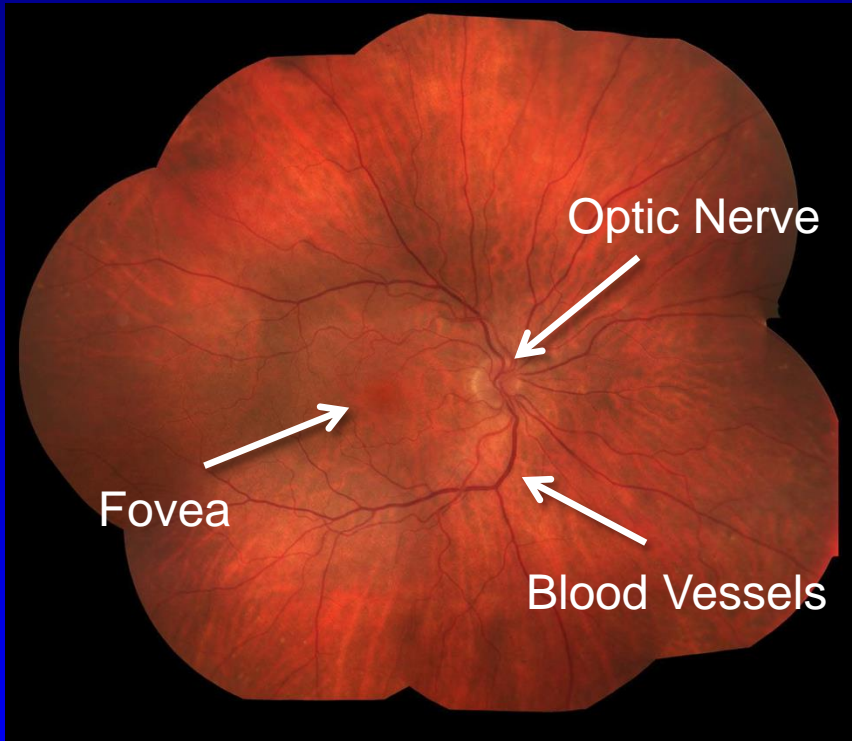
- First reported by Albrecht Von Graefe in 1858 where he described three brothers with deaf-blindness



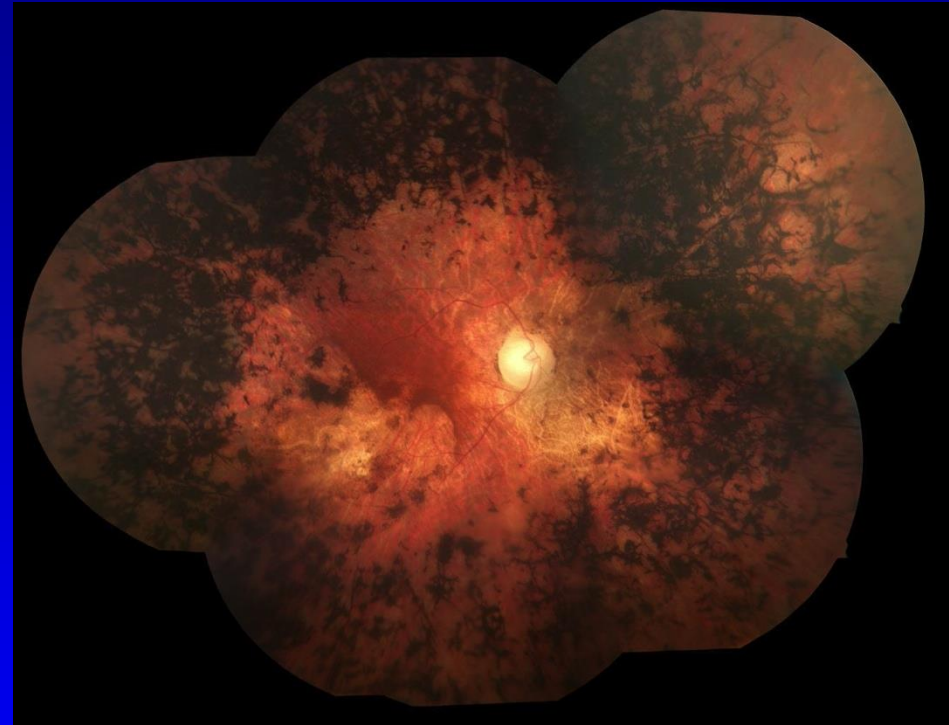
Charles Usher

- Scottish Ophthalmologist Charles Usher described 68 patients in 1912 with retinitis pigmentosa and deafness

Retinal Degeneration in Usher Syndrome



Normal Retina



Usher Syndrome

Anatomy of the Eye and Retina

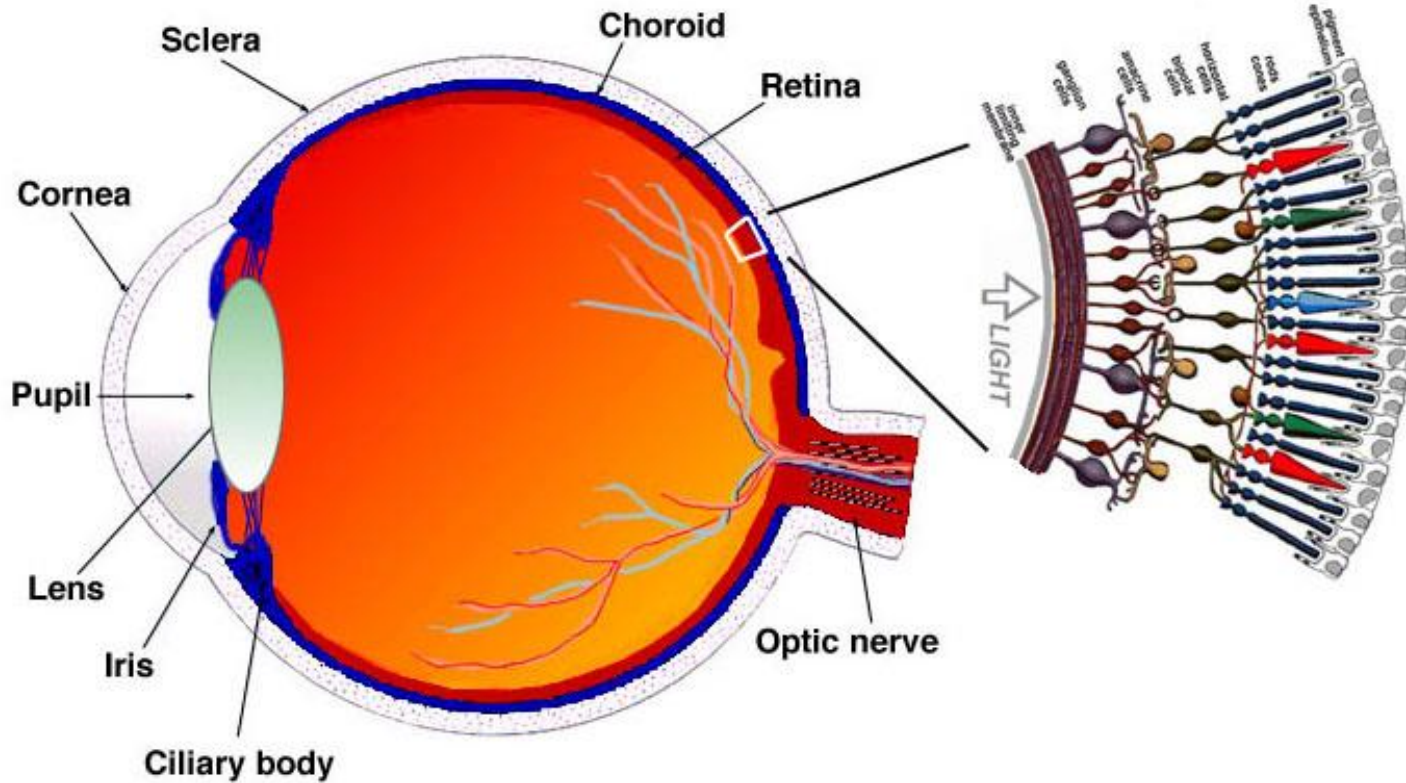


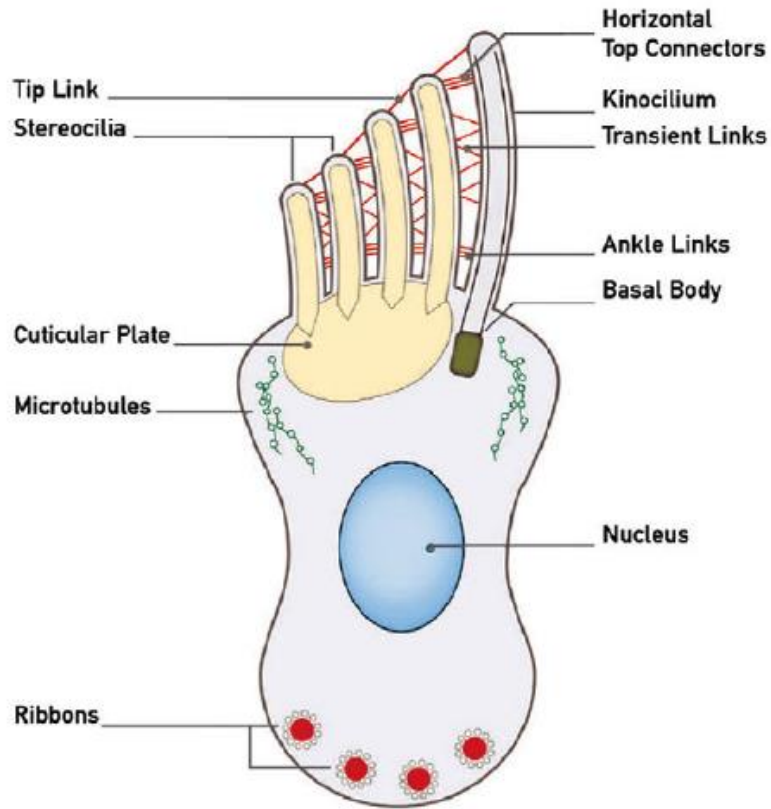
Fig. 1.1. A drawing of a section through the human eye with a schematic enlargement of the retina.

Images from Webvision

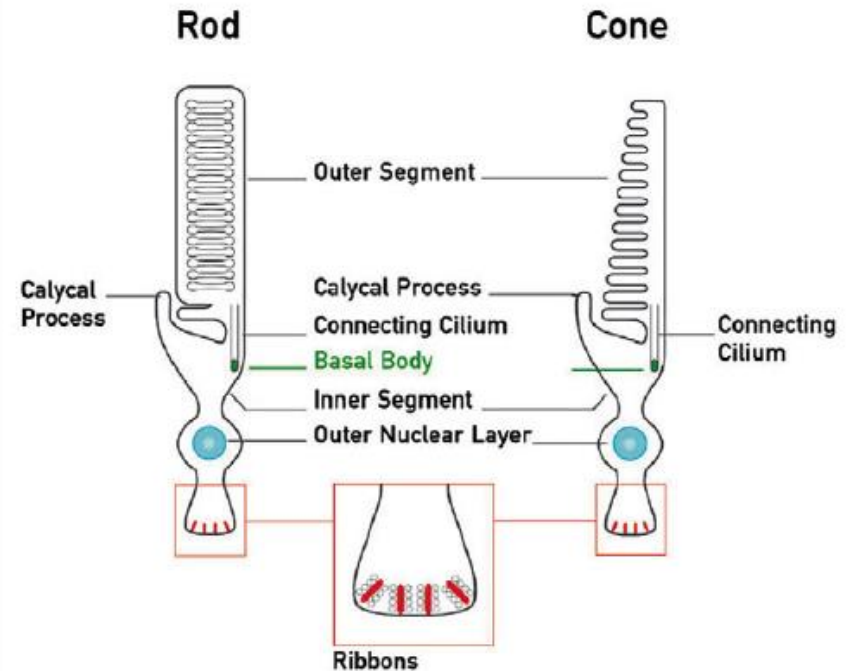
Rods and Cones die in inherited retinal degenerations

Hair cells and photoreceptors share specialized cilia

A



B



Usher Syndrome - Subtypes

Type I:

- Retinitis Pigmentosa – very severe
- Profound congenital deafness (cochlear implants)
- Vestibular dysfunction (balance problems)

Type II:

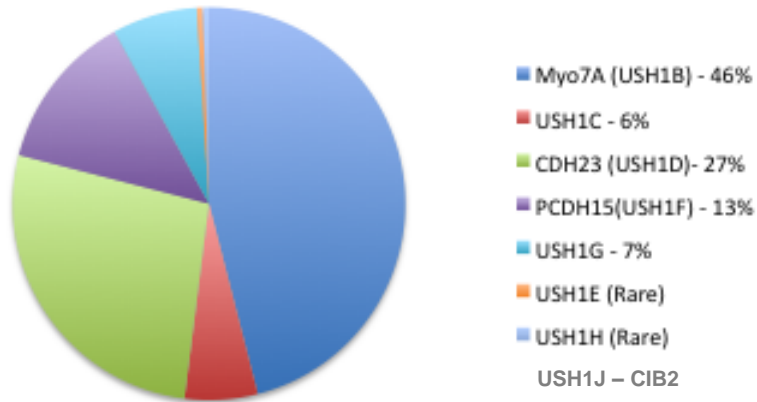
- Retinitis Pigmentosa – moderate to severe
- Severe congenital deafness (hearing aids)

Type III:

- Retinitis Pigmentosa – moderate to severe
- Progressive deafness and vestibular dysfunction

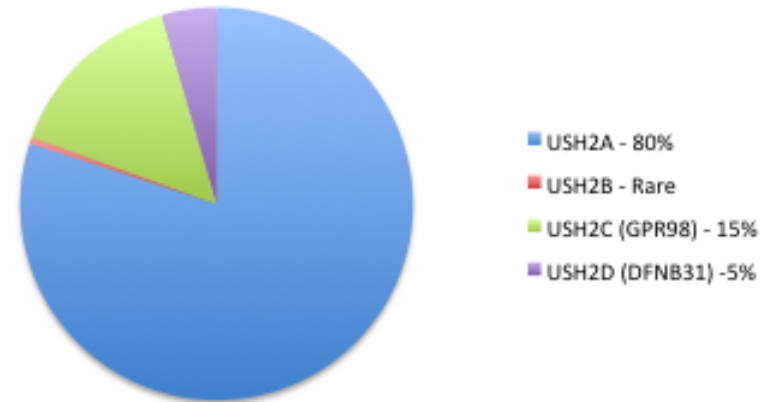
Genes Involved in Usher Syndrome

USHER SYNDROME I



At least 8 genes

USHER SYNDROME II



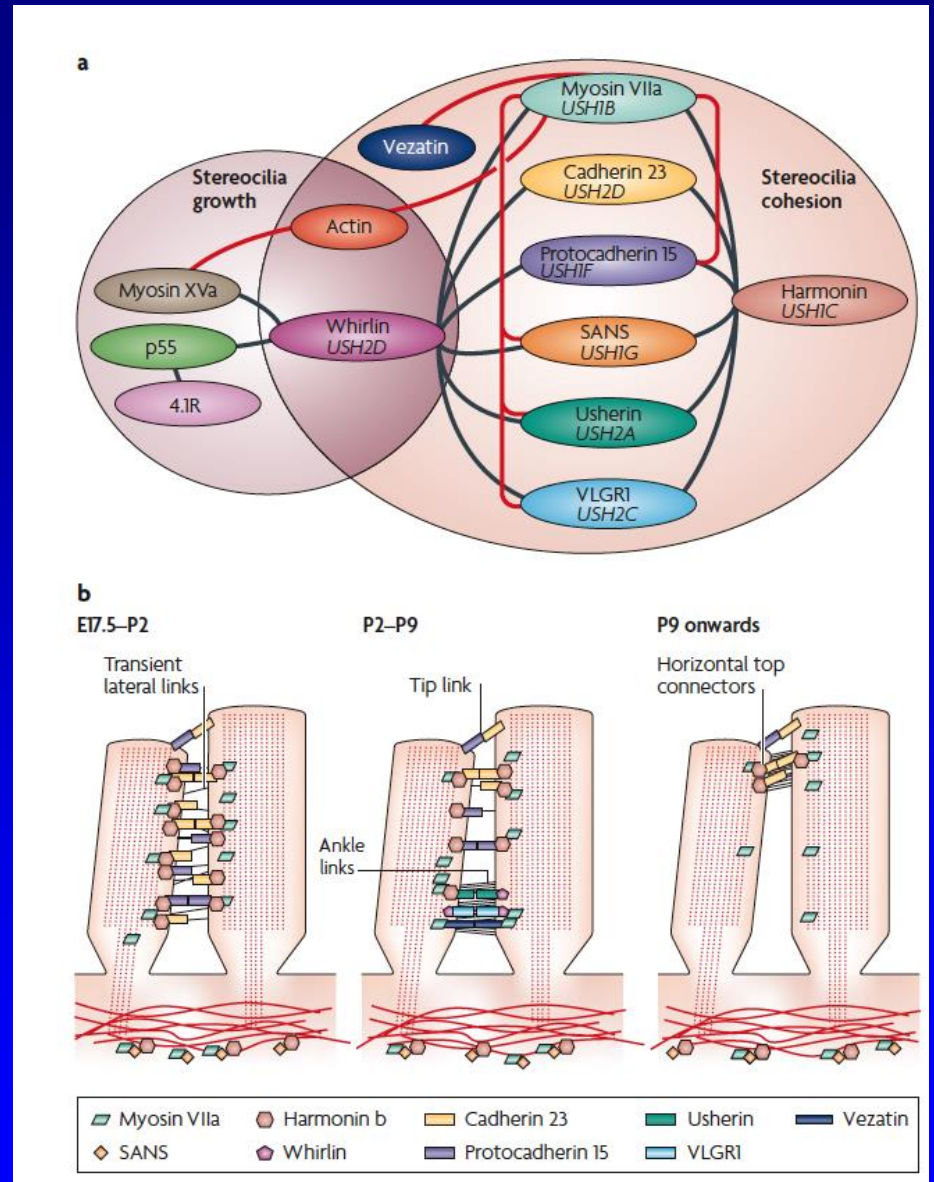
At least 3 genes

USHER SYNDROME III



At least 2 genes

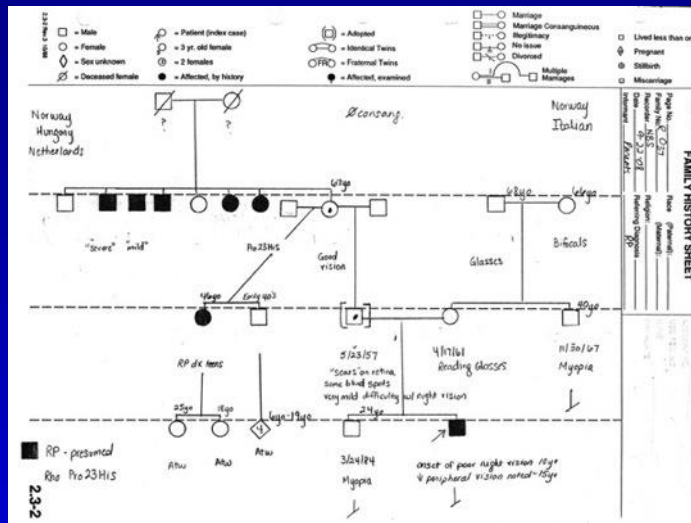
The Usher Interactome



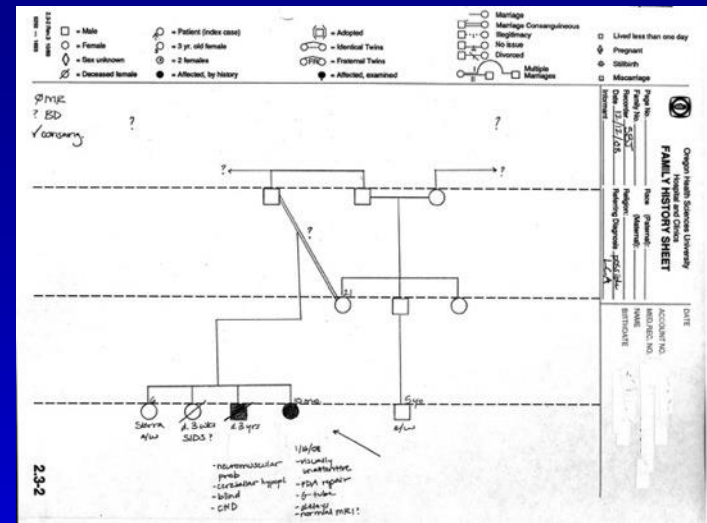
From Brown et al.

Tools to Study Usher Syndrome

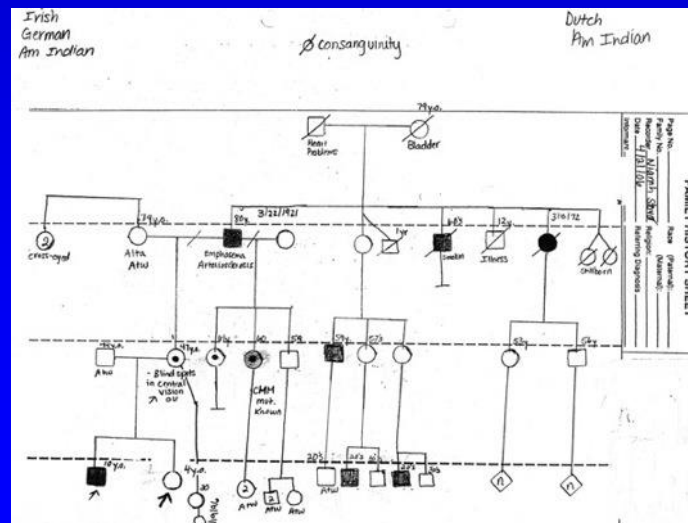
Family History



Autosomal Dominant

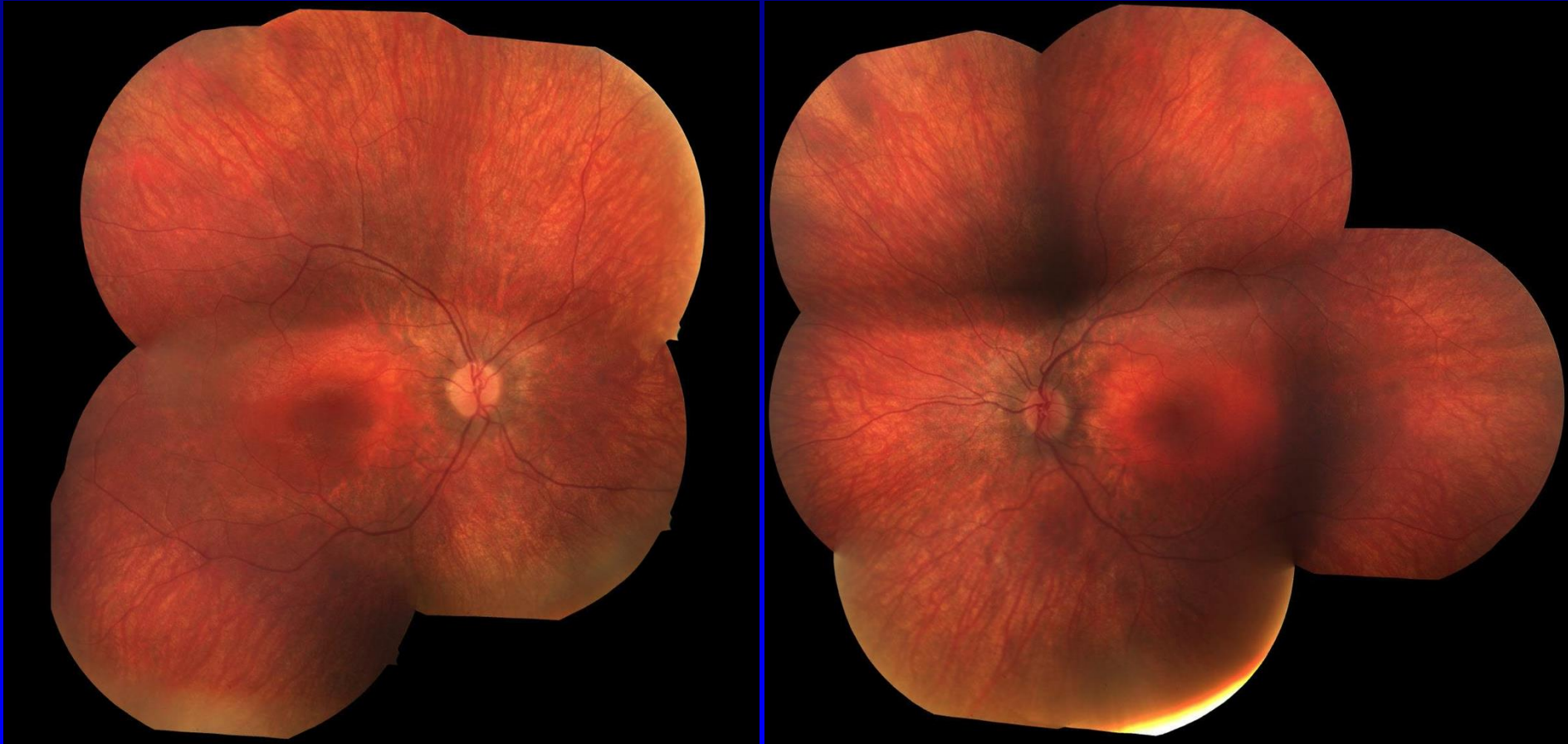


Autosomal Recessive



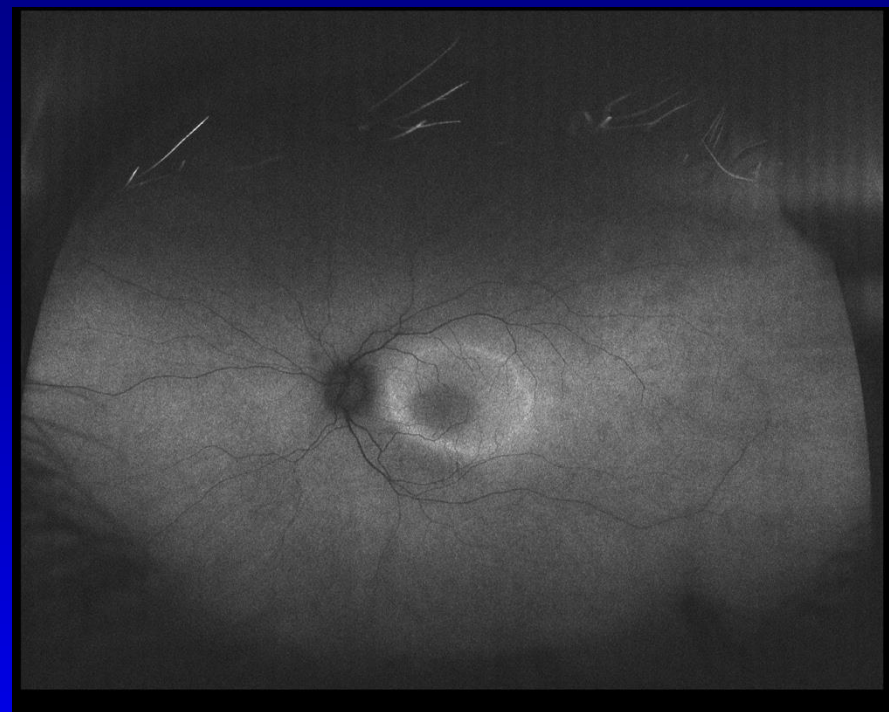
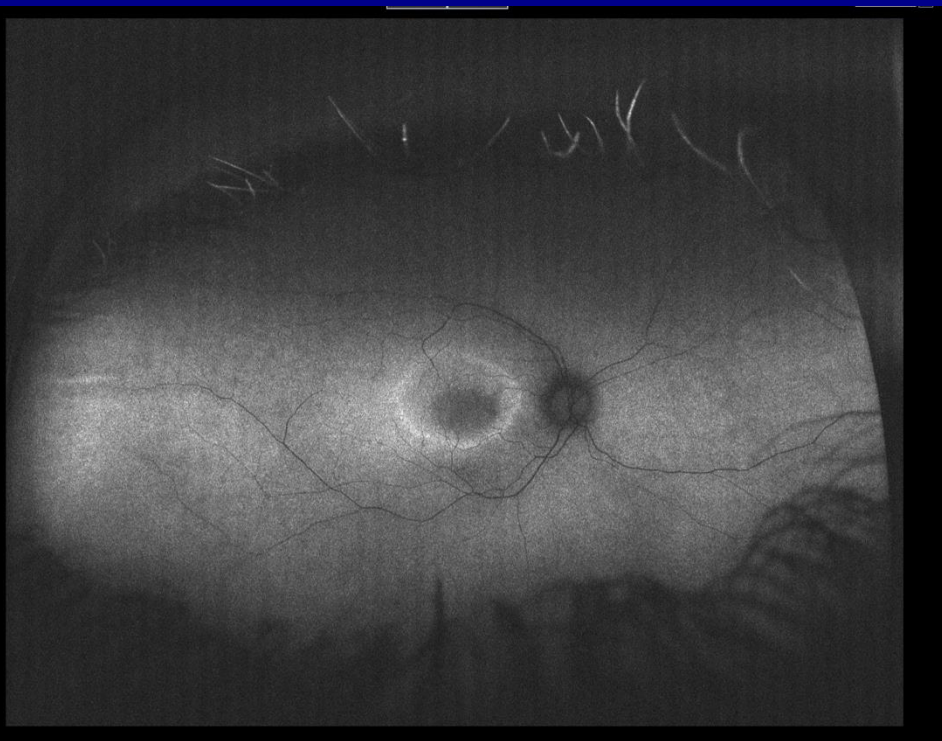
X-Linked Recessive

Fundus Photography



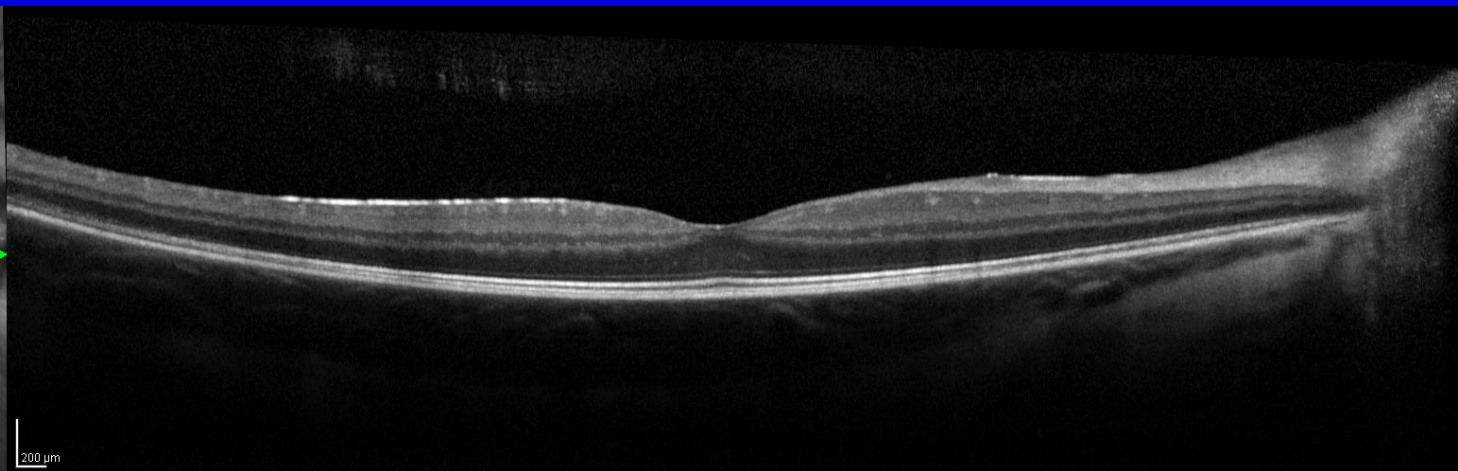
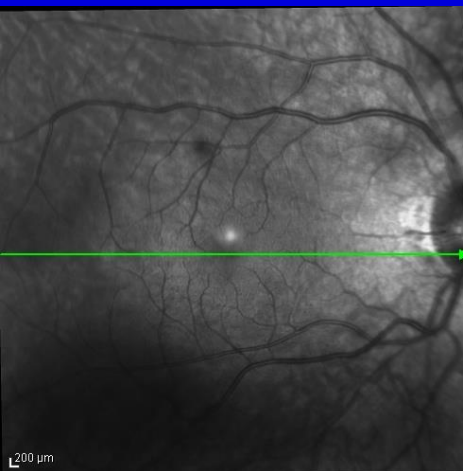
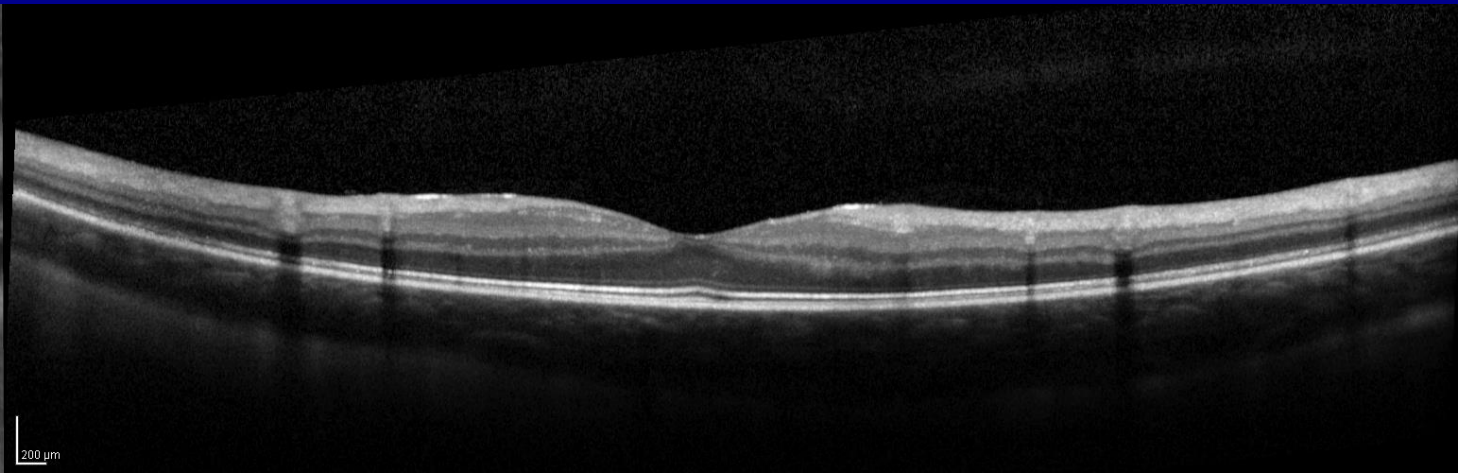
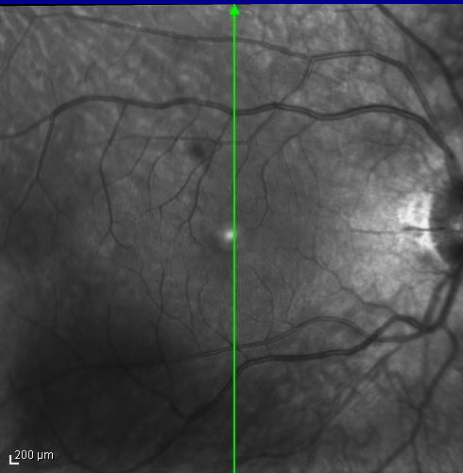
Patient with Type I Usher Syndrome

Short Wavelength Autofluorescence



Patient with Type I Usher Syndrome

15 year old with USH1B from mutations of MYO7A

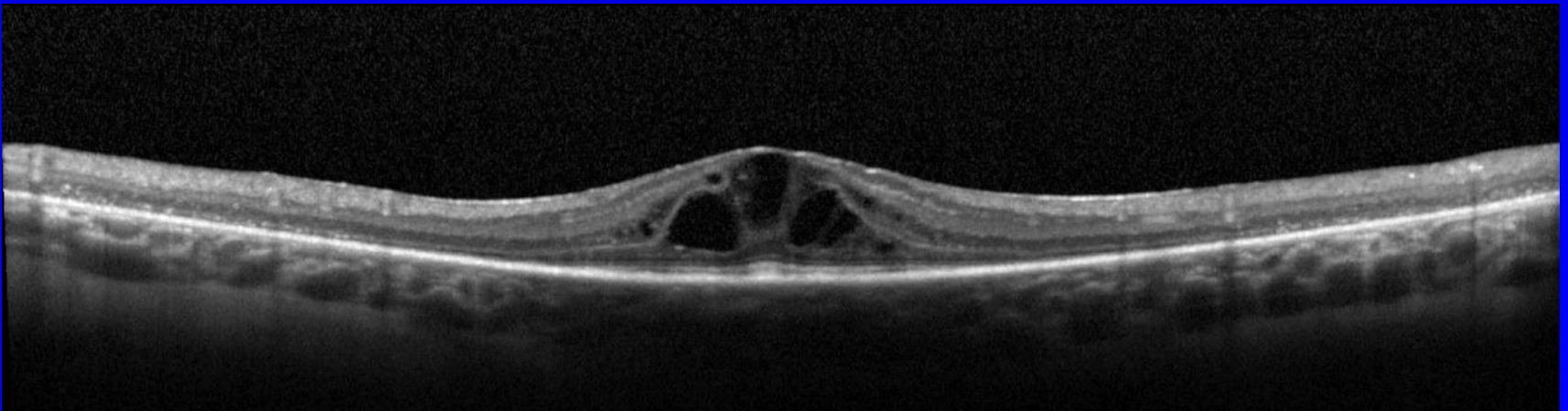


Importance of OCT for detecting Macular Edema

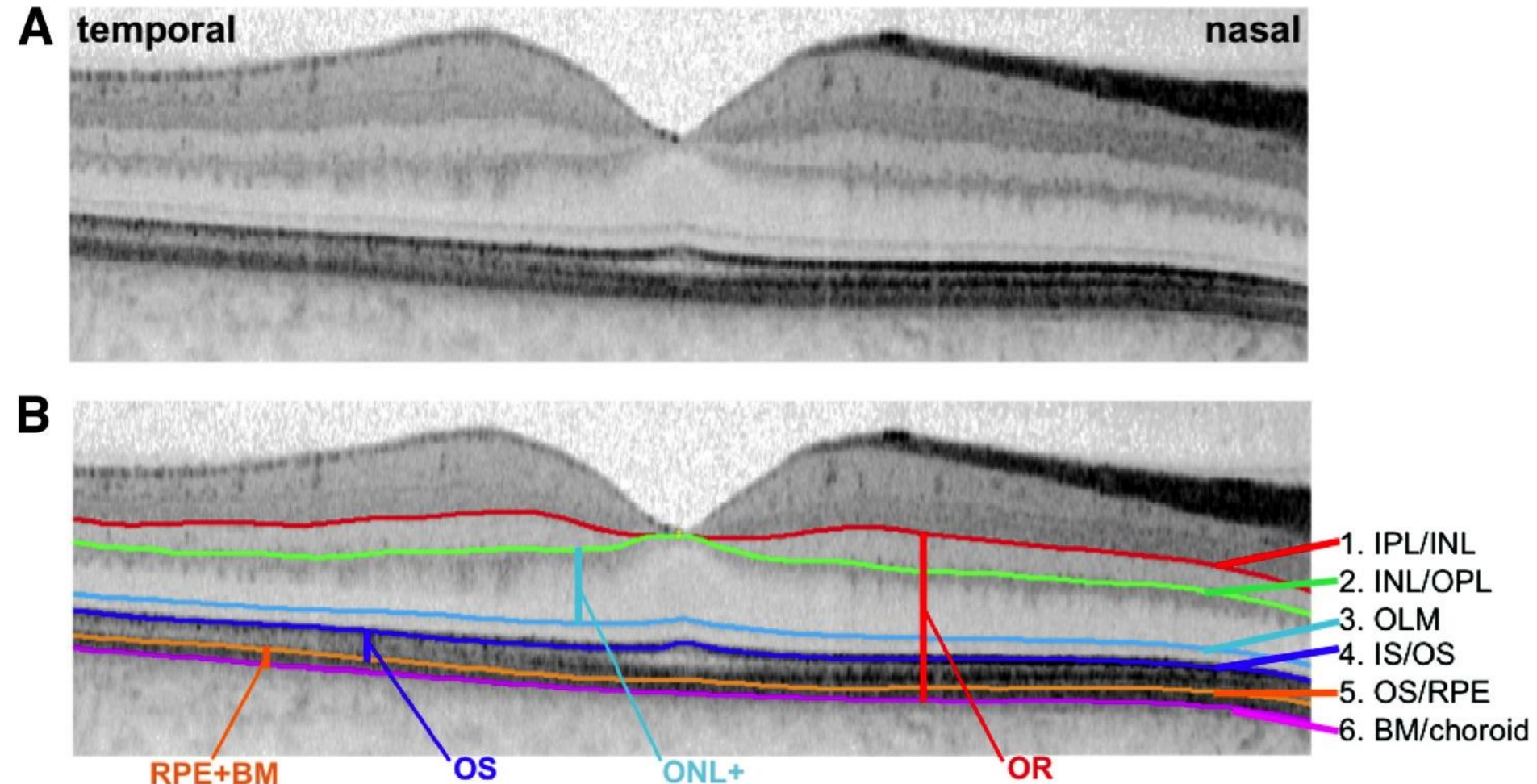
Healthy Eye



Cystoid Macular Edema in an Usher Patient

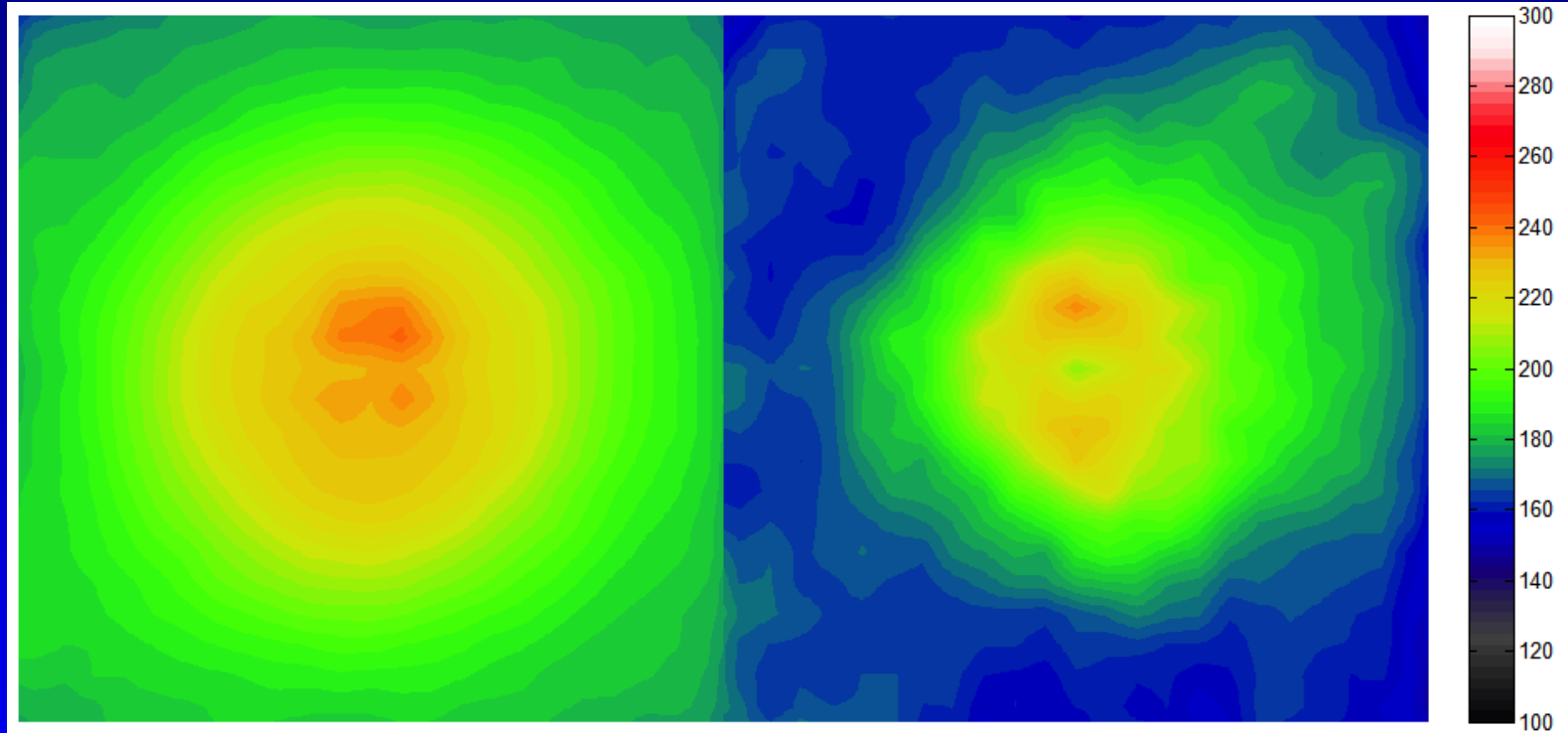


Segmentation of Retinal Layers



Lazow, MA, et al. Transition Zones between Healthy and Diseased Retina in Choroideremia (CHM), Stargardt Disease (STGD) Retinitis Pigmentosa (RP). Invest Ophthalmol Vis Sci 2011; 52: 9581

SD-OCT: Outer Retinal Thickness



Normal group AVG

AMD group AVG

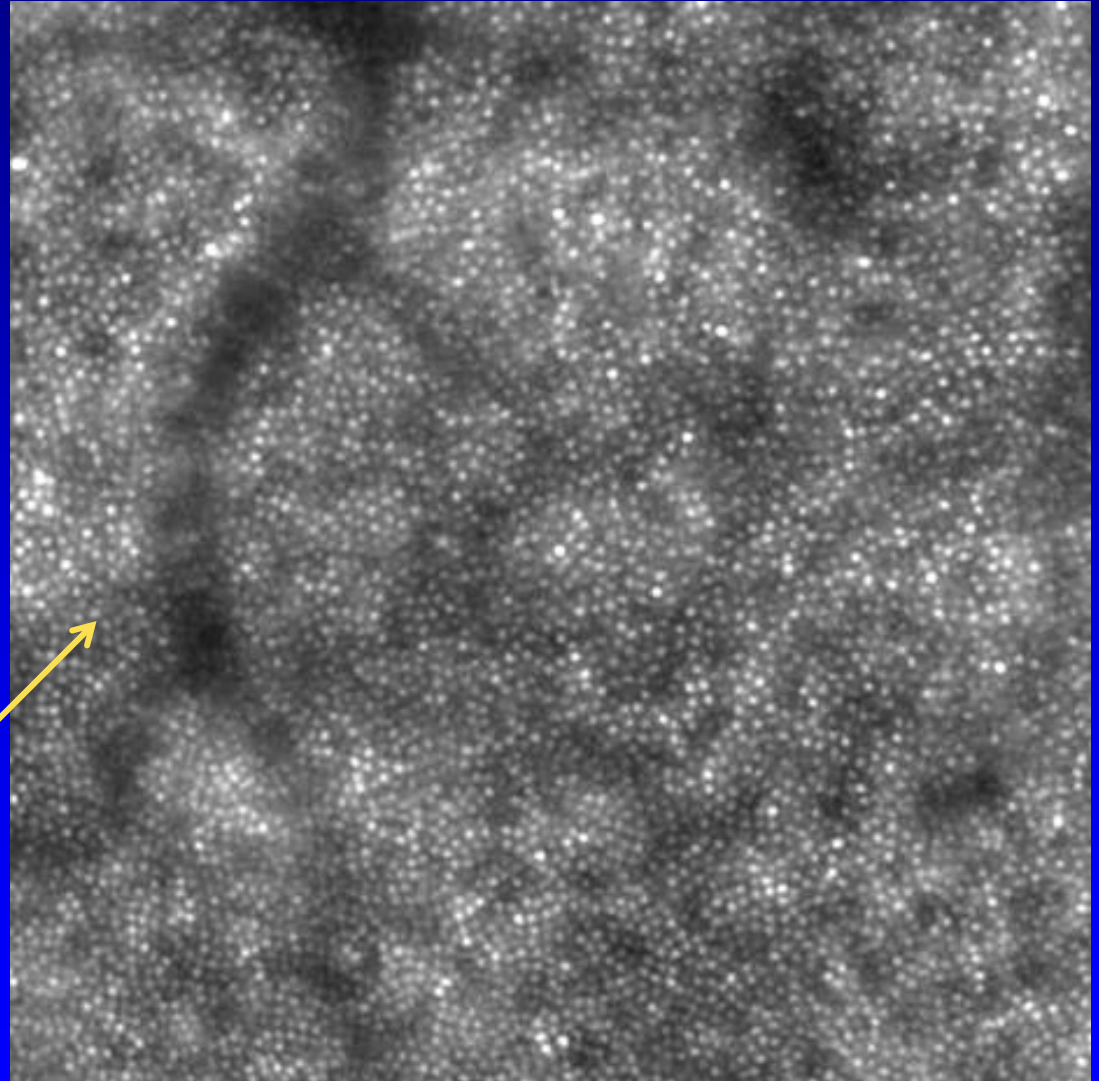
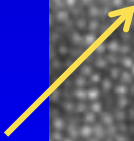
Overall average	Normal (28 Eyes)	AMD (12 Eyes)
Total retinal thickness(μm)	307.6 ± 12.8	271.3 ± 11.2
ORL thickness(μm)	194.5 ± 6.6	175.1 ± 10.1

Adaptive Optics



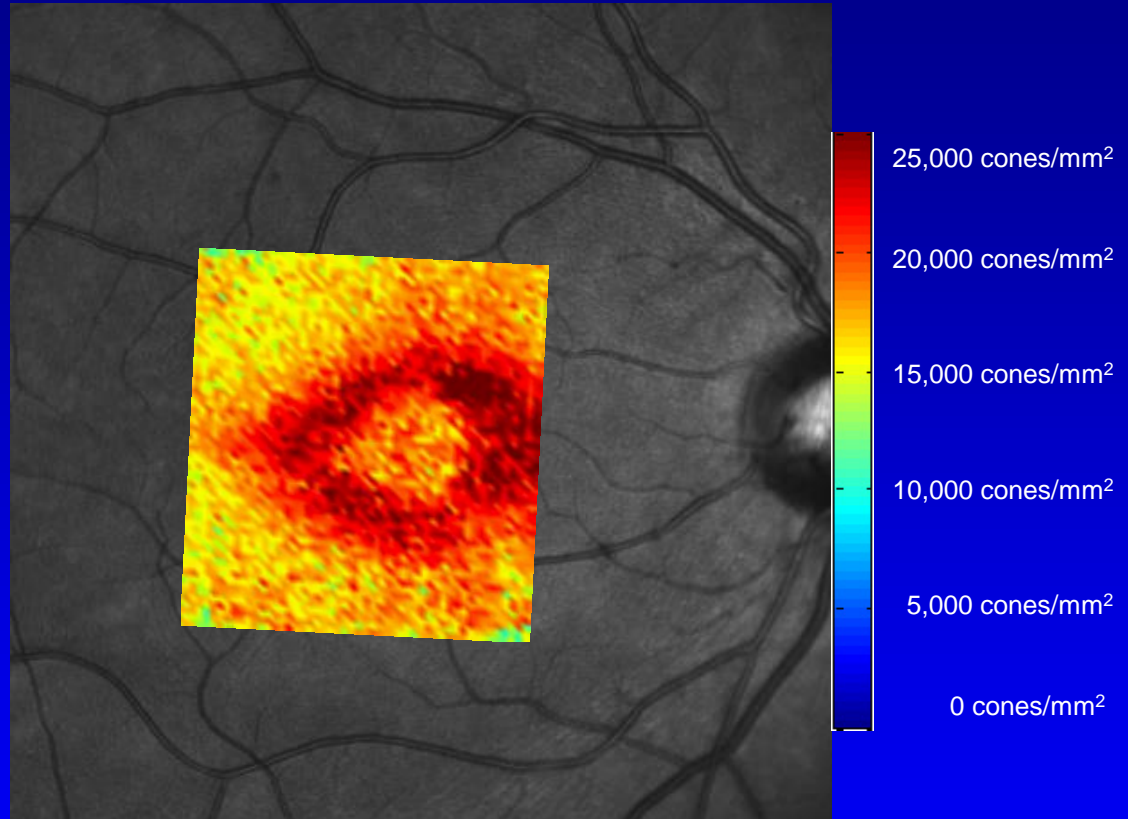
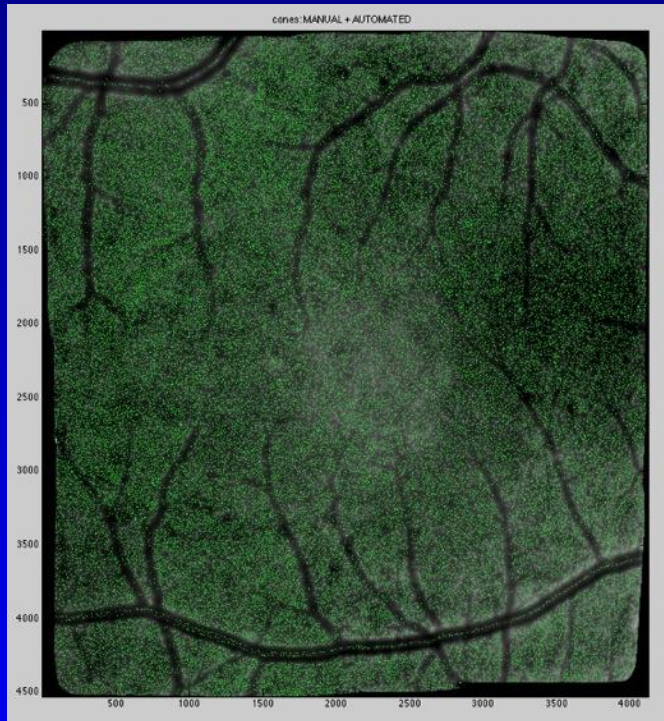
RTx1 from Imagine Eyes

Individual Cone Photoreceptors



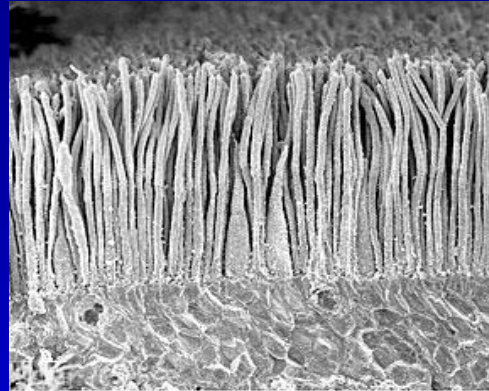
AO image demonstrating normal cone mosaic

Adaptive Optics

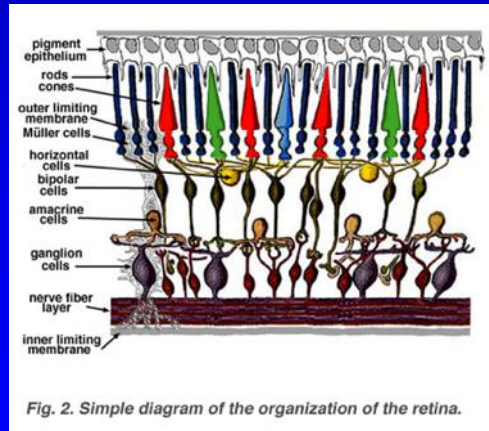
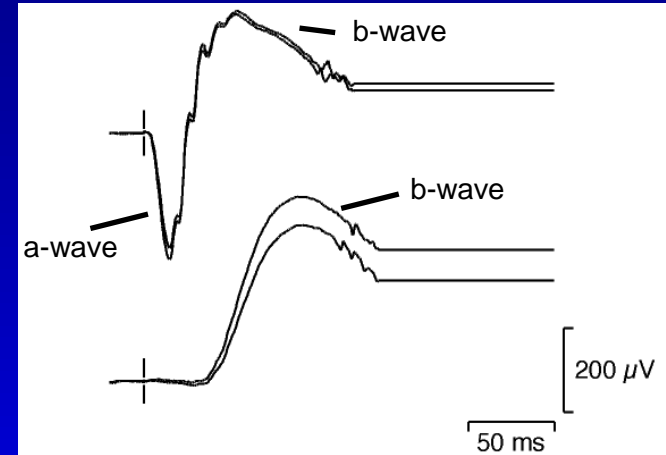


Cone Density Map

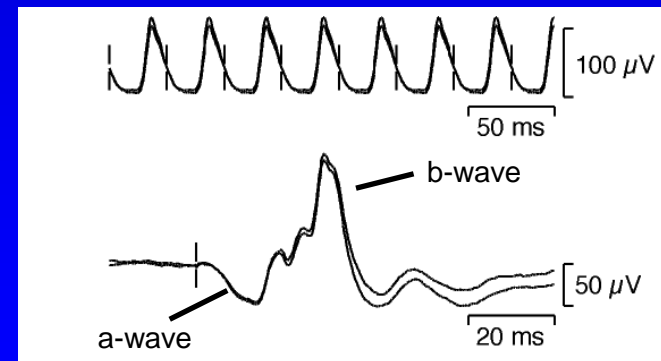
Full Field Electroretinograms (ERG)



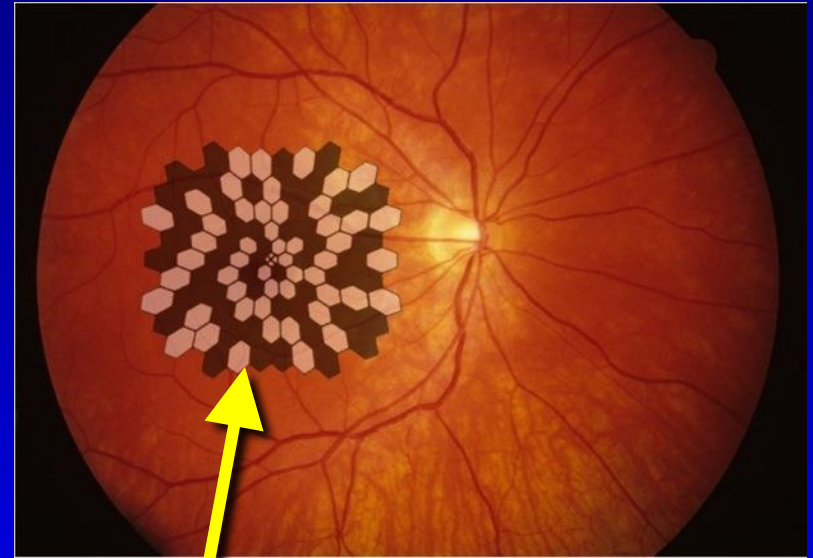
Rod-Driven ERG



Cone-Driven ERG

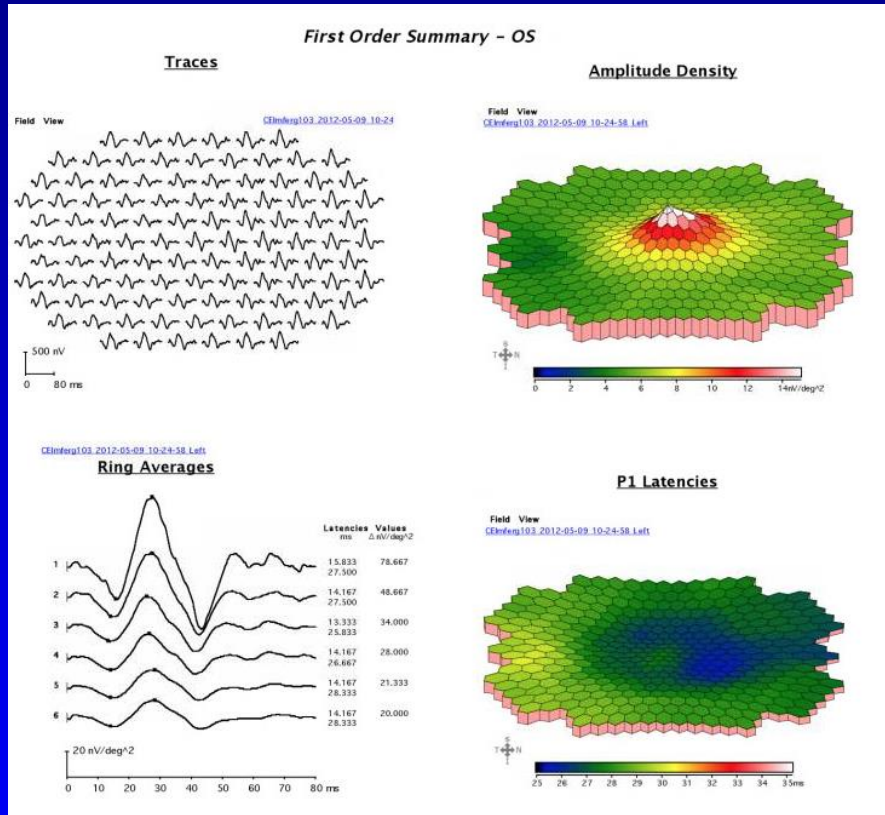


Multifocal Electroretinograms

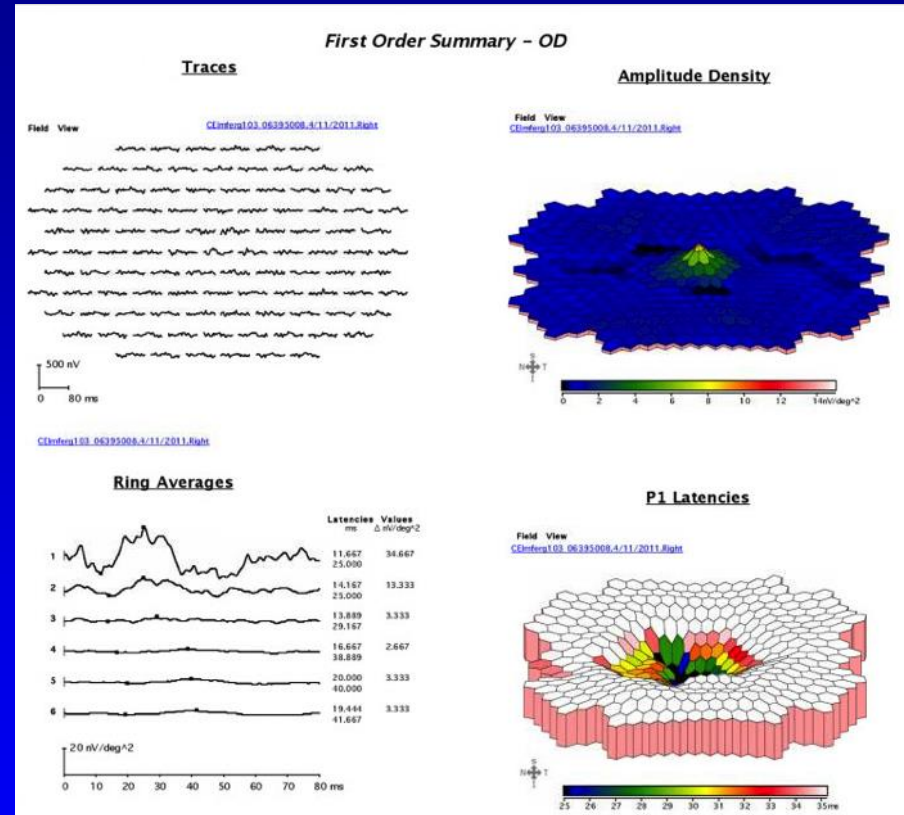


The mfERG only measures 24 deg!

Multifocal Electretinograms



Healthy Eye

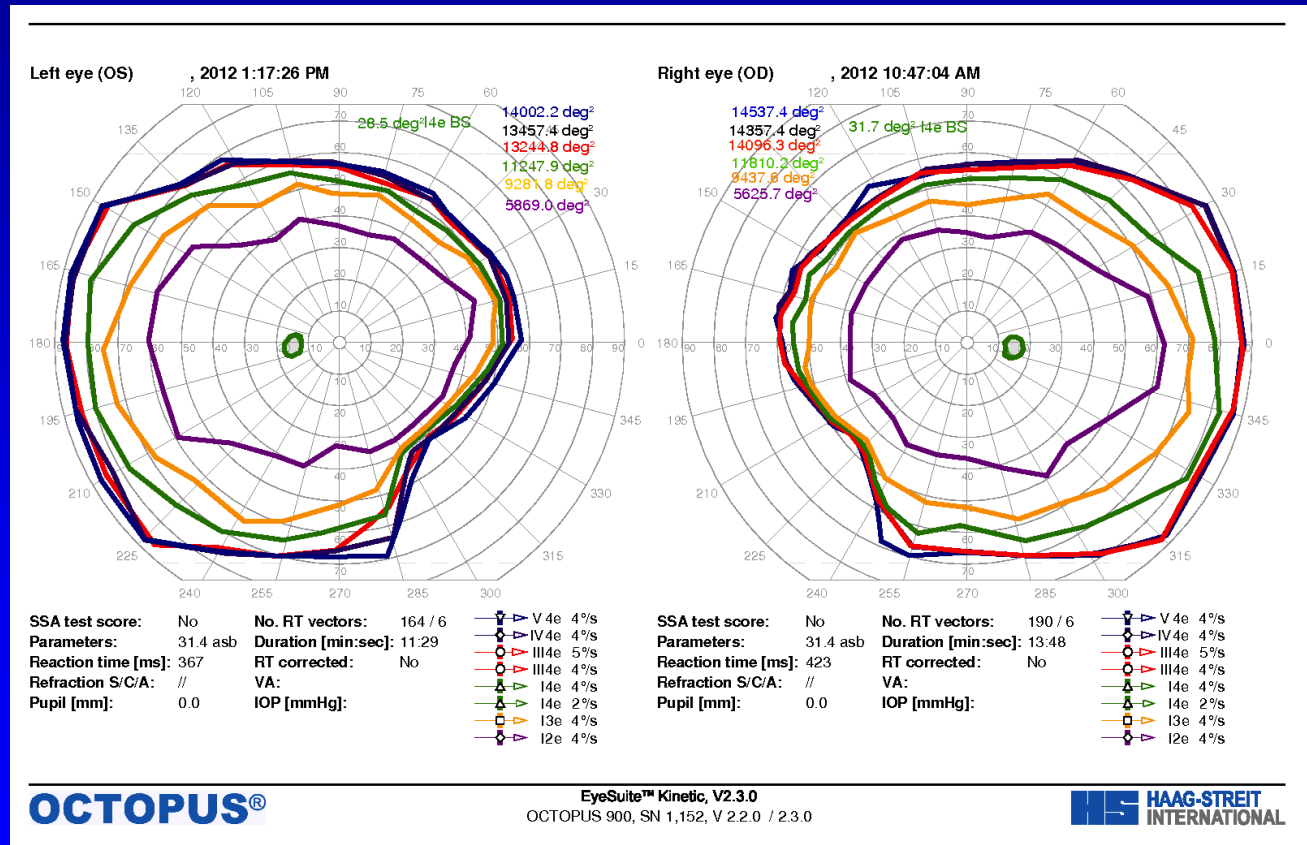


Usher Syndrome

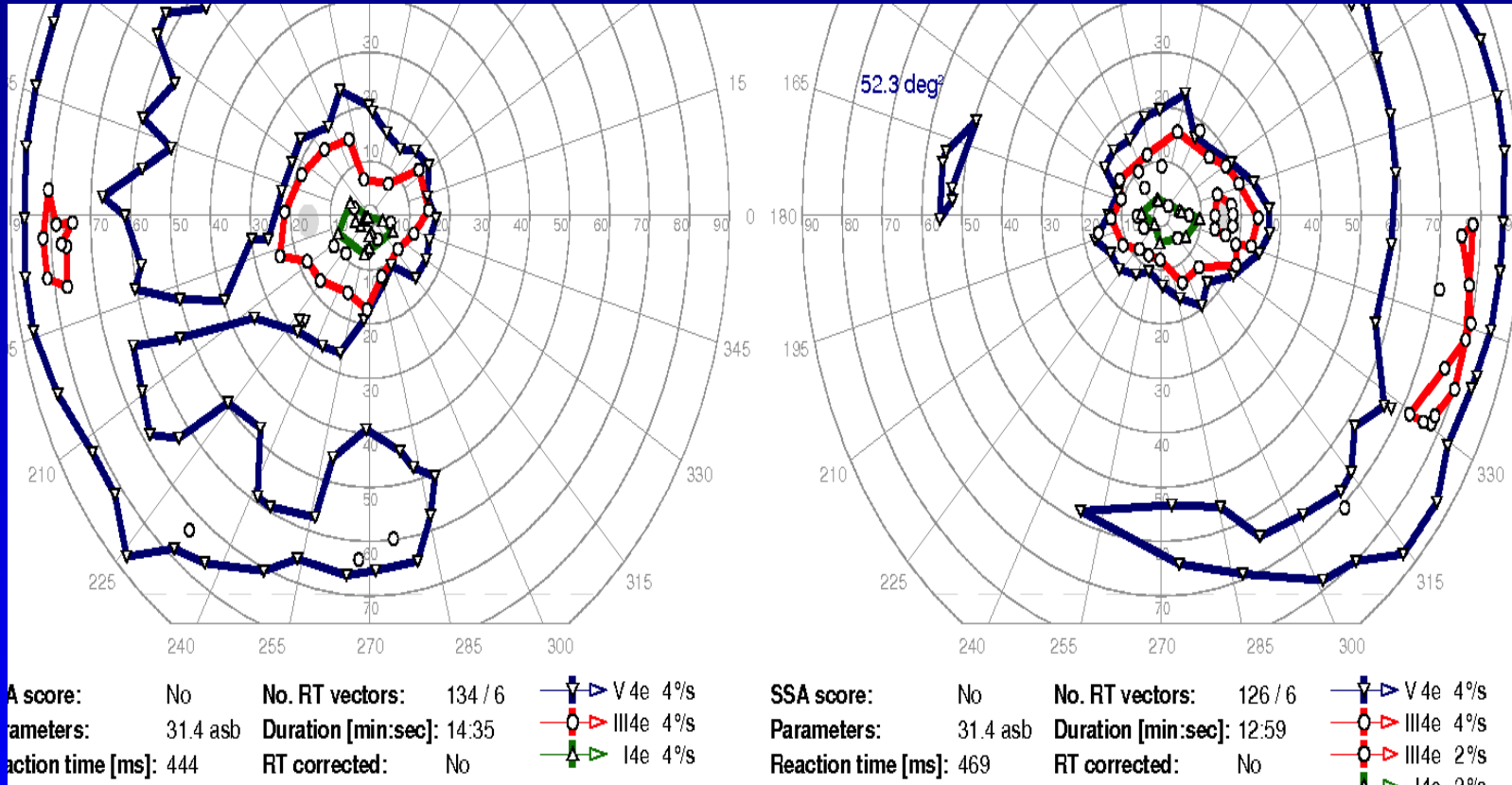
Octopus Visual Fields



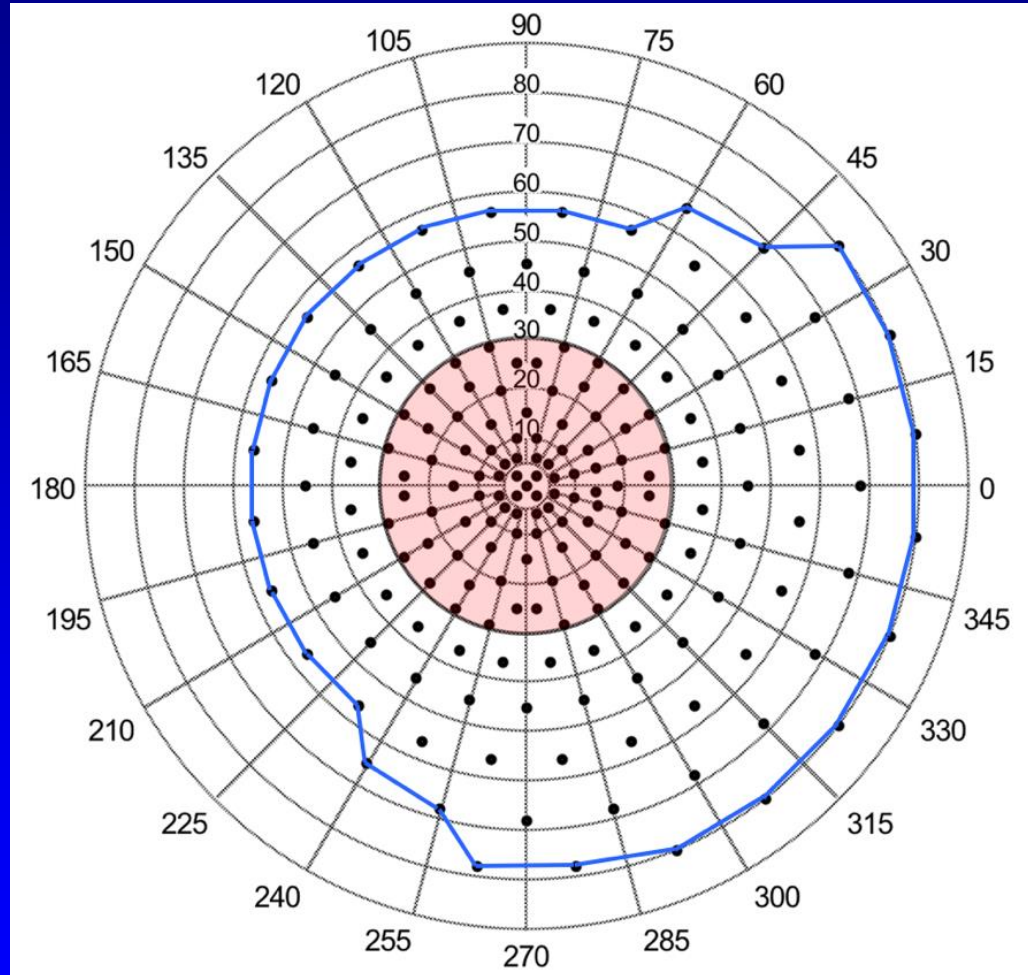
Kinetic Field



Kinetic Perimetry 24yr with USH1B



Faster GATE Algorithm allows 184 points to be sampled and cover entire field

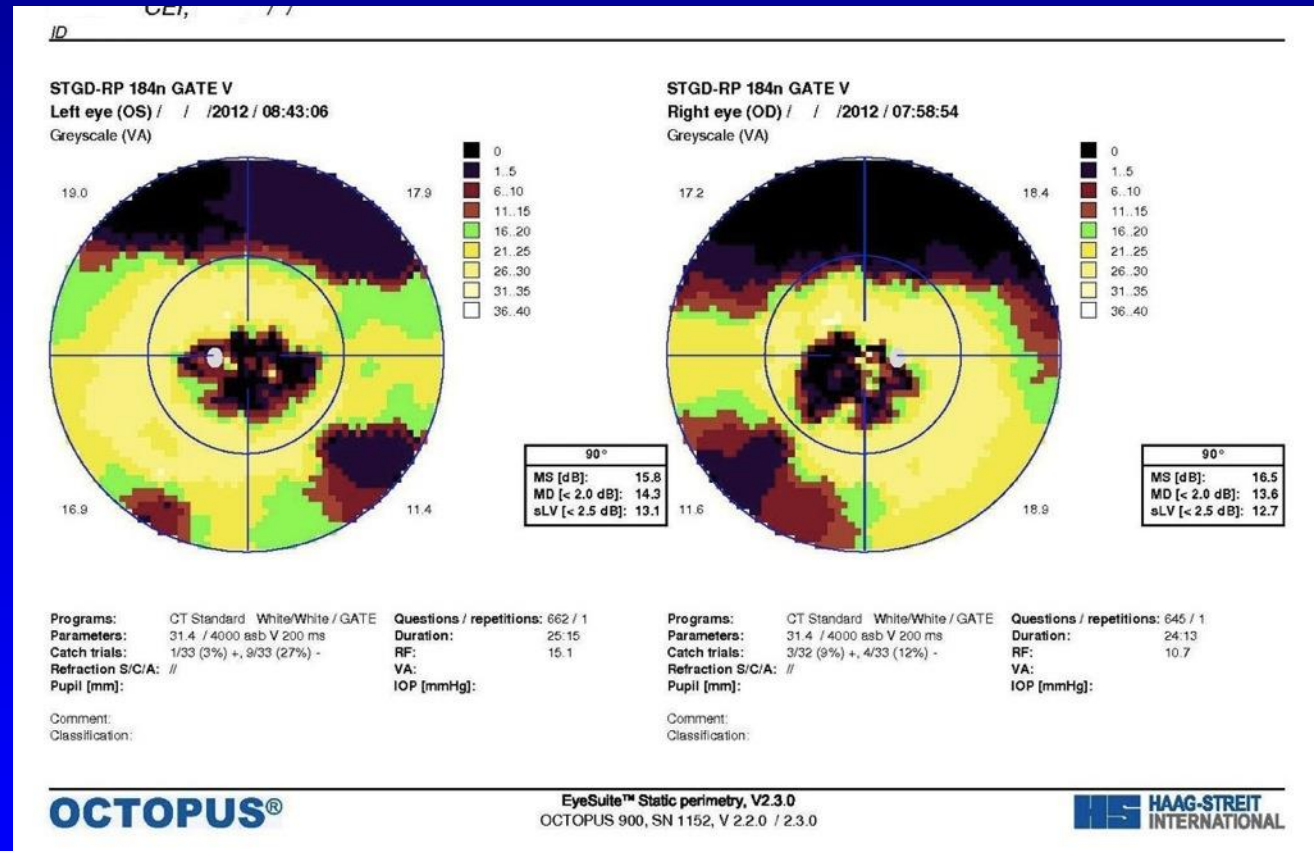


187 pt Grid

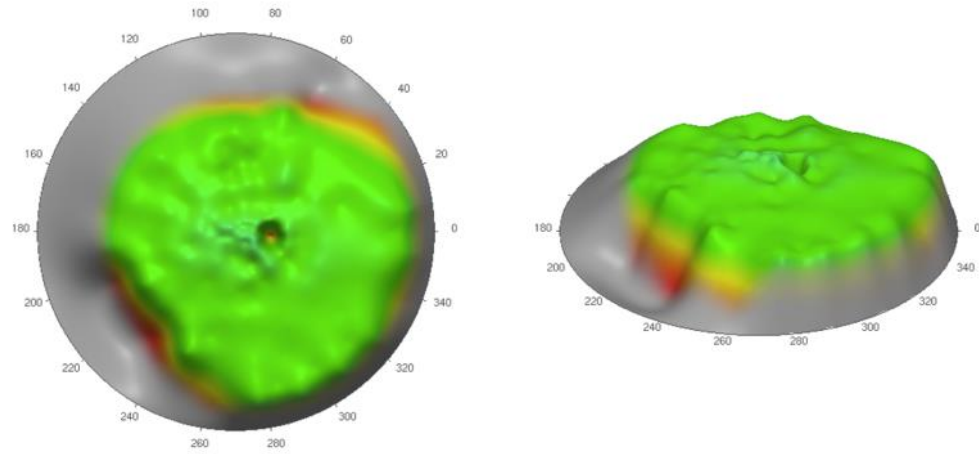
Octopus Visual Fields



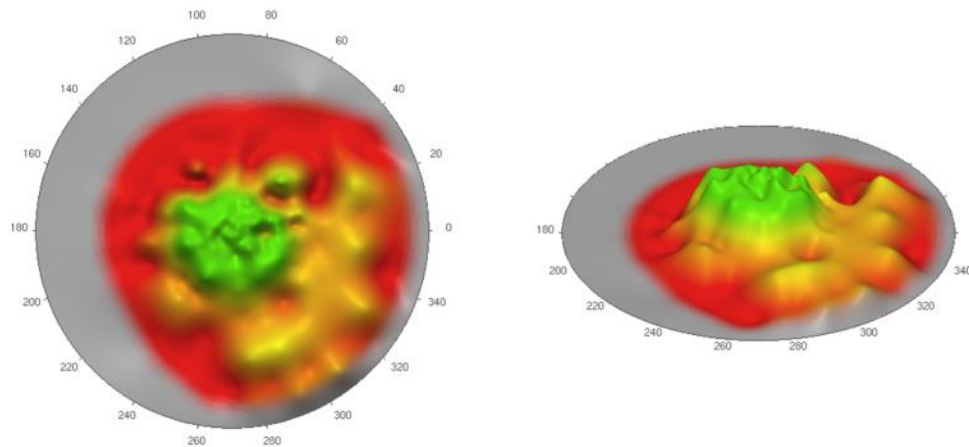
Abnormal Static Field



Visual Field Modeling in Usher Syndrome



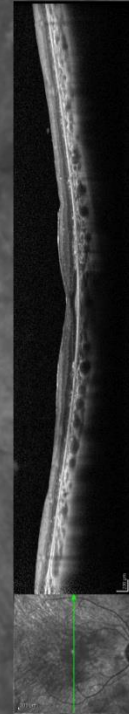
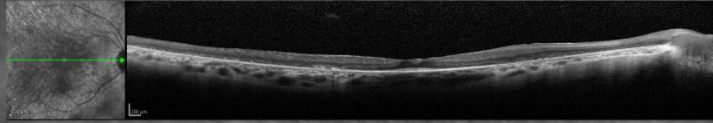
Normal

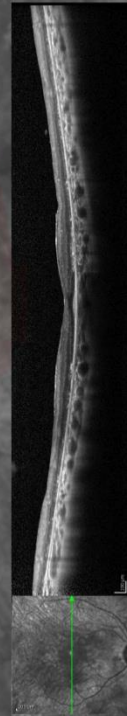
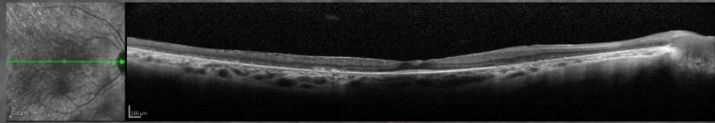
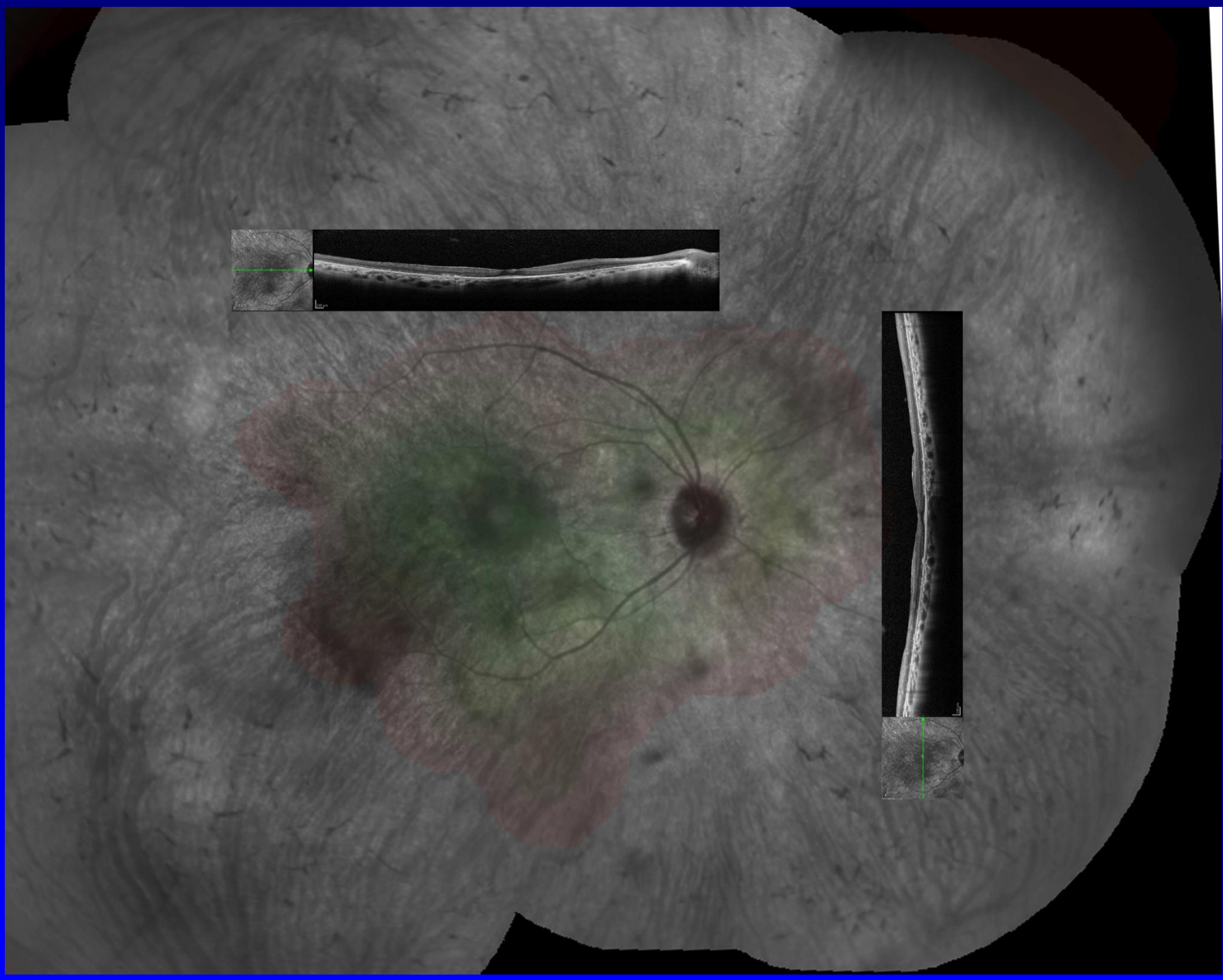


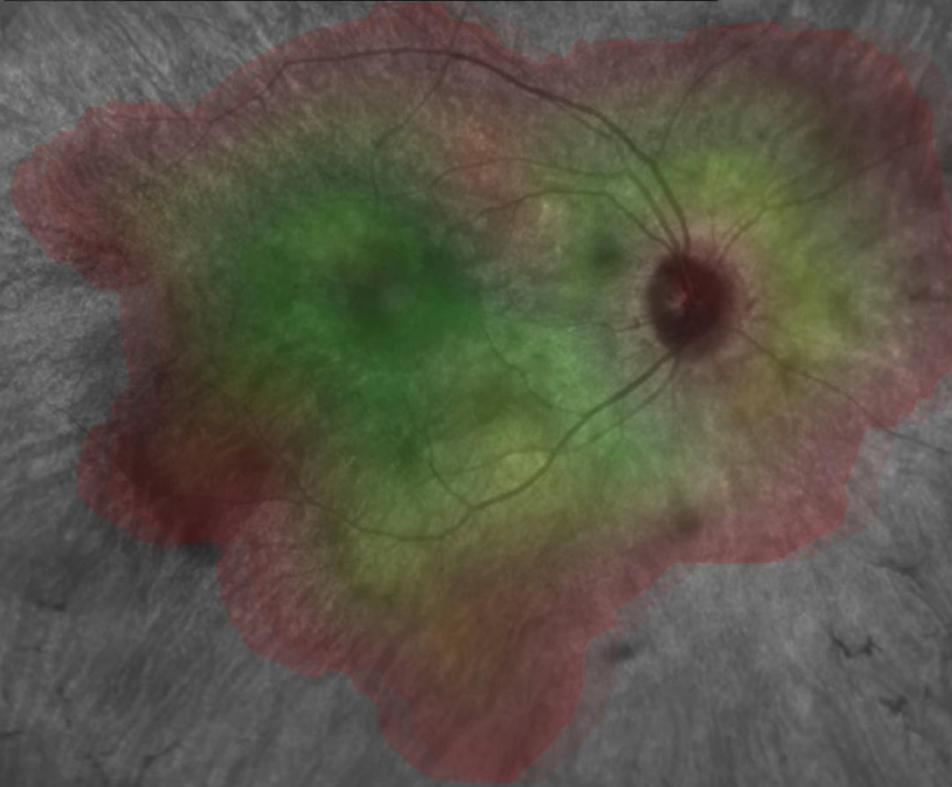
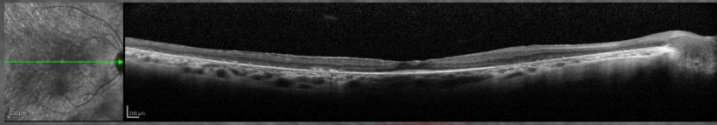
Usher Syndrome

Combining Structural and Functional Information

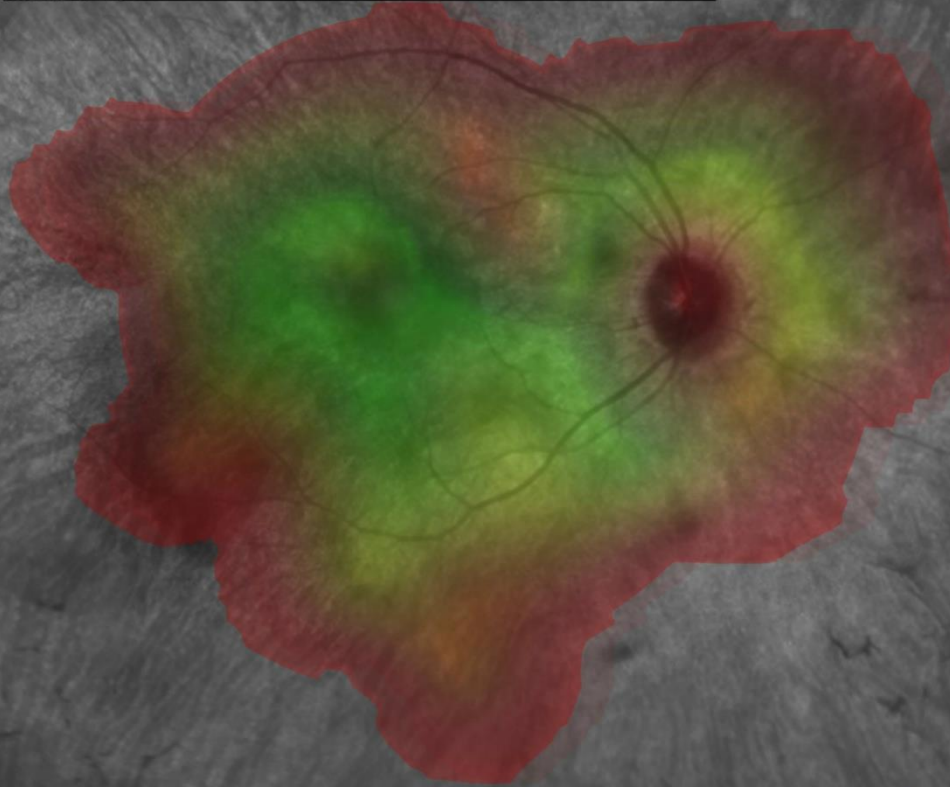
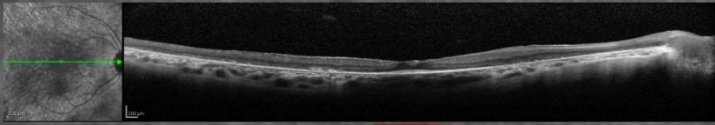
OD







[0.00/26.60]



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Treatments for Retinal Degenerations

- Low Vision Aids
- Micronutrients
- Neuroprotection/Small Molecules
- Transcript Editing (Antisense Oligonucleotides)
 - Cell Based Therapy (Stem Cells)
 - Optogenetics
 - Artificial Retina
 - Gene Therapy

Low Vision Aids



Proper glasses prescription



Night vision goggles



Broad-beam Flashlight



Magnifiers and minifiers



Tablet Devices

Micronutrients and Vitamins

- High dose Vitamin A

 - Precautions for taking high doses of vitamin A

 - Liver Toxicity
 - Osteoporosis
 - Pregnancy
 - contraindicating in some mutations (such as those seen in Stargardt Disease)

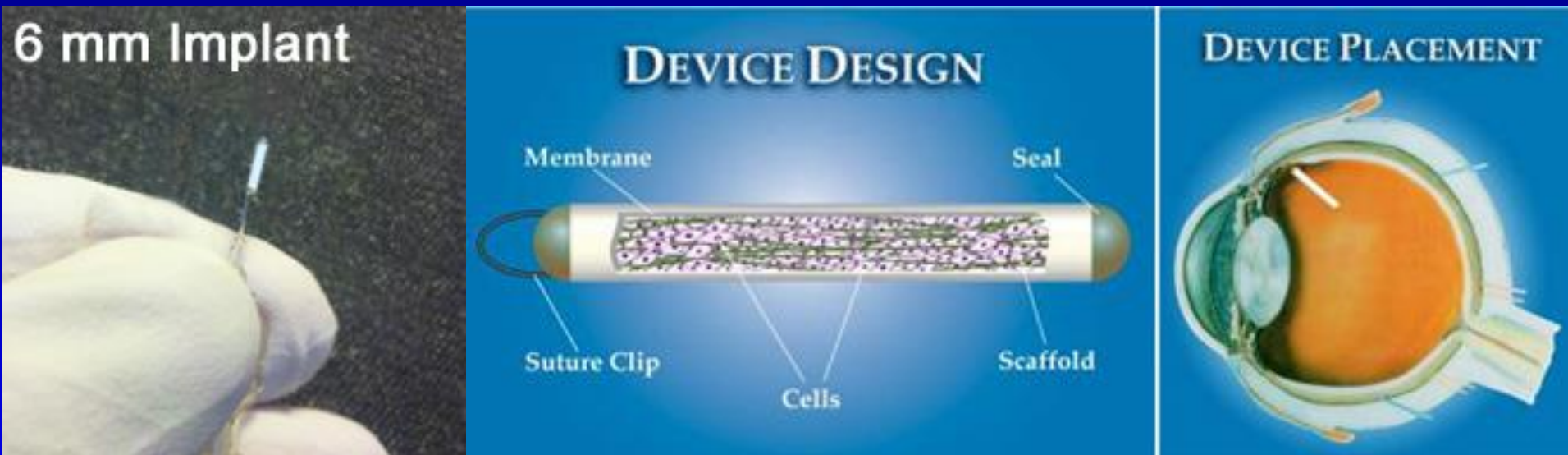
- DHA (docosahexanoic acid)

- Lutein

- ? β -carotene

- **AVOID:** Vitamin E, Smoking

Neuroprotection

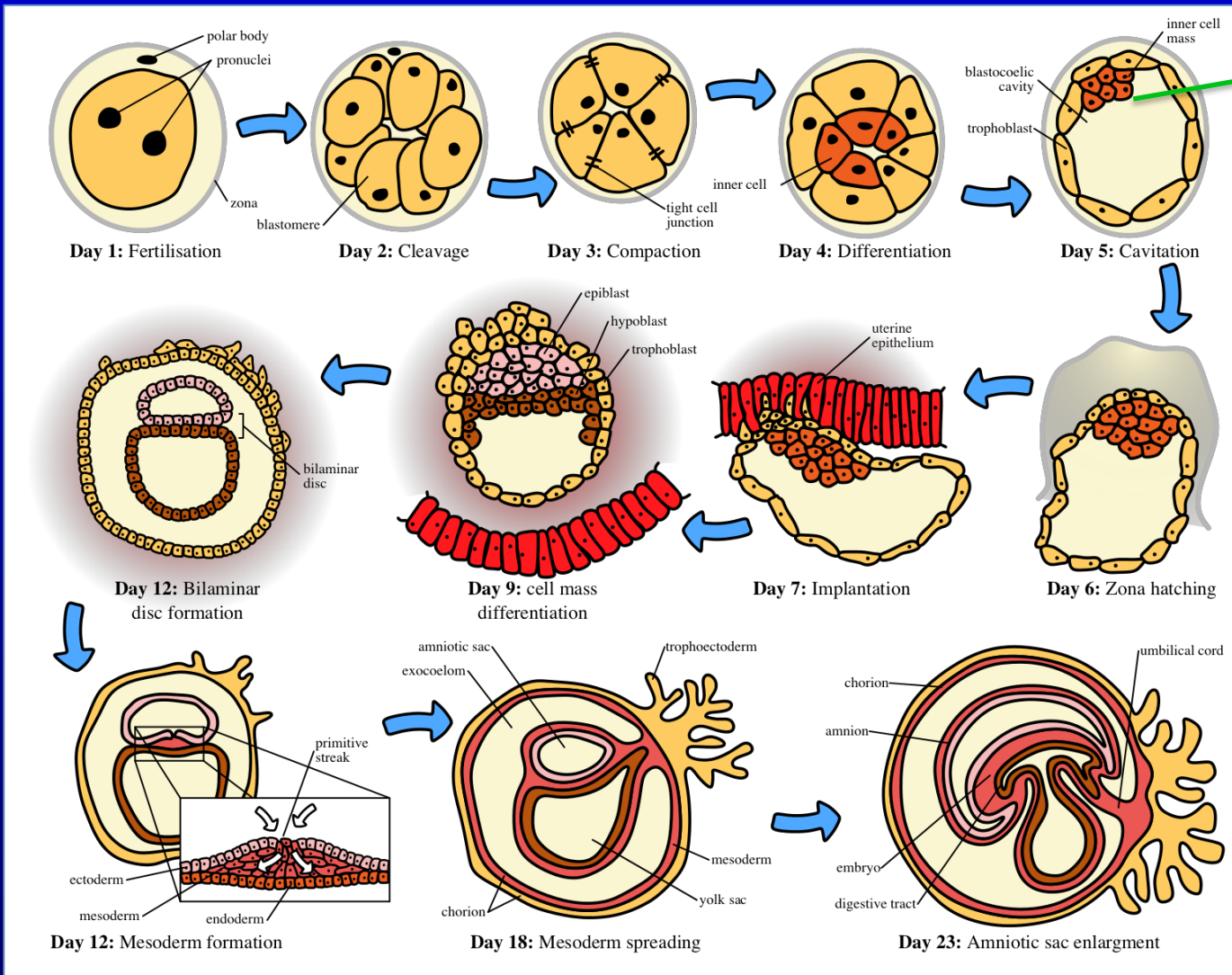


www.medgadget.com

Ciliary Neurotrophic Factor (CNTF)

Results of clinical trials have not been impressive

What is a Stem Cell?



Embryonic Stem (ES) Cells

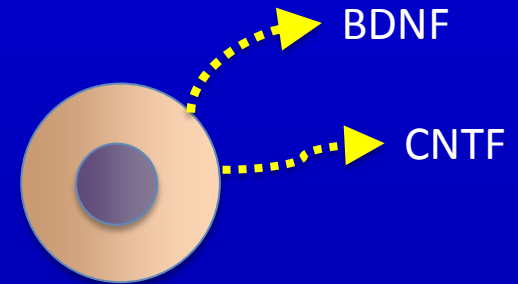
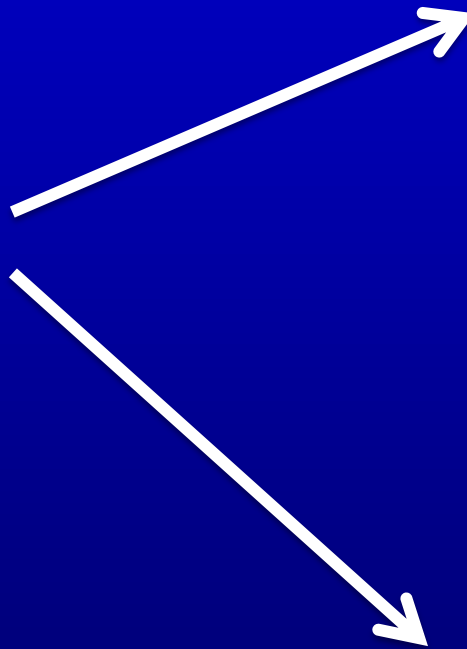
Totipotent {

- Unlimited capacity to divide
- Can become any other tissue

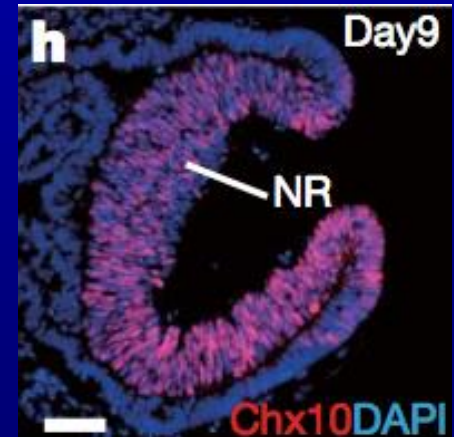
Pluripotent {

- Unlimited capacity to divide
- Can become a subset of tissues

How stem cells might work...



Release Protective Factors

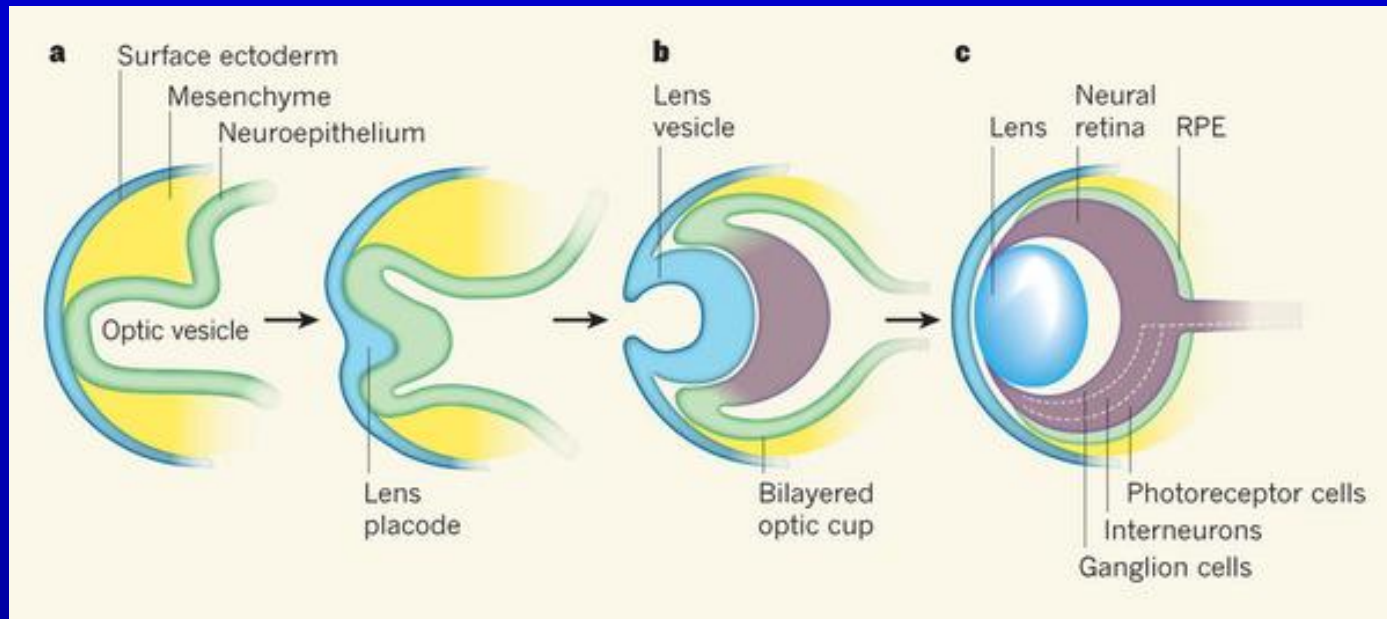


Differentiate in Retinal Tissue

Current Stem Cell Trials

<u>Sponsor</u>	<u>Cell Derivation</u>	<u>Differentiation</u>	<u>Delivery</u>	<u>Disease</u>	<u>Sites</u>
ACT	Embryonic	RPE	subretinal	Stargardt	UCLA, Bascom, Wills, MEEI, Moorfields, Korea
ACT	Embryonic	RPE	subretinal	AMD	UCLA, Bascom, Wills, MEEI, Korea
Pfizer	Embryonic	RPE	subretinal scaffold	AMD	UCL
Stem Cells Inc.	CNS		subretinal	AMD	RFSW
	Bone Marrow		intravitreal	RP, AMD	Univ. Sao Paulo
	Bone Marrow		intravitreal	RP	Madihol Univ (Thailand)
	Bone Marrow	CD34+	intravitreal	RP, DM, AMD	UC Davis

How to build a new retina



Self-Formation of Optic Cups and Storable Stratified Neural Retina from Human ESCs

Tokushige Nakano,^{1,2,4,5} Satoshi Ando,^{1,2,4} Nozomu Takata,¹ Masako Kawada,¹ Keiko Muguruma,¹ Kiyotoshi Sekiguchi,¹ Koichi Saito,⁴ Shigenobu Yonemura,³ Mototsugu Eiraku,^{1,2} and Yoshiki Sasai^{1,2,5,*}

¹Organogenesis and Neurogenesis Group

²Division of Human Stem Cell Technology

³Electron Microscopy Laboratory

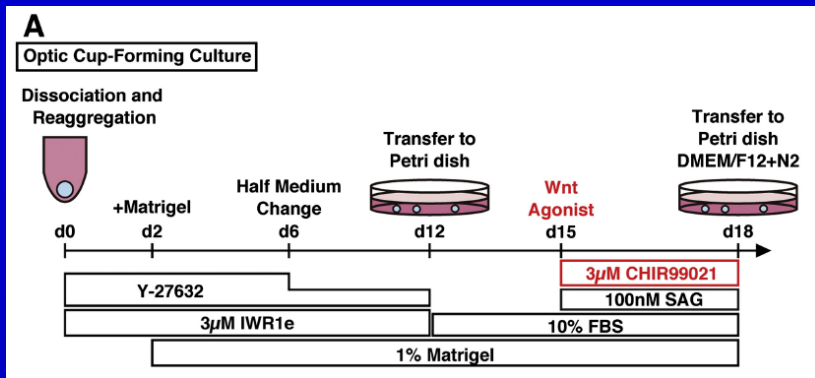
RIKEN Center for Developmental Biology, Kobe 650-0047, Japan

⁴Environmental Health Science Laboratory, Sumitomo Chemical Co., Ltd., Osaka 554-8558, Japan

⁵Department of Medical Embryology, Graduate School of Medicine, Kyoto University, Kyoto 606-8501, Japan

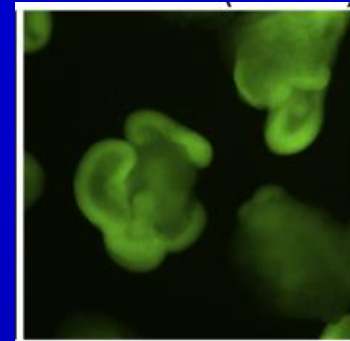
⁶Laboratory of Extracellular Matrix Biochemistry, Institute for Protein Research, Osaka University, Suita 565-0871, Japan

*Correspondence: yoshikisasai@cdb.riken.jp

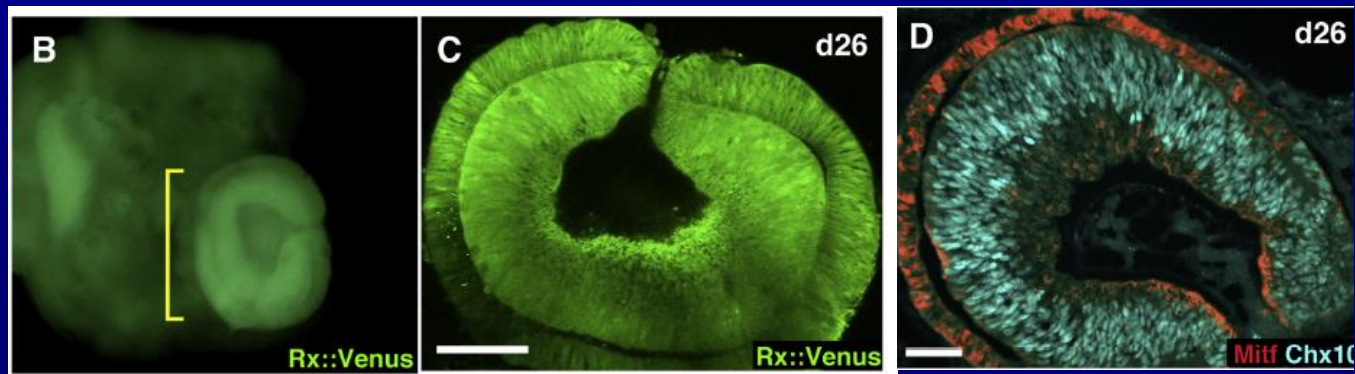
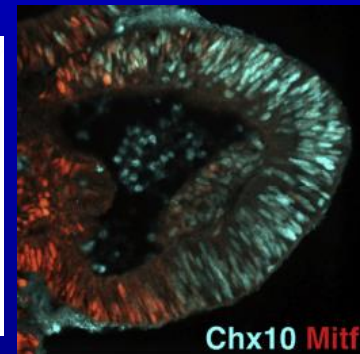
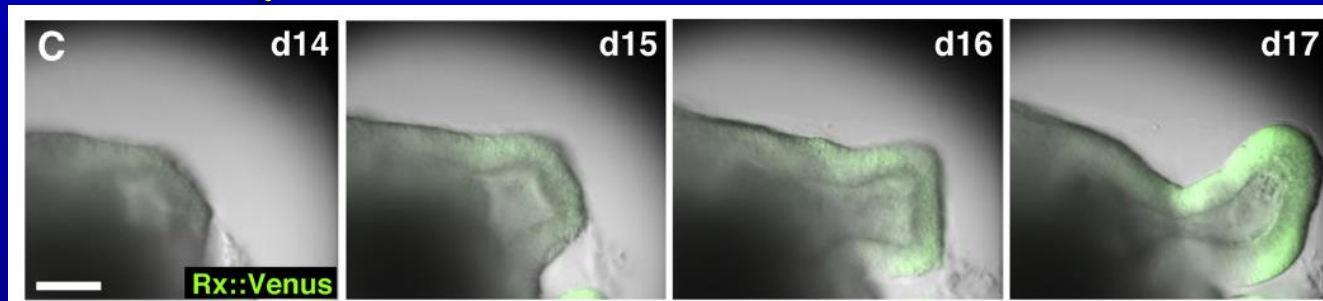


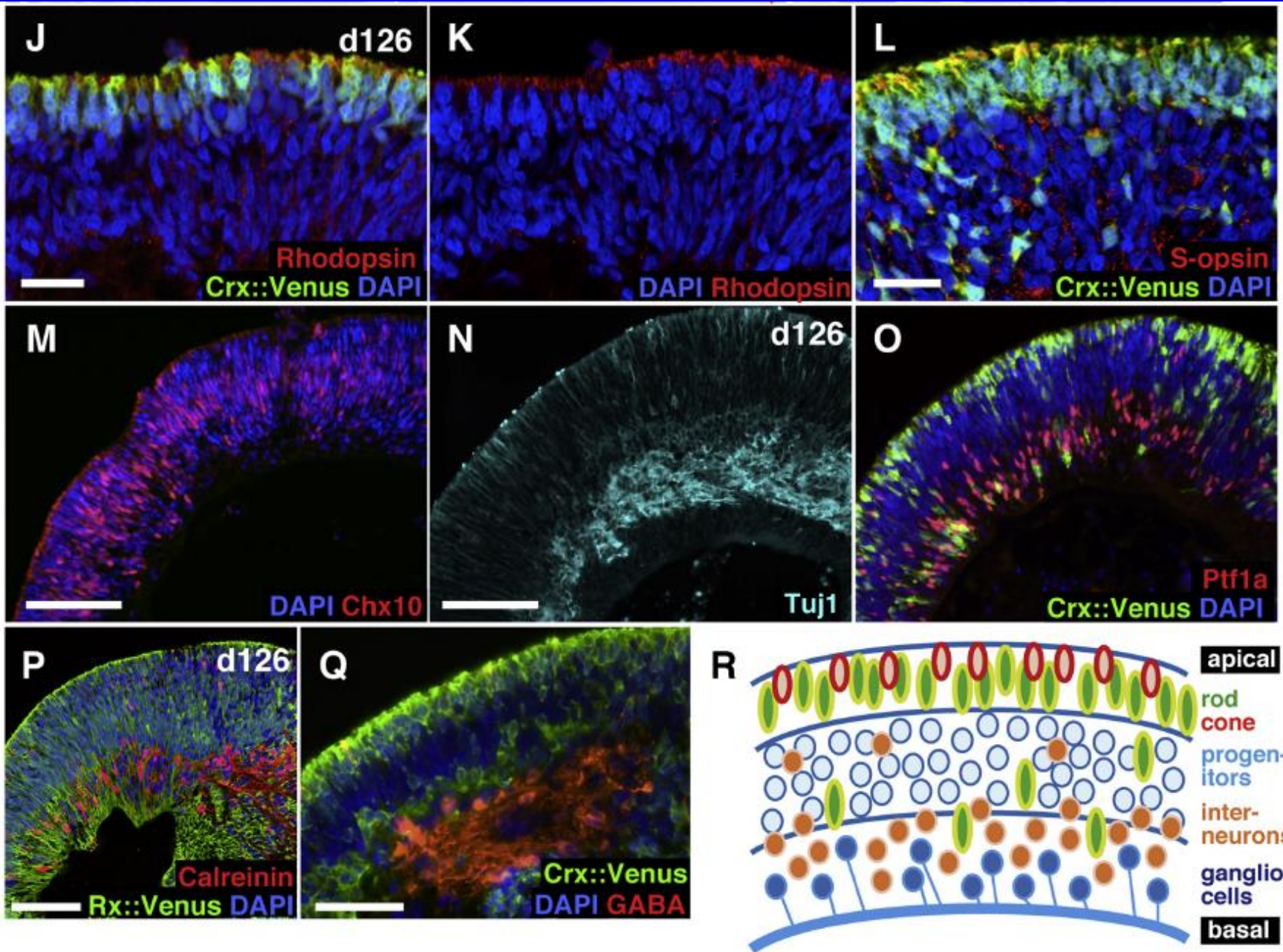
hESC grown in 3D Matrigel with SFEBq media

Transfect venus under RX promoter



FACS for RX:Venus positive cells

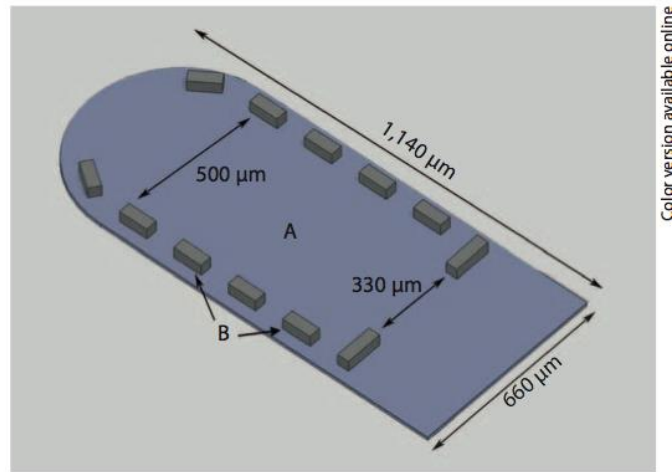




A Novel Approach for Subretinal Implantation of Ultrathin Substrates Containing Stem Cell-Derived Retinal Pigment Epithelium Monolayer

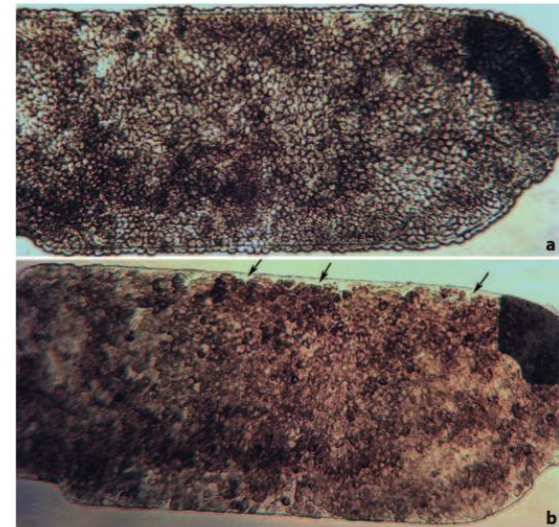
Yuntao Hu^{a,e} Laura Liu^{a,f} Bo Lu^c Danhong Zhu^{a,b} Ramiro Ribeiro^{a,g}
Bruno Diniz^{a,h} Padmaja B. Thomas^a Ashish K. Ahuja^a David R. Hinton^{a,b}
Yu-Chong Tai^c Sherry T. Hikita^d Lincoln V. Johnson^d Dennis O. Clegg^d
Biju B. Thomas^a Mark S. Humayun^a

Ophthalmic Res 2012;48:186–191



Color version available online

Fig. 1. Diagrammatic sketch of the implantation tool. The device consists of a thin parylene plate (A) containing barriers (B) arranged in the form of a 'U'. The substrate for implantation is placed in the middle of the U-shaped area (substrate chamber).



Color version available online

Fig. 2. The ultrathin substrate containing hESC-RPE cells. Images were taken before (a) and after (b) subretinal implantation. b Considerable cell loss can be observed along the edges of the substrate (arrows).

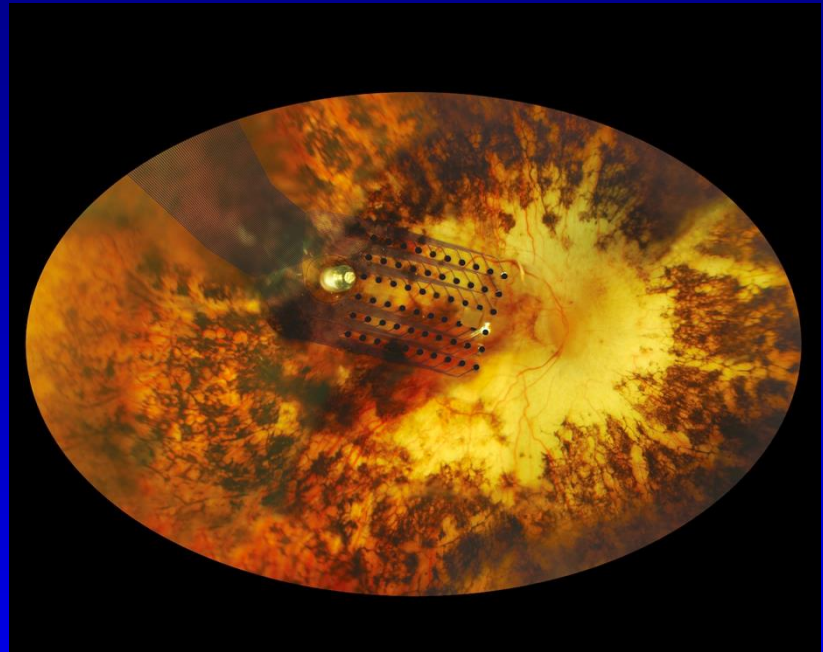
4- μ m-thick parylene substrates containing a monolayer of human embryonic stem cell-derived RPE

Implantable Retinal Silicon Chips



AP Photo/Martin Cleaver

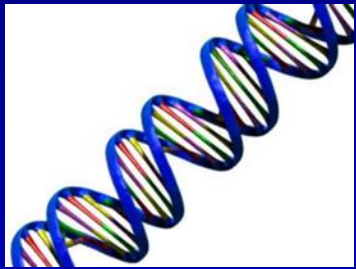
Argus II Retinal Implant



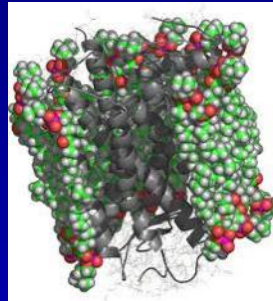
Gene Therapy

What is gene therapy?

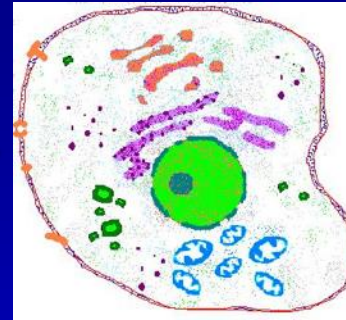
Normal



DNA

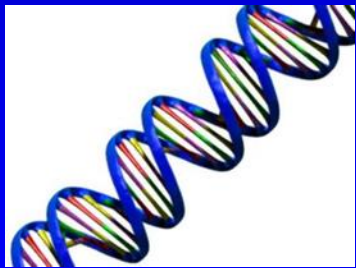


Protein

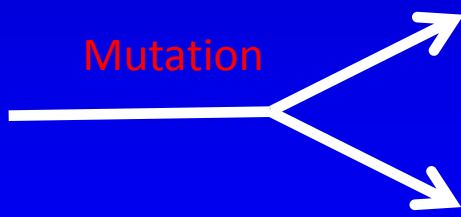


Cells

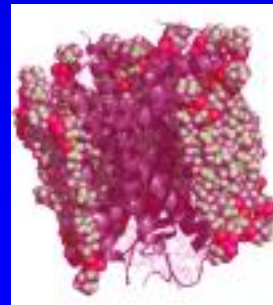
Abnormal



DNA



No Protein



Mutant Protein

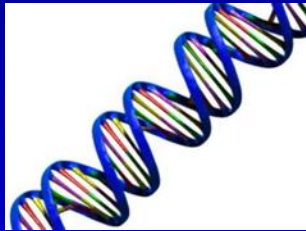
Gene Therapy Approach

Replace missing protein

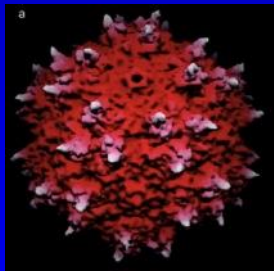
Decrease bad protein

How to Deliver Gene Therapy?

- **Vector** – a mechanism to deliver DNA to a cell



Bare DNA

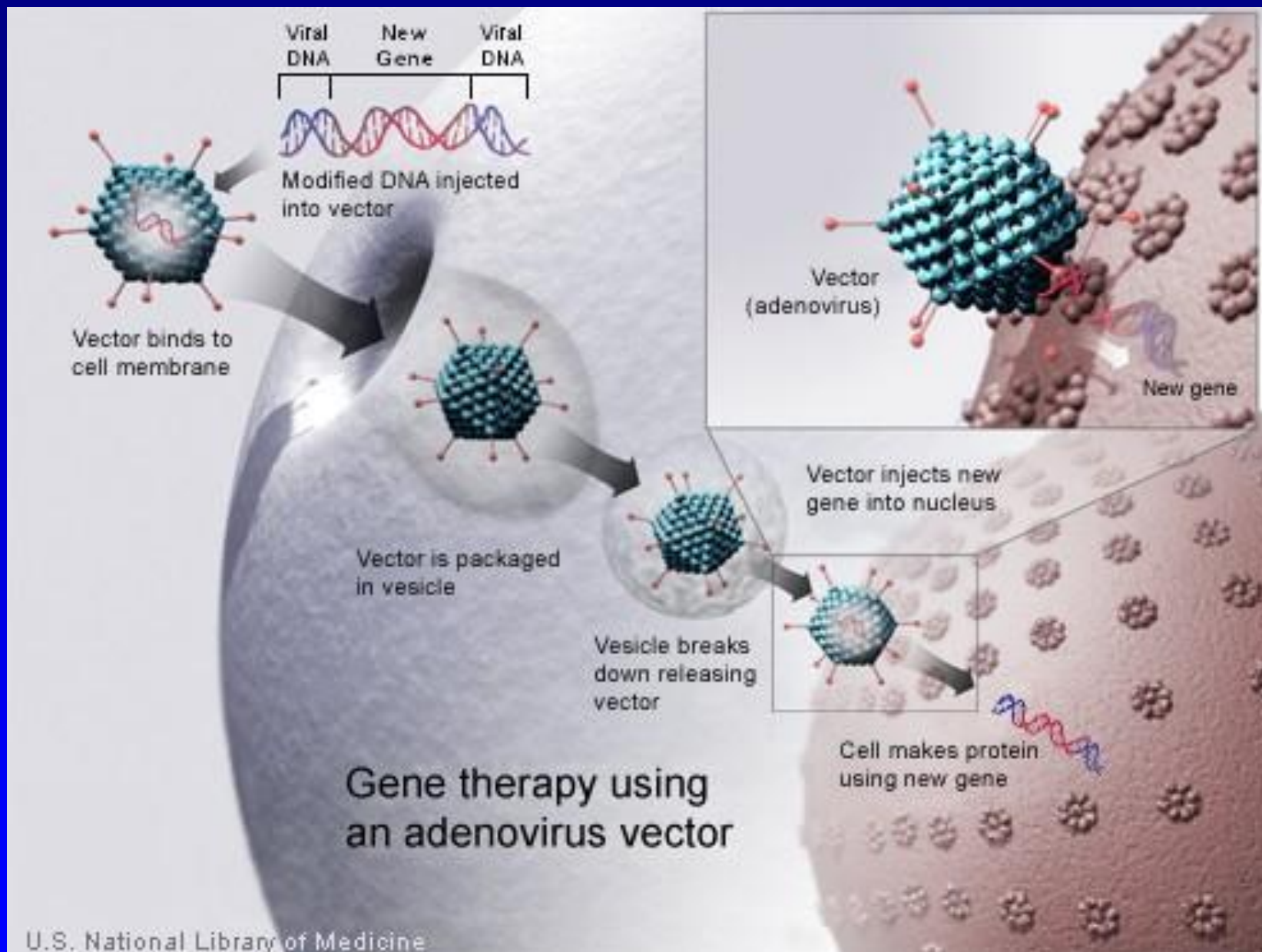


Modified Virus

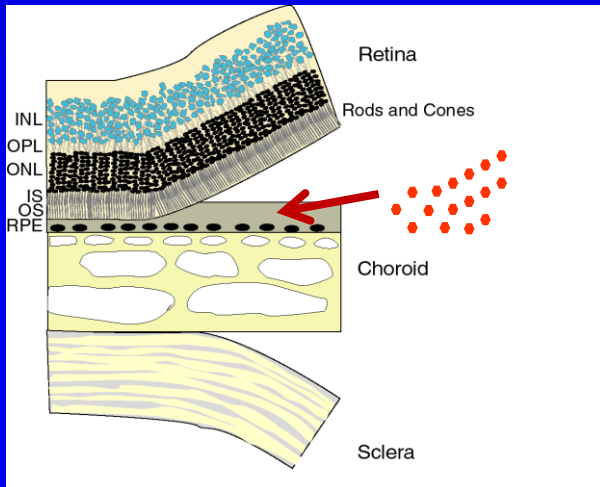
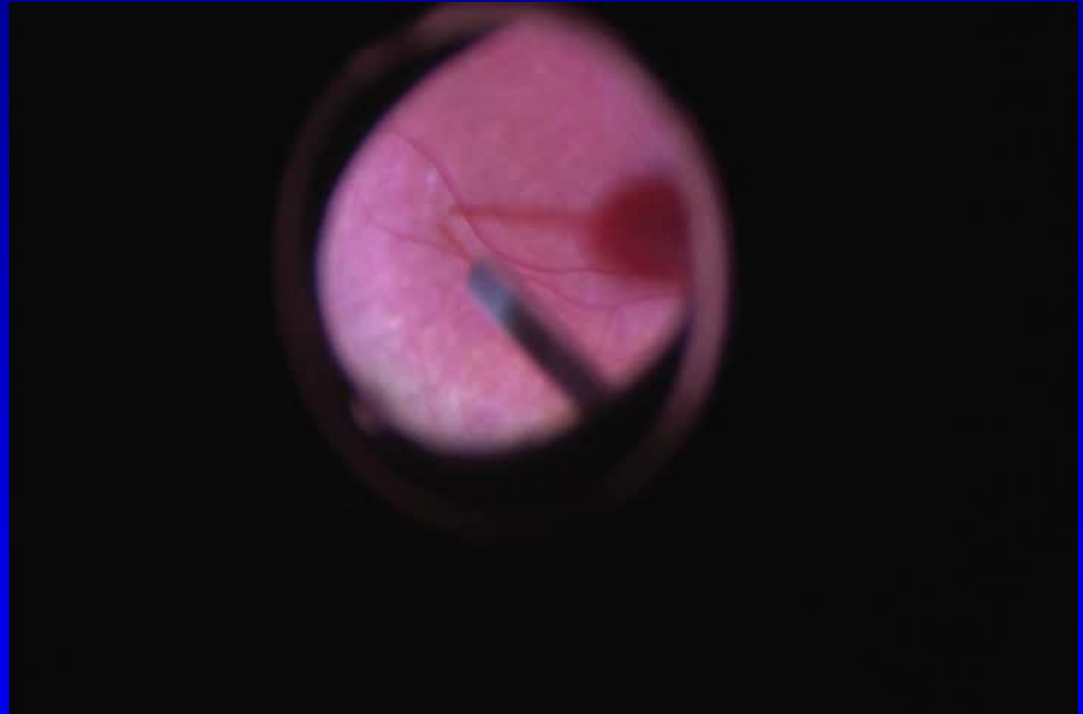
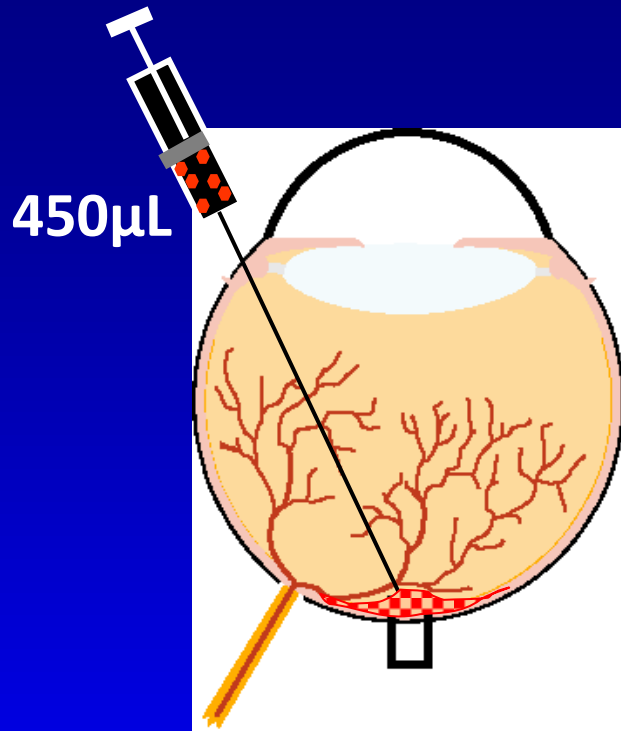


Nanoparticles

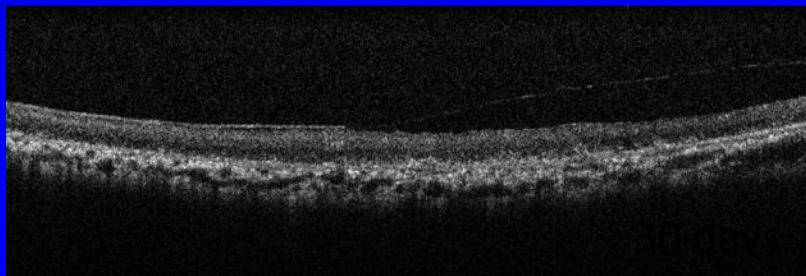
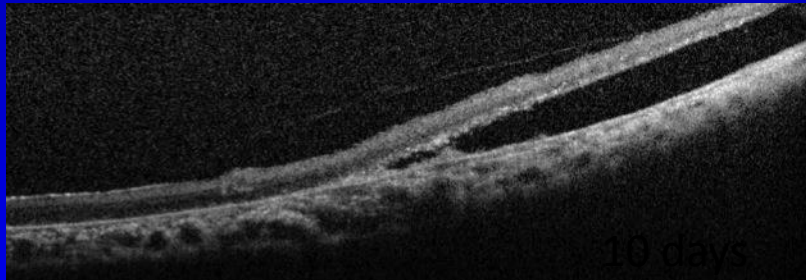
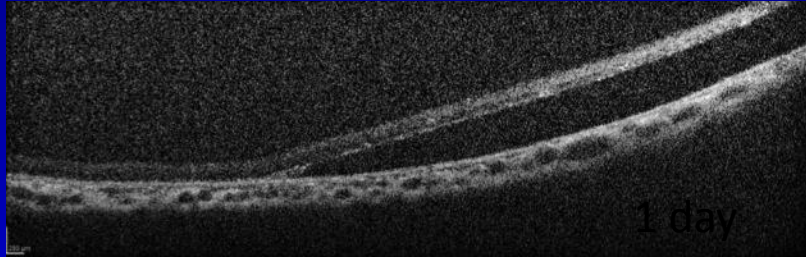
Gene Therapy



Gene Therapy - Subretinal Injection



Recovery of Subretinal Bleb



- Immediately after injection

- Bleb resorbed

Gene Therapy Trials at Casey Eye Institute

Current

- Leber Congenital Amaurosis Type 2 (*RPE65*)
- Stargardt Disease (*ABCA4*)
- Usher Syndrome Type 1B (*MYO7A*)
- Retinostat (Endostatin/Angiostatin for NVAMD)

Future/Planned Trials

- X-Linked Retinoschisis (*RS1*)
- Achromatopsia (*CNGB3*)

Gene Therapy Trials at Other Centers

- LCA (*RPE65*) – University of Pennsylvania, CHOP, Moorfields, Israel
- Choroideremia (*REP1*) – University of Oxford
- Retinitis Pigmentosa (*MERTK*) – King Khalid Eye Specialist Hospital

UshStat

(For Type 1B Usher Syndrome)

Type 1B Usher Syndrome

Genetic Defect

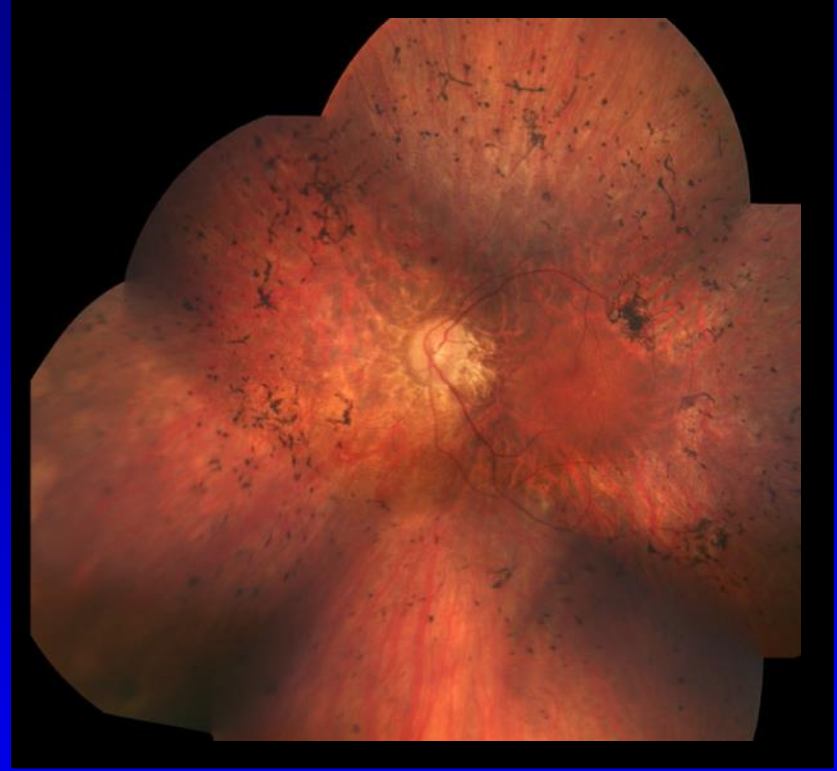
- Autosomal Recessive
- Mutations in *MYO7A*

Clinical Features

- Severe Early Onset Rod-cone dystrophy
- Severe Congenital Deafness
- Balance problems



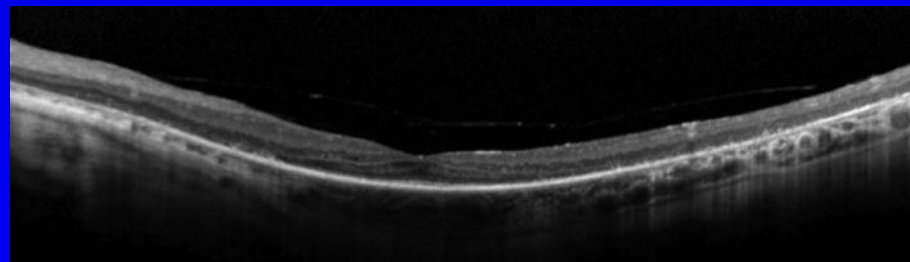
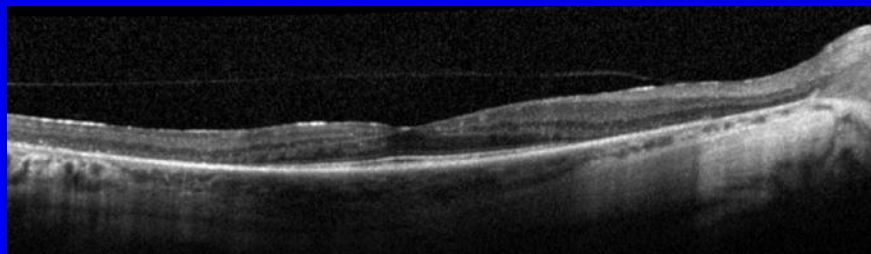
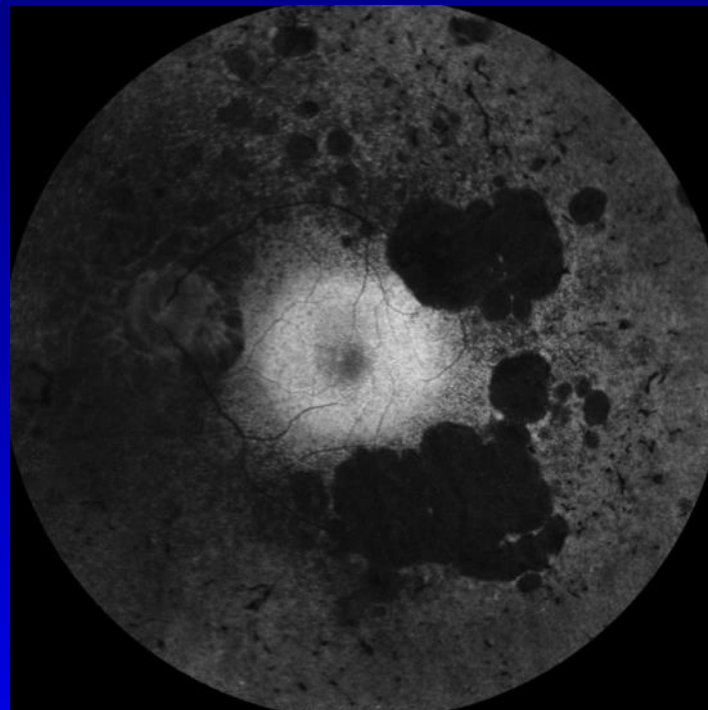
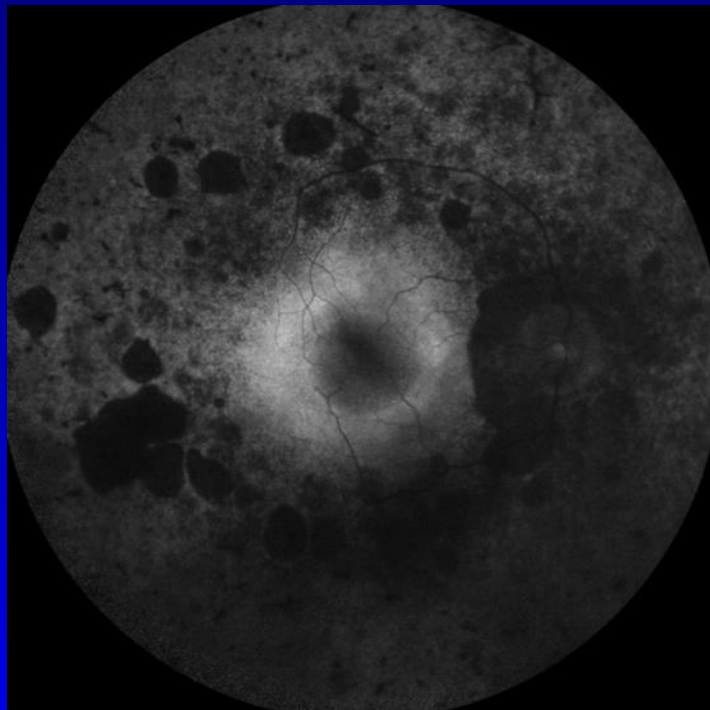
Usher Syndrome – Type 1B



61 yr Female, Va = 20/40 OU

Severe Deafness

Usher Syndrome – Type 1B



61 yr Female, Va = 20/40 OU

A Phase I/IIa Dose Escalation Safety Study of Subretinally Injected USHStat™, Administered to Patients with Usher Syndrome Type 1B

Sponsor: Oxford Biomedica UK / Sanofi

Primary Investigator: Richard Weleber MD

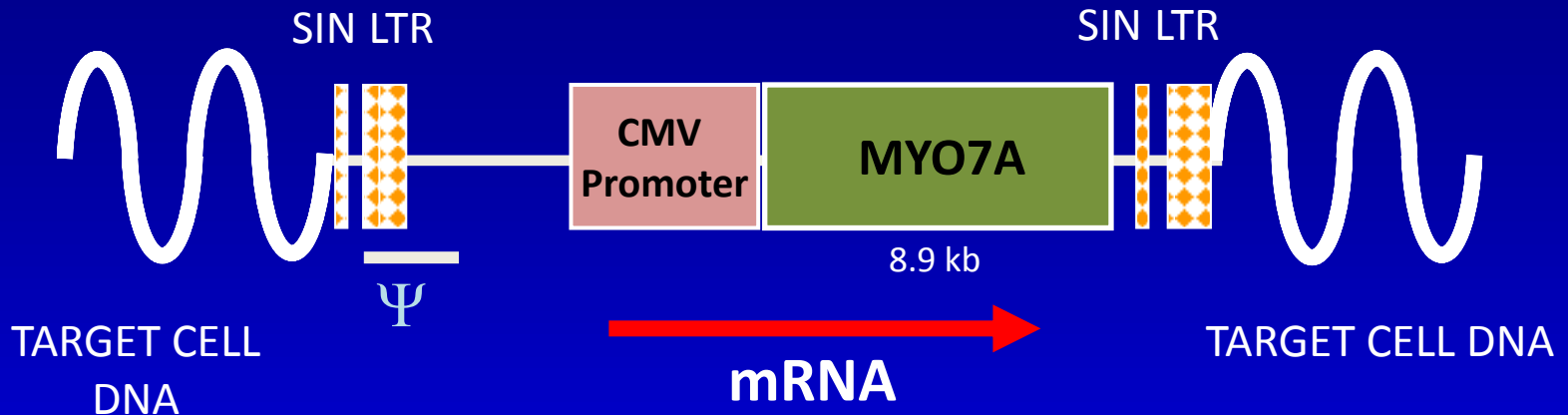
Design: Phase I/IIa dose escalation study

Sites: Casey Eye Institute, OHSU
Hospitalier Nationale d'Ophthalmologie des Quinze-Vingts

Vector: non-primate lentiviral vector based on EIAV

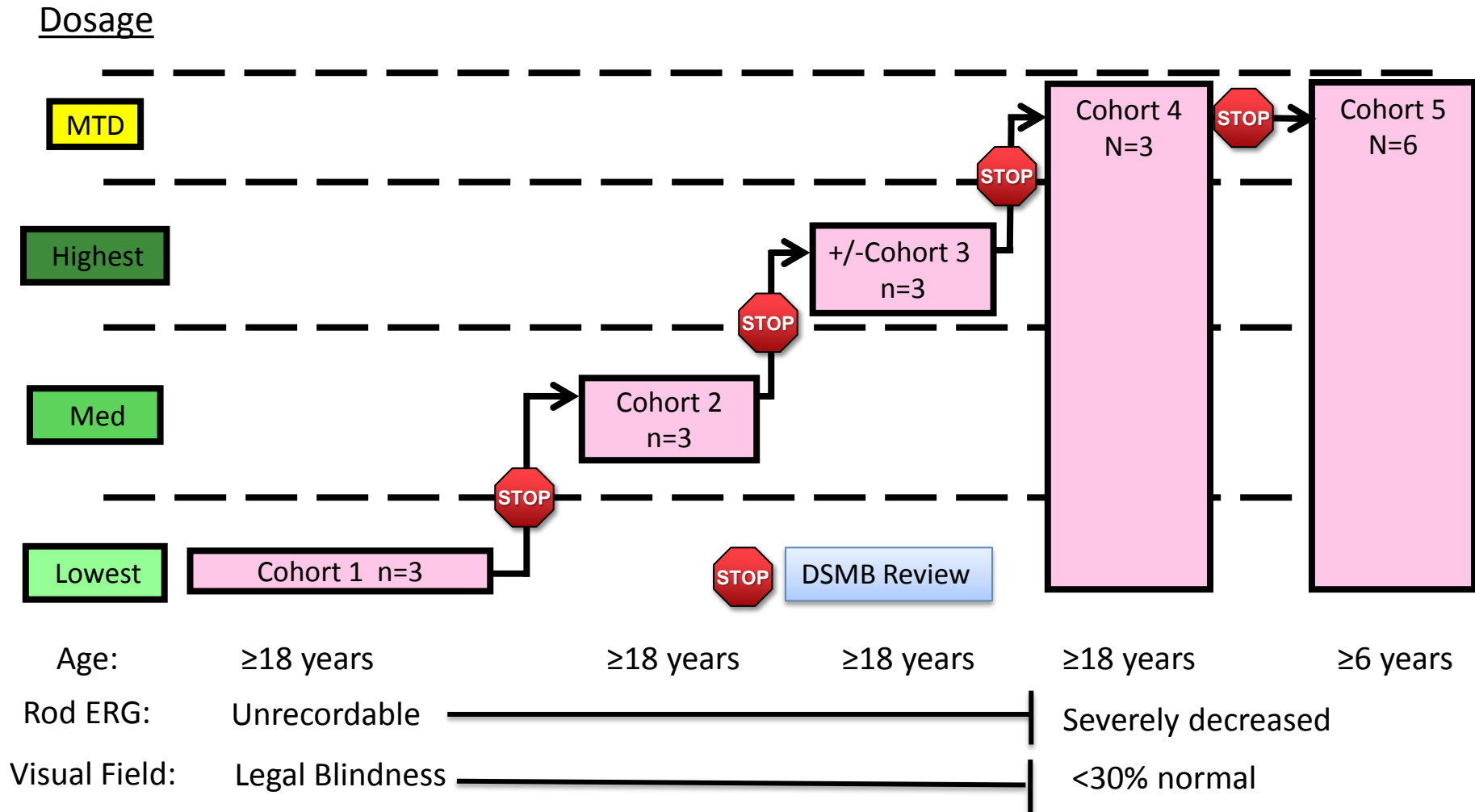
Delivery: Subretinal injection

Ushstattm



- **Based on Equine Infectious Anemia Virus (EIAV)**
 - Wild-type virus causes transient anemia in horses, non-pathogenic in humans
 - Ushstattm vector contains <10% of original viral genome
 - Non-replicating, but does integrate
- **Safety/Transfection Studies in Non-human Primates**
 - Minimal inflammation
 - Low risk for insertional mutagenesis

Ushstat – Study Design



18 Patients Total

Ushstat Inclusion Criteria

****All Patients:** Two confirmed *MYO7A* mutations

Cohorts 1, 2, 3

- ≥ 18 years
- Constriction of Kinetic Visual Field that meets criteria for legal blindness
- No detectable rod ERG

Cohort 4

- ≥ 18 years
- Kinetic Visual Loss - $\geq 30\%$ reduction sensitivity volume
- Evidence of severe rod/cone dysfunction on ERG

Cohort 5

- ≥ 6 years
- Kinetic Visual Loss - $\geq 30\%$ reduction sensitivity volume
- Evidence of severe rod/cone dysfunction on ERG

Ushstat Endpoints

Primary – Safety

- Visual Acuity
- Examination
- Static and Kinetic Visual Field
- OCT
- Laboratory Parameters

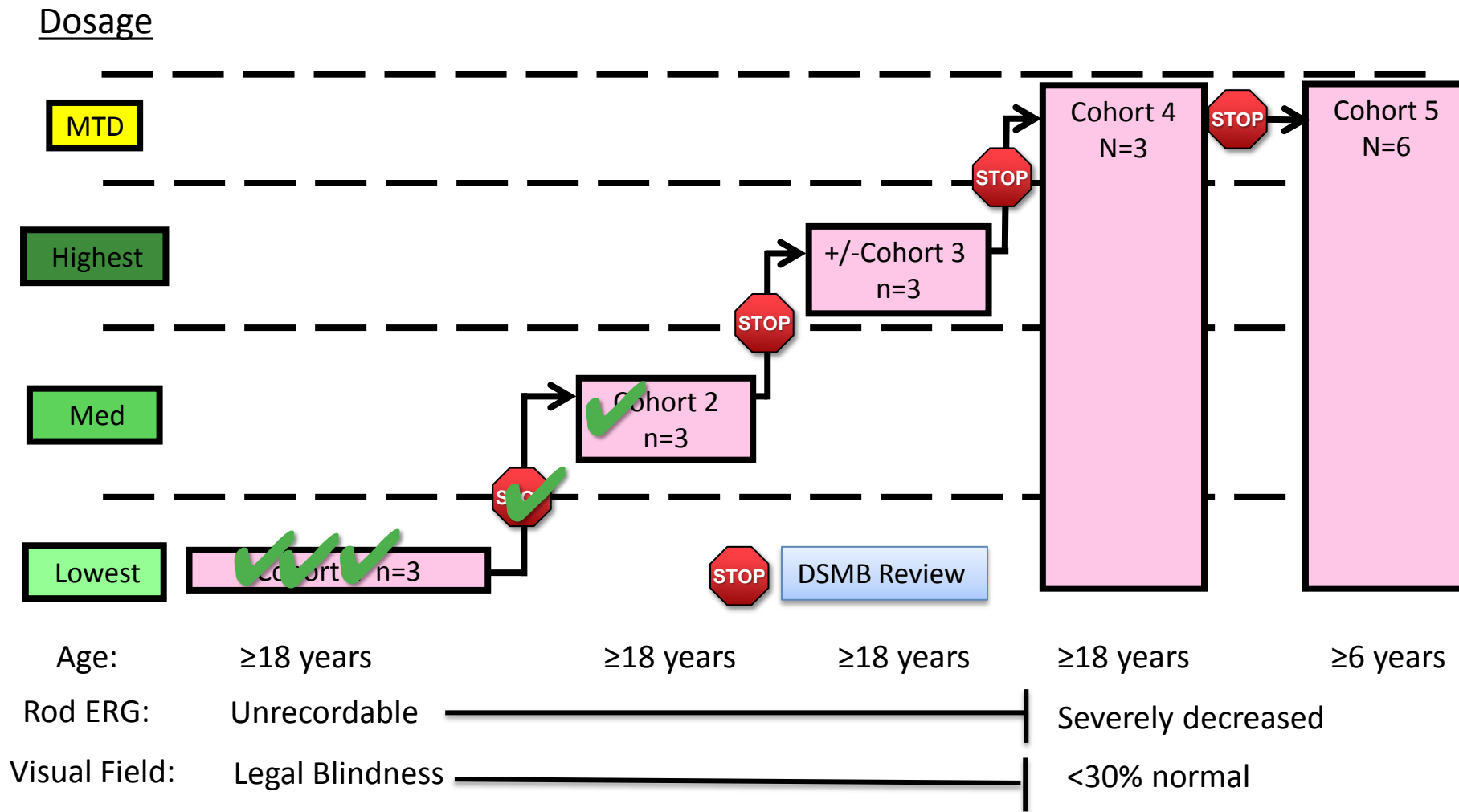
Secondary – Efficacy

- BCVA
- Kinetic and Static Perimetry
- ERG
- OCT
- Adaptive Optics

Recruitment

- First patient dosed April 17, 2012
- Second patient dosed June 28, 2012
- Third patient dosed Oct 4, 2012
- Fourth patient dosed February 16, 2013
- Fifth patient planned, but trial currently on hold

Ushstat – Study Design



- 4 patients treated (3 from Cohort 1, 1 from Cohort 2)
- No serious adverse events

Conclusions

- Multiple gene therapy trials are underway with more planned
- Preliminary safety reports are encouraging

Oregon Retinal Degeneration Center at the Casey Eye Institute



Grant Support

- Hear See Hope
- NIH/NEI 1 K08 EY021186-01
- Foundation Fighting Blindness – CDA
- Research to Prevent Blindness – CDA
- Research to Prevent Blindness
 - Unrestricted grant to CEI
- Foundation Fighting Blindness – Center Grant

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